

### In Silico Docking Analysis Revealed the Potential of Phytochemicals Present in Phyllanthus Amarus and Andrographis Paniculata, Used in Ayurveda Medicine in Inhibiting SARS-CoV-2

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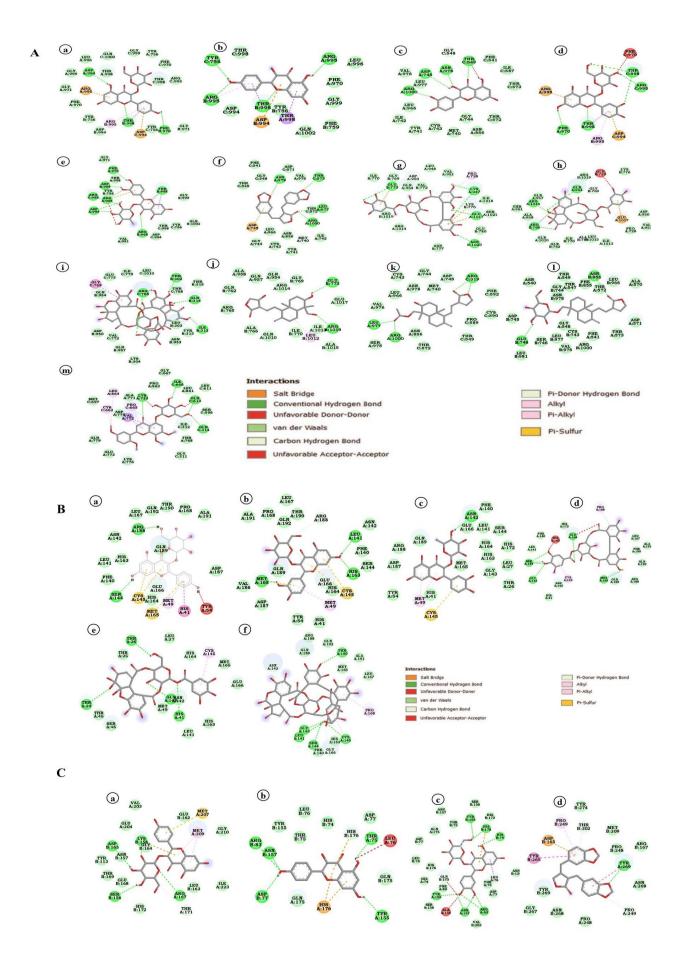
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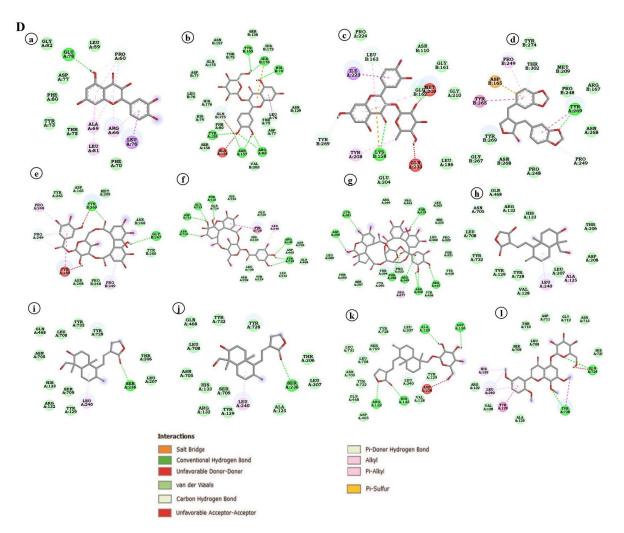
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No therapeutics and vaccines are available against SARS-CoV-2 at present. In the current study we have made an attempt to provide preliminary evidences for interaction of 35 phytochemicals from two plants (Phyllanthus amarus and Andrographis paniculata used in Ayurveda) with SARS-CoV-2 proteins (S protein, 3CLpro, PLpro and RdRp) through in silico docking analysis. The docking was performed with the aid of AutoDock Vina and ADME and other pharmacokinetic properties were predicted using SWISSADME and admetSAR

#### File list (2)

Supplementary figure.pdf (738.72 KiB)	view on ChemRxiv • download file
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**Supplementary figure 1: A,** 2D visualization of interaction between SARS-CoV-2 proteins with phytochemicals from *P. amarus* and *A. paniculata* showing more dock score than remdesivir

**A;** S protein (6VSB) with a) astragalin, b) kaempferol c) quercetin d) quercetin-3-O-glucoside e) quercitrin f) hinokinin g) corilagin h) furosin i) geraniin j) isoandrographolide k) 19-O-acetyl-14-deoxy-11,12-didehydroandrographolide l) neoandrographolide m) 5-hydroxy-7,8,2',5'-tetramethoxy flavone

**B**; 3CLpro (6LU7) with a) astragalin, b) quercetin-3-O-glucoside c) quercitrin, d) corilagin e) furosin f) geraniin

C; PLpro(4OVZ) with a) astragalin, b) kaempferol c) quercetin-3-O-glucoside d) hinokinin

**D;** RdRp (6NUS) with a) quercetin b) quercetin-3-O-glucoside c) quercitrin d) hinokinin e) corilagin f) furosin g) geraniin, h) isoandrographolide i) 14-deoxy-14,15-didehydroandrographolide j) 14-Deoxyandrographolide k) neoandrographolide l) 5-hydroxy-7,8,2',5'-tetramethoxy flavone

# In silico docking analysis revealed the potential of phytochemicals present in *Phyllanthus amarus* and *Andrographis paniculata*, used in Ayurveda medicine in inhibiting SARS-CoV-2

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#### **Abstract**

The Severe acute respiratory syndrome coronavirus-2 (SARS-CoV-2) has resulted in outbreak of global pandemic, fatal pneumonia in human referred as Coronavirus Disease-2019 (Covid-19). No therapeutics and vaccines are available against SARS-CoV-2 at present, which made it difficult to contain this disease worldwide. Drug discovery and vaccine development against this pandemic virus was put in fast pace across the world. Ayurveda, the age old practice of treating human ailments in India can also be considered in this endeavour, if supported by scientific evidences required for regulatory authorities to use against SARS-CoV-2. With this backdrop, attempt was made to provide preliminary evidences for interaction of 35 phytochemicals from two plants (Phyllanthus amarus and Andrographis paniculata used in Ayurveda) with SARS-CoV-2 proteins (S protein, 3CLpro, PLpro and RdRp) through in silico docking analysis. The nucleotide analogue remdesivir, being used in treatment of SARS-CoV-2 was used as a positive control. The results revealed that, 18 phytochemicals from P. amarus and 14 phytochemicals from A. paniculata shown binding energy affinity/dock score (< -6.0 kcal/mol). Fourteen phytochemicals from A. paniculata also shown binding affinity (up to -9.10 kcal/mol). Further, flavonoids (astragalin,

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kaempferol, quercetin, quercetin-3-O-glucoside, quercetin) and tannins (corilagin, furosin, geraniin) present in *P. amarus* have more binding affinity (up to -10.60 kcal/mol) than remdesivir (up to -9.50 kcal/mol). The pharmacokinetic predictions suggest that compounds from the two plants species studied in the current study are found to be non-carcinogenic, water soluble and biologically safe. The phytochemicals present in the extracts of *P. amarus* and *A. paniculata* might may have synergistic effect with action on multiple target sites of SARS-CoV-2. The information generated here might serve as preliminary evidence for anti SARS-CoV-2 activity of phytochemicals present from *P. amarus* and *A. paniculata* and the potential of Ayurveda medicine in combating the virus.

**Keywords**: Coronavirus, COVID-19, Molecular docking, Natural Ligands, *Andrographis* paniculata and *Phyllanthus amarus* 

#### Introduction

The outbreak of highly transmissible fatal pneumonia in human referred as Coronavirus Disease-2019 (Covid-19) has spread globally (215 countries and territories) with over 16.81 M confirmed cases and over 6,62,095 deaths worldwide as on 31st July 2020 (https://www.worldometers.info/coronavirus). Whereas, in India, 15,83,792 people with 34,968 deaths (https://www.worldometers.info/coronavirus). According to the International Committee on Taxonomy of Viruses (ICTV), the novel zoonotic pathogenic virus associated with COVID-19 is Severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) and the virus species is Severe acute respiratory syndrome-related coronavirus (Gorbalenya et al., 2019). The break out of SARS-CoV-2 has been first reported during December 2019 in Wuhan, Hubei Province, China with a fatality rate of 4 percent (Liu, et al., 2020; Wu et al., 2020; Zhu et al., 2020). Sore throat, dry cough, fever, chills, repeated shaking with chills, muscle pain, headache, loss of taste or smell and respiratory distress are major symptoms induced by SARS-CoV-2 (Cheng et al., 2007). In most severe cases, the clinical presentation of SARS-CoV-2 infection includes pneumonia that progresses to multi-organ failure and even death (Zaki et al., 2012; Machhi et al., 2020). Covid-19 has become global pandemic and created the wide spread fear of death everywhere in the world resulting lockdown in human society (Mandal, 2020). The increasing pandemic potential of SARS-CoV-2 is due to its high human to human transmissible efficiency which makes it difficult to contain (Wu et al., 2020; Zhao, et al., 2020).

The SARS-CoV-2 is a β-coronavirus, belongs to family *Coronaviridae* of order Nidovirales (Zhu et al., 2019). It has enveloped positive sense single stranded RNA genome with 33 kilo base (Kb) encoding for 14 open reading frames (ORFs) (Fehr and Perlman, 2015; Guo et al. 2020). The genome of SARS-CoV-2 lacks the hemagglutinin-esterase gene. However, it comprises two flanking untranslated regions (UTRs) at 5' end and 3' with 265 and 358 nucleotides, respectively. The first ORF comprises approximately 67 percent of the genome and encodes 16 non-structural proteins (nsps), which were processed by proteolytic cleavage, whereas the remaining 13 ORFs encode for structural proteins and protein accessory (Boopathi et al., 2020; Chan et al., 2020). The nsps mainly include helicase, RNA dependent RNA polymerase (RdRp), 3-Chymotrypsin-like protease (3CLpro), Papain-like protease (PLpro) and others likely to be involved in the transcription and replication of SARS-CoV-2 (Simmons et al., 2005; Boopathi et al., 2020). Four major structural proteins are spike surface glycoprotein (S protein), membrane protein (M), nucleocapsid protein (N), envelope (E). M protein possess N-terminal glycosylated ectodomain at the N-terminal end that consists of three transmembrane domains (TM) and a long C-terminal CT domain (Boopathi et al., 2020; Chan et al., 2020). The M and E proteins are required for virus morphogenesis, assembly, and budding, whereas, S protein is a fusion viral protein comprising two subunits S1 and S2. The S1 subunit consists of receptor-binding domain (RBD), N-terminal domain (NTD) and signal peptide. The S2 subunit comprises two heptad repeat regions known as HR-N and HR-C, which form the coiled-coil structures surrounded by the protein ectodomain (Walls et al., 2020). A furin cleavage site (PRRARS'V) is present in S protein at the interface between S1 and S2 subunits that will be processed during the biogenesis (Coutard et al., 2020). SARS-CoV-2 S protein facilitate the entry of viral particles into the host cell after binding with the host Angiotensin-converting enzyme 2 (ACE-2) (Tortorici and Veesler, 2019) and also determines the host range and cellular tropism (Simmons et al., 2005). Entry of SARS-CoV-2 into host cells is mediated by the cleavage of viral spike protein by host proteases viz., Serine proteases (TMPRSS2) or Cysteine proteases (Cathepsin B/L) (Boopathi et al., 2020; Yan et al., 2020). The ACE2 and TMPRSS2 proteins are prime targets of the coronavirus (Matsuyama et al., 2010; Glowacka et al., 2011; Zhao et al., 2020).

The most direct and simplest strategy to combat SARS-CoV-2 would be to neutralize the virus from entering cells by targeting the host ACE2 protein, since the chances of mutation in this will be very less (Walker and Burton, 2018; Karakus *et al.*, 2020). Proteases (3CLpro and PLpro) translated from viral RNA are essential for polyprotein processing of

SARS-CoV-2 and involved in replication and transcription of SARS-CoV-2 can serve as potent therapeutic targets (Prussia *et al.*, 2011; Sheahan *et al.*, 2020). The SARS-CoV-2 proteases act on polyproteins (PP1a and PP1ab) to release the nsps which are encoded by open reading frame (ORF1a/b) of virus (Sheahan *et al.*, 2020). Inhibiting virus entry into the cell and its replication combined with modulating the immune system could be a potential target for drug therapy (Sanders *et al.*, 2020). Currently, there are no available vaccines or specific medicines for the treatment of COVID-19.

All the countries and pharma giants in the world are working to come out with either vaccines or drugs which will be helpful in containing this global pandemic. In India, attempts were made to address this problem through Ayurveda and Siddha practices as well apart from development of drugs and vaccines, which were still widely used among the Indian population to combat diseases caused by viral infections such as jaundice, malaria, cholera etc., (Gomathi et al., 2020). Ayurveda is the age old practice of treating human ailments in India and provided remedies against many medical conditions. Medicinal herbs from India are promising in treating various illnesses (Gomathi et al., 2020). Further, identification of phytochemicals from medicinal plants can be used as drugs for viral infection targeting viral receptors (Chang and Woo, 2003; Keyaerts et al., 2007), viral integration (Kim et al., 2010), reverse transcription, viral replication and viral protein translation (Mansouri et al., 2009).

from Andrographis paniculata Wall (Kalmegh) Extracts and *Phyllanthus* amarus Schum. & Thonn were used in Ayurvedic medicinal systems and proven to have numerous medicinal properties (Niranjan et al., 2010; Patel et al., 2011). A. paniculata also known as 'King of bitters' belonging to Acanthaceae family (Pholphana et al., 2013) is widely used to treat fever, diarrhoea, common cold. It consists of wide range of pharmacologically active substances such as andrographolides and its derivatives that have been showed anti-cancer, antimalarial, hepato-protective, antioxidant activities (Niranjan et al., 2010) and reported to have potent antiviral activity against diverse group of viruses belonging to different families (Gupta et al., 2017). It also been implicated in suppression of increased NOD-like receptor protein 3 (NLRP3), caspase-1, and interleukin-1 \beta molecules which are extensively involved in the pathogenesis of SARS-COV and likely SARS-CoV-2 as well (Liu et al., 2020a, Liu et al., 2020b). Phyllanthus amarus Schum. & Thonn. (Family: Euphorbiaceae) is one of a medicinal herb widely distributed in tropical and subtropical countries from Africa to Asia, South America and the West Indies. It is a rich source of secondary metabolites such as alkaloids, flavonoids, hydrolysable tannins, lignins, polyphenols, triterpenes, sterols and volatile oils (Patel et al., 2011). It exhibits an array of ethanopharmacological activities such as anti-inflammatory, hepatoprotective, nephroprotective, anti-amnesia, anti-cancer, diuretic, anti-oxidant, anti-viral, anti-bacterial, anti-hyperglycemic, anti-hypercholesterolemia and so on (Calixto *et al.*, 1998; Patel *et al.*, 2011).

Several Ayurvedic practitioners in India claiming Ayurveda medicine can cure Covid-19, which are not supported by enough scientific evidences. Nevertheless, the role of herbal extracts in addressing the human ailments cannot be ignored totally. Lack of proper experimental evidences might have made regulatory authorities to enforce not to sale the Ayurveda medicines as a curative agent of Covid-19. There are several Ayurveda tablets available in the Indian market claiming to boost immune system in the backdrop of Covid-19 outbreak. Here we discussed on the potential Ayurveda medicine prepared using *A. paniculata* and *P. amarus* against SARS-CoV-2 through the results of *in silico* analysis of interaction between SARS-CoV-2 proteins and secondary metabolites of these plants available in the databases. This provides some preliminary scientific evidence to harness the potential of Ayurveda medicines in combating SARS-CoV-2. However, this needs to be to be validated with experimental evidences required by the regulatory authorities such as large scale clinical trials.

#### **Material and Methods**

#### Sources of ligands and viral receptor proteins

The list of phytochemicals present in *P. amarus* and *A. paniculata* were obtained from previously published scientific literature (Chao and Lin, 2010; Gupta and Vaghela, 2019; Joseph and Raj, 2011; Nisar *et al.*, 2018; Pereira *et al.*, 2016; Tan *et al.*, 2016). A total of 35 compounds were selected for the molecular docking studies (21 from *P. amarus* and 14 from *A. paniculata*) (Table 1). Remdesivir, an antagonist of SARS-CoV-2 RdRp was chosen as a positive control (Yu *et al.*, 2020).

The three-dimensional (3D) conformers of the selected ligands were retrieved from PubChem (https://pubchem.ncbi.nlm.nih.gov) database in SDF and MOL formats. Similarly, the 3D structures of proteins determined using a biophysical technique such as X-ray crystallography or NMR spectroscopy are openly available in protein data bank were retrieved (www.rcsb.org). Four different viral targets were considered for docking studies *viz.*, spike protein (S protein), 3-chymotrypsin-like protease (3CLpro), papain-like protease (PLpro) and RNA dependent RNA polymerase (RdRp). The crystal structure of S protein and 3CLpro of SARS-CoV-2 available in the protein databank were retrieved for the current study. The PDB ID for S protein and 3CLpro are PDB ID: 6VSB, chains A, B, C) (Wrapp *et* 

al., 2020), and PDB ID: 6LU7, chain A) (Jin et al., 2020), respectively. Crystal protein structures of PLpro and RdRp of SARS-CoV-2 were not available in the protein databank at the time of analysis. Hence, the crystal structure developed for the both proteins using the X-ray structure of PLpro of human coronavirus papain-like proteases (PDB ID: 4OVZ, chain A) (Baez-Santos et al., 2014) and the solved RdRp structure (97.08% sequence identity with SARS-CoV-2) of SARS-HCoV (PDB ID: 6NUS, chain A) (Kirchdoerfer et al., 2019) were downloaded for docking studies.

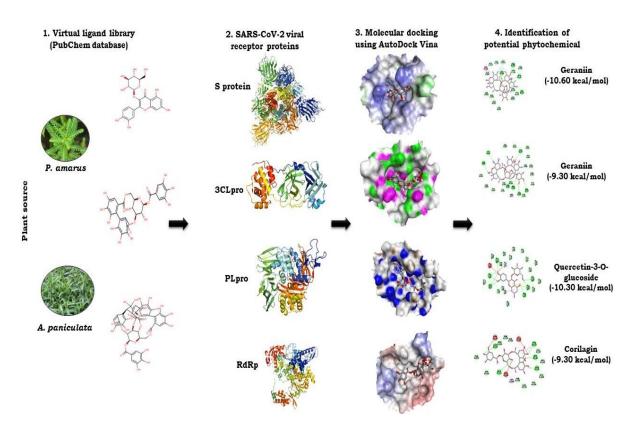
#### Preparation of ligands and target proteins

The retrieved 3D structures of the ligands were optimized using Avogadro version 1.2.0 (Hanwell *et al.*, 2012) with force filed type MMFF94 and converted to PDB format using Open Babel version 3.1.1. (O'Boyle *et al.*, 2011). For further simplification of analysis, the optimized ligands with the lowest energy were loaded to AutoDock-MGL Tools (Morris *et al.*, 2009), the gasteiger charges were added and standard processes were used to obtain the PDBQT files. The downloaded 3D structures were checked for the presence of any improper bonds, side-chain anomalies and missing hydrogens using PyMOL software (DeLano, 2009). All the water molecules, complex molecules, ions and ligands of the proteins were removed in Biovia Discovery Studio 2020 (Biovia, 2020). The optimized PDB structures were uploaded to AutoDock-MGL Tool (Morris *et al.*, 2009), succinctly, polar hydrogens were added and standard processes were used to obtain the PDBQT files.

#### Active site prediction and molecular docking

The active sites of the proteins were determined using the Computed Atlas for Surface Topography of Proteins (CASTp) (http://sts.bioe.uic.edu/castp/index.html?2011) and Biovia Discovery Studio 2020. In the case of 3CLpro protein, the binding site was selected based on the location of co-crystallized inhibitor N3, which was developed using computer-aided drug design and has shown specific inhibition to SARS and MERS with good binding affinity in the binding pockets of crystal structure of 3CLpro (Jin *et al.*, 2020). The optimized ligands and proteins in PDBQT format were subjected to molecular docking with the aid of AutoDock Vina software (Trott and Olson, 2010). AutoDock Vina predicts the interaction between protein and ligand by utilizing its scoring function (binding affinity). In the process of molecular docking, based on predicted active sites, the grid box was set to 60 Å×60 Å×60 Å centered at 225.616, 226.490, 225.337 (XYZ coordinates) for spike protein, grid box 26 Å×26 Å centered at -10.712, 12.411, 68.8312 (XYZ coordinates) for 3CLpro, grid box 60 Å×60 Å centered at -8.611, 38.916, -41.012 (XYZ coordinates) for PLpro and grid box 60 Å×60 Å centered at 138.751, 163.504, 136.636 (XYZ coordinates) for RdRp.

The output file obtained after docking analysis consists of top 9 binding poses with their respective binding affinity in kcal/mol. The ligand binding pose showing highest binding affinity with least root mean square deviation (RMSD) was selected. The protein-ligand interaction in 3D structures was visualized in PyMOL. Similarly, 2D structure was visualized in Biovia Discovery studio 2020. The 3D visualization represents the exact location or the binding pocket of the target protein. Whereas, the 2D structure visualization represents the different bonds formed between the amino acid residues of the viral target protein and the ligand. The workflow of *in silico* molecular docking analysis has been represented in figure 1.



**Figure 1:** The workflow of molecular docking analysis of phytochemicals present in *P. amarus* and *A. paniculata* with four proteins (S protein, 3CLpro, PLpro and RdRp) targets of SARS-CoV-2. The phytochemicals of the two plant species were obtained from PubChem (https://pubchem.ncbi.nlm.nih.gov) database in SDF and MOL formats. The 3D structures of proteins available in protein data bank were retrieved (www.rcsb.org). The detailed protocol is given in the material and methods.

#### **ADME** and other pharmacokinetic properties

The drug-likeness analysis was carried out based on Lipinski's rule of five (Lipinski, 2004). Absorption, Distribution, Metabolism, and Excretion (ADME) are pharmacokinetic properties of a ligand that deals primarily with its absorption, distribution, metabolism and excretion in the human body. The major parameter, Lipinski's rule of five was generated using SWISSADME (Daina *et al.*, 2017) for evaluating the pharmacological significance. The ligands in PDB format were converted to canonical SMILES format and uploaded to SWISSADME prediction (http://www.swissadme.ch). Properties such as solubility (logS), human intestinal absorption (HIA) and carcinogen were obtained from admetSAR (http://lmmd.ecust.edu.cn/admetsar2).

#### **Results and Discussion**

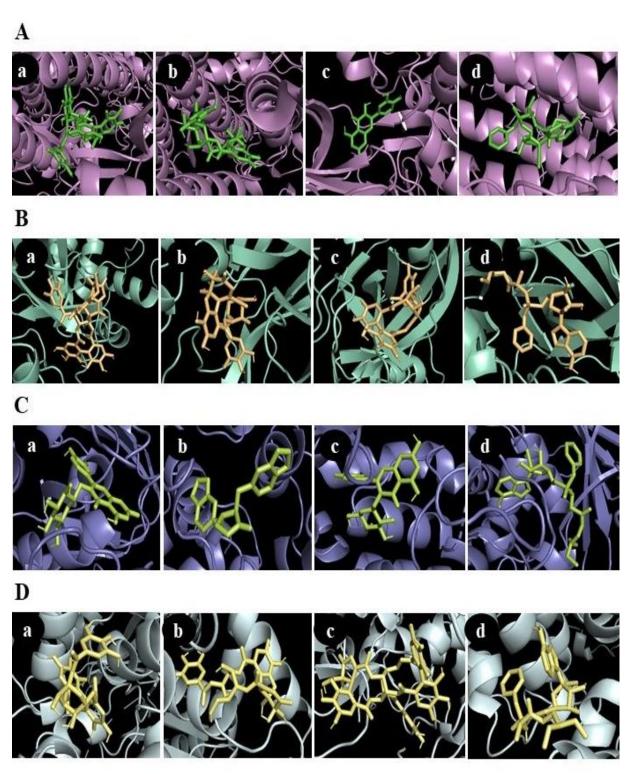
Impact of Covid-19 is having catastrophic effect on human health and life. The scientific communities from the nuke and corner of the world have scrupulously engaged in developing the drug or vaccine against SARS-CoV-2, causal agent of Covid-19. The current study discusses on the potential of phytochemicals present in two medicinal plants used in the Ayurveda, an age old medical practice in India. Plants/herbal extracts part of Ayurveda medicines are known to boost immunity in humans, which is crucial for fighting against microbial enemies (Kumar *et al.*, 2017). Plant extracts from *P. amarus* and *A. paniculata* have been extensively used in Ayurveda for curing various human ailments (Niranjan *et al.*, 2010; Patel *et al.*, 2011). The herbal extracts may have several phytochemicals, few of them might be playing role in neutralizing the pathogens. Binding interaction between a pathogen protein and ligand derived from herbal extracts may result in inhibition of the activity of pathogen protein. Hence, to understand and determine the potency of the plant extracts against cataclysmic virus SARS-CoV-2, about 21 compounds present in *P. amarus* and 14 compounds present in *A. paniculata* were used for molecular docking simulation with four SARS-CoV-2 receptor targets.

Prior to the docking analysis, the ligands were optimized by minimizing the energy with force field type MMFF94, this helps in removing clashes among atoms and develop a stable starting pose of the ligands for binding interaction (Klebe *et al.*, 2006). The docking coupled with a scoring function can be utilized to evaluate a large number of potential drugs *in-silico* (Monika *et al.*, 2010). In the process of molecular docking, generally, the binding affinity lesser than the upper threshold (-6 kcal/mol) is regarded as a *cut off* value for determining good binding affinity between ligand and protein (Shityakov *et al.*, 2014). Of the

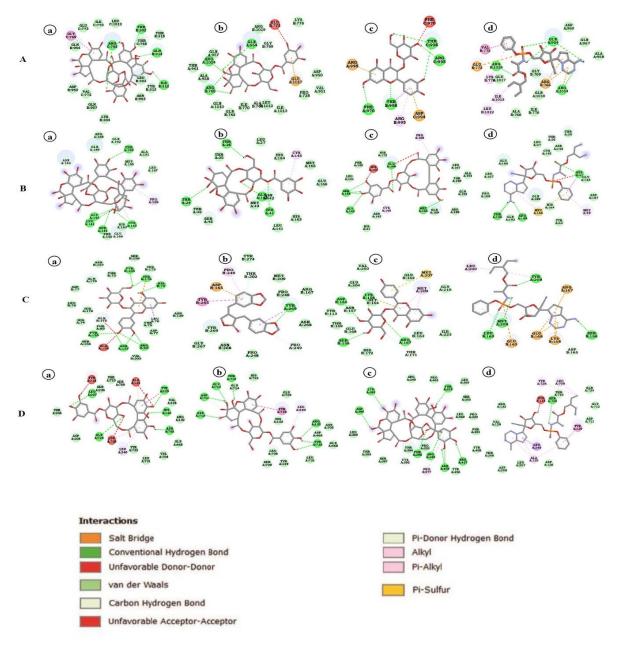
35 compounds screened from two plant species (Table 1), three compounds (cinnamic acid, Phenylalanine and farnesyl farnesol) did not qualify as per the threshold binding affinity to neutralize SARS-CoV-2 proteins.

In the recent clinical trials, ramdesivir, a nucleotide analogue of adenosine triphosphate has been reported to be a potential prodrug against SARS-CoV-2 infection (Gao *et al.*, 2020). It was originally developed for treating viral infection caused by Ebola and Marburg virus but was also found effective against SARS and MERS (Elfiky, 2020). Hence, remdesivir has been used as a positive control in the current study. During virus replication, the RdRp mistakenly incorporates ramdesvir into its genome, which results in stalling of replication (Wang *et al.*, 2020). In our study, remdesivir has shown good interaction with RdRp protein (Table 1) with the binding affinity of -7.6 kcal/mol. Similarly, it has shown a good binding affinity with 3CLpro (-7.90 kcal/mol), S protein (-8.00 kcal/mol) and PLpro (-9.50 kcal/mol) (Table 1) and might be having some role in inhibiting SARS-CoV-2. Many recent computational investigations also showed inhibition activity of remdesivir with various viral receptors of SARS-CoV-2 (Murugan *et al.*, 2020; Elfiky, 2020; Hall and Feng, 2020). This shows that the prodrug might be effective against all the four viral receptors.

The 3D and 2D visualization of top three phytochemicals along with positive control remdesivir based on binding affinity with respective viral receptor protein has been represented in figure 2 and 3, respectively. Hydrogen bond energy is the major contributor in dock score. Hydrogen bond formation between ligand structures and viral receptors is responsible for inhibition of the target protein and it reflects the firmness of bonding between the protein and ligand (Chen *et al.*, 2016). Selected phytochemicals from *P. amarus* and *A. paniculata* used in the current docking studies against SARS-CoV-2 proteins shown very good dock score above the threshold cut off of -6 kcal/mol. Ligand structures and necessary hydrogen bond formation between phytochemical with respective viral receptor has been illustrated in table 2 and supplementary figure 1.



**Figure 2:** 3D visualization of interaction between SARS-CoV-2 proteins with representative phytochemicals. **A:** S protein (6VSB) with a) Geraniin, b) Furosin, c) Quercetin-3-O-glucoside, d) Remdesivir (positive control). **B:** 3CLpro (6LU7) with a) Geraniin, b) Furosin, c) Corilagin, d) Remdesivir (positive control). **C:** PLpro (4OVZ) with a) Quercetin-3-O-glucoside, b) Hinokinin, c) Astragalin, d) Remdesivir (positive control). **D:** RdRp (6NUS) with a) Corilagin, b) Furosin, c) Geraniin, d) Remdesivir (positive control).



**Figure 3:** 2D visualization of interaction between SARS-CoV-2 proteins with representative phytochemicals. **A:** S protein (6VSB) with a) Geraniin, b) Furosin, c) Quercetin-3-O-glucoside, d) Remdesivir (positive control). **B:** 3CLpro (6LU7) with a) Geraniin, b) Furosin, c) Corilagin, d) Remdesivir (positive control). **C:** PLpro (4OVZ) with a) Quercetin-3-O-glucoside, b) Hinokinin, c) Astragalin, d) Remdesivir (positive control). **D:** RdRp (6NUS) with a) Corilagin, b) Furosin, c) Geraniin, d) Remdesivir (positive control).

#### Interaction between spike glycoprotein (S protein) (6VSB) and phytochemicals

S protein of the SARS-CoV-2 latch onto human angiotensin-converting enzyme 2 (ACE2) receptor and undergo structural changes that allow the viral membrane to fuse with the cell membrane of the host (Walls *et al.*, 2020). Despite the high similarity between sequences and structures of SARS and SARS CoV-2, the drugs successfully used to cure

SARS are ineffective in case of this novel virus (Fani *et al.*, 2020). This indicates that more number of potential molecules should be screened to find an effective molecule.

The molecular docking analysis of S protein of the SARS-CoV-2 with 21 selected phytochemicals from P. amarus showed that majority of the compounds bind to S protein with favourable binding energy ranging from -6.10 kcal/mol (for Phyllanthin) to -10.60 kcal/mol (for geraniin) (Table 1). When the binding energy of phytochemicals was investigated, nine compounds viz., geraniin (-10.60 kcal/mol), furosin (-10.20 kcal/mol), astragalin (-9.80 kcal/mol), quercetin-3-O-glucoside (-9.80 kcal/mol), quercitrin (-9.20 kcal/mol), corilagin (-8.80 kcal/mol), hinokinin (-8.40 kcal/mol), kaempferol (-8.30 kcal/mol) and quercetin (-8.10 kcal/mol) exhibited highest binding affinity. Interestingly, the phytochemicals belonging to the flavonoids and tannins of *P. amarus* have shown a very high affinity with the target protein. Molecular dynamic simulation studies also reported that quercetin and kampferol bind with the ACE2-S complex with low binding free energy (Pandey et al., 2020). The antiviral bioactivities of many flavonoids from medicinal plants were reported by several studies (Thayil et al., 2016; Zakaryan et al., 2017; Jo et al., 2020). Among all compounds, geraniin exhibited (-10.60 kcal/mol) highest docking score with S protein. THR302, ILE312, GLN314, ARG765 amino acid residues were involved in forming four hydrogen bonds in the binding pocket of S protein (Table 2, Figure 3)

In binding interaction between phytochemical compounds (14) from *A. paniculata* with S protein of the SARS-CoV-2 the docking score ranged from -7.30 (for andrograpanin) and -9.10 kcal/mol (for isoandrographolide) (Table 1). Of the 14 compounds, five compounds; isoandrographolide (-9.10 kcal/mol), 19-O-acetyl-14-deoxy-11, 12-didehydroandrographolide (-8.40 kcal/mol), neoandrographolide (-8.20 kcal/mol) and 5-hydroxy-7, 8, 2', 5'-tetramethoxy flavone (-8.20 kcal/mol) shown better binding affinity as compared to remdesivir (-8.00 kcal/mol). Isoandrographolide has properly positioned into the binding pocket constructed by hydrogen bonding with GLU773 and ARG1019 amino acids of S protein exhibited highest (-9.10 kcal/mol) binding affinity (Table 2, Figure 2). Totally, 13 compounds from *P. amarus* and *A. paniculata* have shown better inhibitory activity against S protein.

#### Interaction between 3-chymotrypsin like protease (3CLpro) (6LU7) and phytochemicals

The SARS-CoV-2 protease enzyme, 3CLpro, involved in C-terminal cleavage of virus polyprotein is a potential target for drug discovery to SARS-CoV-2, since it controls replication (Qmar *et al.*, 2020). Hence, this potential target was evaluated with the selected 35 compounds from *P. amarus* and *A. paniculata*. Before docking simulation, the target site for

viral receptor protein was selected based on the location of co-crystallized inhibitor N3 as this target site has been considered to be the most commending site for ligand interaction (Murugan *et al.*, 2020).

The binding affinity of compounds to 3CLpro ranged from -6.10 kcal/mol (for andrograpaninn) to -9.30 kcal/mol (for geraniin) (Table 1). Among 21 phytochemicals derived from P. amarus, six compounds viz., geraniin (-9.30 kcal/mol), furosin (-8.70 kcal/mol), corilagin (-8.70 kcal/mol), quercitrin (-8.60 kcal/mol), astralagin (-8.40 kcal/mol) and quercetin-3-O-glucoside (-8.20 kcal/mol) have shown better binding affinity as compared to remdesivir (-7.90 kcal/mol) (Table 1). The compounds belong to tannins (geraniin, furosin and corilagin) and flavonoids (quercitrin, astralagin, and quercetin-3-O-glucoside) are found to have good inhibitory action on 3CLpro. Several reports have been shown antiviral effect of flavonoids (Gescher et al., 2010; Chen et al., 2012). Recently, Adem et al., (2020) evaluated 80 flavonoids compounds present in different medicinal plants against 3CLpro of SARS-CoV-2. Among them, hesperidin, rutin, diosmin, apiin, diacetylcurcumin, (E)-1-(2-Hydroxy-4-methoxyphenyl)-3-[3-[(E)-3-(2-hydroxy-4-methoxyphenyl)-3-oxoprop-1-enyl] phenyl] prop-2-en-1-one, and beta. beta'-(4-Methoxy-1,3- phenylene) bis(2'-hydroxy-4',6'dimethoxyacrylophenone have shown effective binding affinity with SARS-CoV-2 3Clpro. The flavonoids from the two plant species used in the present study have higher binding affinity to protease than flavonoids reported by Adem et al. (2020). Geraniin, with the highest binding affinity of -9.30 kcal/mol has formed five hydrogen bonds with the amino acid residues, LEU141, GLY143, SER144, CYS145, THR190 of 3CLpro (Table 2, Figure 3).

The docking score for compounds present in *A. paniculata* ranged from -6.10 kcal/mol (for andrograpanin) to -7.80 kcal/mol (for neoandrographolide) (Table 1) which is on par with the remdesivir showing the score of -7.90 kcal/mol. During replication of the viral genome, the 3CLpro cleaves the C-terminal of polypeptide protein at 11 distinct sites to generate various nsps (Qmar *et al.*, 2020). Hence, the phytochemicals present in the extracts of *P. amarus* might hamper the activity of the protease enzyme there by inhibiting the process of virus multiplication.

#### Interaction between papain like protease (PLpro) (4OVZ) and phytochemicals

The SARS-CoV-2 another protease enzyme, PLpro known to be involved in cleaving of N-terminal of viral polyprotein which is also an important antiviral drug target. It also plays a crucial role in its replication along with 3CLpro (Baez-Santos, 2015). The 3D structure of PLpro used in the docking simulation is the x-ray structure of SARS-CoV, which has the highest similarity with the PLpro of SARS-CoV-2 (Zhou *et al.*, 2020).

The binding energy of phytochemicals present in *P. amarus* ranged from -6.30 kcal/mol (for cinnamic acid) to -10.30 kcal/mol (for quercetin-3-O-glucoside) (Table 1). Four compounds; quercetin-3-O-glucoside (-10.30 kcal/mol), hinokinin (-9.80 kcal/mol), astragalin (-9.70 kcal/mol) and kaempferol (-9.60 kcal/mol) from *P. amarus* have exhibited better binding energies compared to remdesivir (-9.50 kcal/mol). Apart from this, the binding affinities of furosin (-9.20 kcal/mol) and geraniin (-9.20 kcal/mol) are reasonably well with that of the positive control, remdesivir. Quercetin-3-O-glucoside has the maximum dock score of -10.30 kcal/mol and shown its binding to HIS74, ARG83, TYR155, ASN157, HIS176 amino acid residues of viral protein at the binding pocket of PLpro with eight hydrogen bonds (Table 2, Figure 3)

The binding affinity of compounds from *A. paniculata* with PLpro ranged from -6.80 kcal/mol (for andrograpanin) to -9.00 kcal/mol (for 5-Hydroxy-7,8-dimethoxyflavone). Even though binding affinities of all the 15 compounds falls between -6.00 kcal/mol and less than the dock score of positive control remdesivir (-9.5 kcal/mol), they also might have inhibitory effects on the activity of PLpro. The results suggest that, among the promising phytochemicals from *P. amarus* and *A. paniculata* docked with PLpro, flavonoids (quercetin-3-O-glucoside and astragalin) and phenyl propanoid (hinokinin) have exhibited better binding affinity. The SARS-CoV-2 PLpro is essential virus replication (Baez-Santos *et al.*, 2015) and any compound having good binding affinity with inhibitory effects will be of immense potential in containing this virus. The flavonoids and phenyl propanoids showing good affinity binding abilities might serve as lead molecules for the discovery of novel drugs to SARS-CoV-2.

## Interaction between RNA dependent RNA polymerase (RdRp) (6NUS) and phytochemicals

The RdRp plays a major role in the replication and transcription of SARS-CoV-2 genome (Aftab *et al.*, 2020). The RdRp used in the current study is the solved structure of SARS-CoV having 97.08 percent sequence identity with the RdRp of SARS-CoV-2 (Kirchdoeffer and Ward, 2020).

Among 21 compounds from *P. amarus* screened against RdRp, epibubbialine has shown lowest dock score of -6.30 kcal/mol (below -6.0 kcal/mol is considering as having good binding affinity) and corilagin has shown highest dock score of -9.30 kcal/mol (Table 1). Seven compounds; corilagin (-9.30 kcal/mol), furosin (-9.10 kcal/mol), geraniin (-9.10 kcal/mol), quercetin (-8.60 kcal/mol), hinokinin (-8.20 kcal/mol), quercetrin (-8.10 kcal/mol) and quercetin-3-O-glucoside (-7.70 kcal/mol) from *P. amarus* exhibited better dock score

compared to the RdRp antagonist remdesivir (-7.60 kcal/mol). Corilagin with high binding affinity of -9.30 kcal/mol has interacted with active site residues, TYR129, HIS133, LEU207, ASN705, GLN724 of RdRp of virus through five hydrogen bonds (Table 2, Figure 3)

Five compounds, neoandrographolide (-8.50 kcal/mol), isoandrographolide (-8.30 kcal/mol), 14-deoxy-14,15-didehydroandrographolide (-7.90 kcal/mol), 14-Deoxyandrographolide (-7.80 kcal/mol) and 5-hydroxy-7, 8, 2',5'-tetramethoxy flavone (-7.80 kcal/mol) from *A. paniculata* have shown better binding energy affinity compared to remdesivir (-7.60 kcal/mol) (Table 1). Among these, neoandrographolide exhibited higher dock score of -8.50 kcal/mol and interacted with ALA125, ASP126, ARG132, HIS133 amino acid residues of RdRp through four hydrogen bonds (Table 2, Figure 3).

In general, the flavonoids (astragalin, kaempferol, quercetin, quercetin-3-O-glucoside and querectrin), a phenyl propanoid (hinkonin) and tannins (coriligan, furosin and gerannin) from P. amarus and a bioactive compound (isoandrographolide), diterpenes (14-deoxy-14,15didehydroandrographolide, 14-Deoxyandrographolide, 19-O-acetyl-14-deoxy-11,12 didehydroandrographolide, Neoandrographolide) and a flavonoid (5-hydroxy-7,8,2',5'tetramethoxy flavone) from A. paniculata have shown good binding affinities with all the four SARS-CoV-2 target proteins. The natural compounds obtained from the medicinal plants have been used in case of two previous coronavirus outbreaks of MERS-CoV and SARS-CoV (Li et al., 2005; Lin et al., 2005). Similarly, strong interaction of tannins (polyphenols) from green tea, epigallocatechin gallate, epicatechingallate and gallocatechin-3-gallate with viral receptor (3CLpro) of SARS-Cov-2 was shown (Ghosh et al., 2020). The flavonoids and tannins interaction with proteins of SARS-CoV-2 in vitro may provide more insights into exploiting these compounds as potential SARS-CoV-2 inhibiting compounds and lead to novel drug development. Further, among all the phytochemicals, the geraniin has shown consistent interaction and very good binding affinity with all the viral protein receptors. The geraniin, extracted from P. amarus has shown to inhibit HIV-1 replication in HeLa CD4+ cells with 50% effective concentration (EC50) values ranging from 0.9 to 7.6 μg/ml (Notka et al., 2004). This suggests that geraniin with multiple target binding ability in docking analysis with high dock score than remdesvir can be a potential candidate to harness as anti SARS-CoV-2 drug. However, this needs in vitro evaluation to authenticate the prediction studies.

**Table 1:** Dock score of interaction between phytochemicals from *Phyllanthus amarus* and *Andrographis paniculata* with SARS-CoV-2 proteins; S protein (PDB ID:6VSB), 3 chymotrypsin like protease (3CLpro) (PDB ID:6LU7), papain like protease (PLpro) (PDB ID:4OVZ) and RNA dependent RNA polymerase (RdRp) (PDB ID:6NUS).

  -			S protein	3CLpro	PLpro	RdRp	
Group Sl. No		Compound	Binding affinity (kcal/mol)	Binding affinity (kcal/mol)	Binding affinity (kcal/mol)	Binding affinity (kcal/m ol)	
Positive control	1	Remdesivir	-8.00	-7.90	-9.50	-7.60	
Phytochemicals fr	rom <i>P</i> .						
	1	Epibubbialine	-6.80	-5.90	-7.50	-6.30	
Alkaloids	2	Isobubbialine	-7.40	-6.00	-7.70	-6.40	
Aikaivius	3	nor-securinine	-6.80	-5.70	-7.30	-6.80	
	4	Securinine	-7.10	-5.90	-7.70	-7.30	
	5	Astragalin*	-9.80	-8.40	-9.70	-7.60	
	6	Kaempferol*	-8.30	-7.70	-9.60	-7.00	
Flavanoids	7	Quercetin*	-8.10	-7.50	-8.60	-8.60	
	8	Quercetin-3-O-glucoside*	-9.80	-8.20	-10.30	-7.70	
	9	Quercitrin*	-9.20	-8.60	-8.80	-8.10	
	10	Cinnamic acid	-5.80	-4.70	-6.30	-5.60	
	11	Hinokinin*	-8.40	-7.40	-9.80	-8.20	
Phenyl	12	Niranthin	-6.30	-6.30	-7.60	-6.10	
propanoid	13	Nirtetralin	-7.40	-6.60	-6.70	-7.00	
 	14	Phenylalanine	-5.50	-4.80	-6.00	-5.60	
	15	Phyllanthin	-6.10	-5.70	-7.60	-6.40	
	16	Corilagin*	-8.80	-8.70	-8.50	-9.30	
Tr	17	Furosin*	-10.20	-8.70	-9.20	-9.10	
Tannins	18	Geraniin*	-10.60	-9.30	-9.20	-9.10	
	19	Melatonin	-6.60	-5.80	-7.00	-5.50	
T.::4	20	farnesyl farnesol	-5.20	-4.50	-5.80	-4.70	
Triterprnes	21	henazine -6.70		-5.80	-7.50	-7.00	
Phytochemicals fr	om A.	paniculata					
Bioactive	1	Andrograpanin	-7.30	-6.10	-6.80	-7.20	
	2	Andrographolide	-7.90	-6.50	-8.00	-7.30	
compounds	3	Isoandrographolide* -9.10 -7.0		-7.00	-9.30	-8.30	
	4	14-acetylandrographolide	-7.60	-6.70	-8.40	-7.00	
	5	14-deoxy-11,12- didehydroandrographolide	-7.70	-7.60	-7.30	-7.10	
Diterpenes	6	14-deoxy-14,15- didehydroandrographolide -8.00 -6.80			-7.80	-7.90	
Diter penes	7	14-Deoxyandrographolide	-8.00	-7.30	-8.30	-7.80	
	8	19-O-acetyl-14-deoxy-11,12- didehydroandrographolide	-8.40	-6.80	-8.80	7.10	
	9	Deoxyandrographolide	-7.80	-7.30	-7.00	7.50	
	10	Neoandrographolide	-8.20	-7.80	-7.80	-8.50	
	11	5-hydroxy-7,8,2',3'- tetramethoxyflavone,	-7.60	-6.90	-8.40	-7.00	
Flavanoids	12	5-hydroxy-7,8,2',5'-tetramethoxy flavone	-8.20	-7.60	-8.60	-7.80	
	13 14	5-hydroxy-7,8-dimethoxyflavanone 5-Hydroxy-7,8-dimethoxyflavone	-7.80 -7.90	-7.20 -7.50	-8.90 -9.00	-7.30 -7.50	

Note: The docking score of respective phytochemicals higher than the positive control (remdesivir) has been denoted in **bold.** 

<sup>&</sup>quot;\*" indicates the phytochemicals showing better binding affinity with all the viral proteins.

**Table 2:** Number of hydrogen bonds formed during the interaction between phytochemical structures with SARS-CoV-2 Proteins; S protein (PDB ID:6VSB), 3 chymotrypsin like protease (3CLpro) (PDB ID:6LU7), papain like protease (PLpro) (PDB ID:4OVZ) and RNA dependent RNA polymerase (RdRp) (PDB ID:6NUS). The interactions showing more binding affinity than remdesivir and SARS-CoV-2 are presented here.

Remdesivir   121304016	receptor ing with
1 121304016	9
121304016	
2   Astragalin   5282102     3   S protein   PHE970, ASP994, THR998   2   3CLpro   SER144, ARG188   2   3CLpro   SER156, LYS158, ASP165, ARG915, ARG915, THR756   3   S protein   TYR756, ARG995, THR756   4   S protein   TYR756, ARG995, THR756   5280863   C <sub>15</sub> H <sub>10</sub> O <sub>6</sub>   5   PLpro   THR75, ASP77, ARG83, ASN1   4   S protein   THR549, ASP745, ASN978, Algorithm   S protein   THR549, ASP745, ASN978, Algorithm   S protein   S protein   S protein   S protein   PHE970, ARG995, THR998   S protein   PHE970, ARG995, THR998   S protein   S protein   PHE970, ARG995, THR998   S protein   S protein   PHE970, ARG995, THR998   S protein   PHE970,	ET209
2	
2 5282102 4 PLpro SER156, LYS158, ASP165, AR  3 Kaempferol 5280863 5 PLpro THR75, ASP77, ARG83, ASN1  4 Quercetin 5280343 4 RdRp GLU350, PRO323, ASN628, V  5 Sprotein PHE970, ARG995, THR998  4 RdRp GLU350, PRO323, ASN628, V  5 Sprotein PHE970, ARG995, THR998  4 RdRp GLU350, PRO323, ASN628, V  5 Sprotein PHE970, ARG995, THR998  4 RdRp GLU350, PRO323, ASN628, V  5 Sprotein PHE970, ARG995, THR998  6 PLpro HIS74, ARG83, TYR155, ASN157, HIS176  7 RdRp ALA125, HIS133	
SZ82102   4	
3	RG167
5 5280863 5 PLpro THR75, ASP77, ARG83, ASN1  Quercetin 5280343 4 S protein THR549, ASP745, ASN978, AI  Quercetin 5280343 5 S protein PHE970, ARG995, THR998  quercetin-3-O-glucoside 5280804 8 PLpro HIS74, ARG83, TYR155, ASN157, HIS176  RdRp GLU350, PRO323, ASN628, V  B PLpro HIS74, ARG83, TYR155, ASN157, HIS176  RdRp ALA125, HIS133	
4 Quercetin 5280343 4 RdRp GLU350, PRO323, ASN628, V  5 Sprotein PHE970, ARG995, THR998  3 3CLpro LEU141, HIS163, MET 165  glucoside 5280804 8 PLpro HIS74, ARG83, TYR155, ASN157, HIS176  C <sub>21</sub> H <sub>20</sub> O <sub>12</sub> 3 RdRp ALA125, HIS133	157
4 S280343 4 RdRp GLU350, PRO323, ASN628, V  quercetin-3-O- glucoside 5280804 5 S protein PHE970, ARG995, THR998  8 PLpro HIS74, ARG83, TYR155,ASN157,HIS176  RdRp GLU350, PRO323, ASN628, V  9 PHE970, ARG995, THR998  1 PHE970, ARG995, THR998  3 RdRp ALA125, HIS133	RG1000
5     quercetin-3-O-glucoside 5280804     3     3CLpro     LEU141, HIS163, MET 165       8     PLpro     HIS74, ARG83, TYR155,ASN157,HIS176       C <sub>21</sub> H <sub>20</sub> O <sub>12</sub> 3     RdRp     ALA125, HIS133	AL675
5 glucoside 5280804 8 PLpro HIS74, ARG83, TYR155,ASN157,HIS176 C <sub>21</sub> H <sub>20</sub> O <sub>12</sub> 3 RdRp ALA125, HIS133	
5 glucoside 5280804 8 PLpro HIS74, ARG83, TYR155,ASN157,HIS176 C <sub>21</sub> H <sub>20</sub> O <sub>12</sub> 3 RdRp ALA125, HIS133	
C211120O12	
	HR998
6 Quercitrin 1 3CLpro ASN142	
5280459	
4 S protein THR573, LEU977, ASN978, AC	GR1000
7 Hinokinin 0 RdRp	
' 442879 1 PLpro TYR269	
5 S protein GLU773, LYS947, GLU1017, A	ASN1023
Corilagin 73568 6 3CLpro GLY143, SER144, MET165, G	LU166
5 6NUS TYR129, HIS133, LEU207, AS GLN724	5N705,
4 S protein ARG765, GLN954, AGG1014	
Furosin 4 3CLpro THR24, THR26, ASN142, GLY	7143
9 10416810 6 PLpro ASP165, PRO249, GLN270, TY	YR274
C <sub>27</sub> H <sub>22</sub> O <sub>19</sub> 7 RdRp ARG132, THR710, ASP711, G ASN713, TYR732	LY712,
4 S protein THR302, ILE312, GLN314, AF	RG765
10 Geraniin 3001497 5 3CLpro LEU141, GLY143, SER144, CY	YS145,
2 PLpro GLU251, TYR269	

		C <sub>41</sub> H <sub>28</sub> O <sub>27</sub>	8	RdRp	THR319, ASP390, LYS391, ARG349, PHE396, ARG457, ASN459
			2	S protein	GLU773, ARG1019
	Isoandrographo	CH.	1	PLpro	TYR155
11	lide 343585	C <sub>20</sub> H <sub>30</sub> O <sub>5</sub>	0	RdRp	
12	14-deoxy- 14,15- didehydroandr ographolide 6473762	C <sub>20</sub> H <sub>28</sub> O <sub>4</sub>	1	RdRp	SER236
13	14- Deoxyandrogra pholide 11624161	C <sub>20</sub> H <sub>30</sub> O <sub>4</sub>	1	RdRp	SER236
14	19-O-Acetyl- 14-deoxy- 11,12- didehydroandr ographolide 46179874	C <sub>22</sub> H <sub>30</sub> O <sub>5</sub>	4	S protein	ARG319, LEU977, ARG1000
	Neoandrograph	C <sub>26</sub> H <sub>40</sub> O <sub>8</sub>	2	S protein	GLU748, ASN856
15	olide 9848024		4	RdRp	ALA125, ASP126, ARG132, HIS133
16	5-hydroxy- 7,8,2',5'-	N.C. CH.	4	S protein	GLN314, GLN613, ILE666, LYS733
10	tetramethoxy flavone 16215023	C <sub>25</sub> H <sub>28</sub> O <sub>12</sub>	2	RdRp	GLN728, TYR728

#### **ADME** and other pharmacokinetic properties

The pharmacokinetic evaluation and ADME (absorption, distribution, metabolism and excretion) properties predictions of pharmacological compounds in advance are important methods to save huge monetary expenses in drug discovery (Zhu *et al.*, 2018). Apart from that, it mainly helps in reducing the potential risk during clinical development as it provides very crucial information to determine whether a compound is suitable to proceed to clinical stage. SWISS-ADME (http://www.swissadme.ch) and admetSAR (http://lmmd.ecust.edu.cn/admetsar2) web servers were used to obtain the ADME and other pharmacokinetic properties of the selected compounds. Among the 35 compounds from *P. amarus* and *A. paniculata* evaluated against SARS-CoV-2 by *in silico* docking analysis, 15

compounds (9 compounds from *P. amarus* and 6 compounds from *A. paniculata* were selected for ADME and other pharmacokinetic properties predictions. All these 15 compounds selected showed better binding ability than the ramdesivir.

The ADME and pharmacokinetic properties of selected phytochemicals are given in the Table 3. Lipinski rule of five (Lipinski, 2004) is a thumb rule to evaluate the drug likeliness or to determine the pharmacological activities that would make it orally active drug in humans. According to the rule the compound should abide following criteria i.e., i) molecular mass less than 500 Daltons, ii) high lipophilicity (expressed as logP less than 5), iii) less than five hydrogen bond donors and iv) less than 10 hydrogen bond acceptors (Petit et al., 2012). Of 15 compounds along with ramdesivir screened, eight compounds (kaempferol, hinokinin, isoandrographolide, quercetin, 14-deoxy-14,15didehydroandrographolide, 14-Deoxyandrographolide, 19-O-Acetyl-14-deoxy-11,12didehydroandrographolide and neoandrographolide) did not violate any of the above rule. However, four compounds viz., astralagin (HBA>10, HBD>5), quercetin-3-O-glucoside (HBA>10, HBD>5), quercitrin (HBA>10, HBD>5) and 5-hydroxy-7, 8, 2', 5'-tetramethoxy flavone (MW>500, HBA>10) along with positive control remdesivir have violated two rules. Three compounds belong to tannins i.e., coriligan (MW>500, HBA>10, HBD>5), furosin (MW>500, HBA>10, HBD>5) and geraniin (MW>500, HBA>10, HBD>5) have violated 3 rules (Table 3). The violation of two or more criteria predicts a molecule to be a non-orally available drug (Lipinski, 2004). However, this rule applies only for oral drugs, the compounds violating the Lipinski's rule of five can be administered through other means after determining its side effects (Benet et al., 2016).

The logS value is directly related to the solubility of the drug in water which ideally ranged between -6.5 and 0.5 (Nisha *et al.*, 2016). All the screened compounds lie within this range. The solubility of the molecules in water significantly facilitates drug developmental activities (Diana *et al.*, 2017). The selected phytochemicals have passed the carcinogenic test and all exhibit non-carcinogenic nature (Table 3). Among 15 phytochemicals, seven phytochemicals (kaempferol, querecetin, hinokinin, isoandrographolide,14-deoxy-14,15-didehydroandrographolide, 14-Deoxyandrographolide and 19-O-acetyl-14-deoxy-11, 12-didehydroandrographolide) have shown high gastro-intestinal or human intestinal absorption. Remaining 8 phytochemicals (astragalin, quercetin-3-O-glucoside, quercetin, coraligin, furosin, geraniin, neoandropholidae and 5-hydroxy-7, 8, 2', 5'-tetramethoxy flavone) have exhibited low gastro-intestinal or human intestinal absorption. The positive control remdesivir also has low human intestinal absorption value. As all the compounds have

exhibited non-carcinogenic nature and good pharmacokinetic properties these compounds individually or in combination may be effective against the SARS-CoV-2. Further, inflammation of lungs resulting in breathing problem is a major symptom in the critical patients suffering from Covid-19 (Li and Ma, 2020). In addition to all the prediction studies, the plant extracts of *P. amarus* have shown anti-inflammatory properties in rat Kupffer cells (Kiemer *et al.*, 2003). The pharmacokinetic predictions suggest that compounds from the two plants species studied in the current study are found to be biologically safe and can be utilized as therapeutics against SARS-CoV-19. However, more studies need to be conducted to decipher the exact mechanism involved in the interaction between these phytochemicals and the viral receptor proteins.

To summarize, all these current studies were done by *in silico* predictions of individual phytochemical originated from *P. amarus* and *A. paniculata*. The information generated here may provide some insights into exploring for potential drugs against SARS-CoV-2 from plants, which can result in discovery of novel drugs. Further, the Ayurveda medications using herbal extracts cannot be undermined in addressing human ailments. Because these extracts contain several compounds, which might have synergistic effect with action on multi-target sites of pathogen. However, the biggest problem in Ayurveda medicine is fixing the dose against the target pathogens. Optimization of this issue supported by scientific evidences for getting required clearances which will have far fetching impact in the harnessing Indian ancient knowledge of medication "Ayurveda" in treating the human ailments including Covid-19 caused by SARS-CoV-2, which threatened the mankind with global pandemic outbreak.

Table 3: Absorption, Distribution, Metabolism and Excretion (ADME) and other pharmacokinetic properties of selected phytochemicals from *Phyllanthus amarus* and *Andrographis paniculata* 

S. L. No	Group	Compound	НІА	Carcinogens	logS value	MW	HBD	НВА	logP value	Rule of 5 violations
1	Positive control	Remdesivir	0.9135 (low)	NC	-3.474	602.58	4	12	3.4	2
2		Astragalin	0.6347 (low)	NC	-2.449	448.38	7	11	1.29	2
3		Kaempferol	0.9881 (high)	NC	-3.142	286.24	4	6	1.7	0
4	Flavanoids	Quercetin	0.9883 (high)	NC	-2.999	302.24	5	7	1.63	0
5		quercetin-3-O-glucoside	0.6468 (low)	NC	-2.449	464.38	8	12	0.94	2
6		Quercitrin	0.7322 (low)	NC	-3.497	448.38	7	11	1.6	2
7	Phenyl propanoid	Hinokinin	0.9784 (high)	NC	-3.113	354.35	0	6	3.08	0
8		corilagin	0.8598 (low)	NC	-2.727	634.45	11	18	1.48	3
9	Tannins	Furosin	0.8598 (low)	NC	-2.75	634.45	11	18	1.48	3
10		geraniin	0.8592 (low)	NC	-3.172	954.66	14	27	0.67	3
11	Bioactive compound	Isoandrographolide	0.9822 (high)	NC	-2.853	350.45	3	5	2.45	0
12		14-deoxy-14,15- didehydroandrographolide	0.9773 (high)	NC	-2.927	332.43	2	4	2.96	0
13	Diterpenes	14-Deoxyandrographolide	0.9899 (high)	NC	-2.816	334.45	2	4	2.79	0
14		19-O-Acetyl-14-deoxy- 11,12- didehydroandrographolide	0.9764 (high)	NC	-4.139	374.47	1	5	3.48	0
15		Neoandrographolide	0.8124 (low)	NC	-3.313	480.59	4	8	3.37	0
16	Flavanoids	5-hydroxy-7,8,2',5'- tetramethoxy flavone	0.4104 (low)	NC	-2.678	520.48	4	12	3.22	2

**HIA**- Human intestinal absorption (probability), **logS**- Solubility, **MW**-Molecular weight (Dalton), **HBD**- Hydrogen bond donor, **HBA**- Hydrogen bond acceptor, **logP**-Liphophilicity, **Rule of 5 violations**- Lipinski rule of 5 and **NC**-Non carcinogen

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