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Carbon dioxide (CO<sub>2</sub>) is an attractive one-carbon (C1) building block in terms of sustainability and abundance. However, its low reactivity limits applications in organic synthesis as typically high-energy reagents are required to drive transformations. Here, we present a redox-neutral C–H carboxylation of arenes and styrenes using a photocatalytic approach. Upon blue-light excitation, the anthrolate anion photocatalyst is able to reduce many aromatic compounds to their corresponding radical anions, which react with CO<sub>2</sub> to afford carboxylic acids. High-throughput screening and computational analysis suggest that a correct balance between electron affinity and nucleophilicity of substrates is essential. This novel methodology enables the carboxylation of numerous aromatic compounds, including many that are not tolerated in classical carboxylation chemistry. Over 50 examples of C–H functionalizations using CO<sub>2</sub> or ketones illustrate a broad applicability. The method opens new opportunities for late-stage C–H carboxylation and valorization of common arenes.

# File list (2)

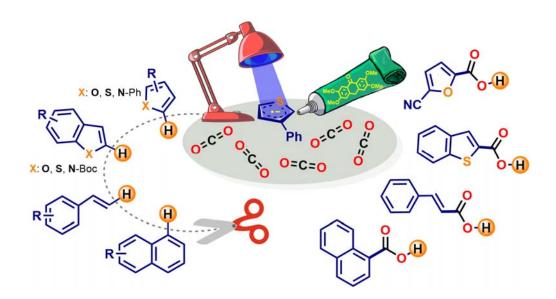
C-H Carboxylation.pdf (657.79 KiB)	view on ChemRxiv • download file
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# Redox-neutral Photocatalytic C–H Carboxylation of Arenes and Styrenes with CO<sub>2</sub>

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Abstract: Carbon dioxide (CO<sub>2</sub>) is an attractive one-carbon (C1) building block in terms of sustainability and abundance. However, its low reactivity limits applications in organic synthesis as typically high-energy reagents are required to drive transformations. Here, we present a redox-neutral C-H carboxylation of arenes and styrenes using a photocatalytic approach. Upon blue-light excitation, the anthrolate anion photocatalyst is able to reduce many aromatic compounds to their corresponding radical anions, which react with CO<sub>2</sub> to afford carboxylic acids. High-throughput screening and computational analysis suggest that a correct balance between electron affinity and nucleophilicity of substrates is essential. This novel methodology enables the carboxylation of numerous aromatic compounds, including many that are not tolerated in classical carboxylation chemistry. Over 50 examples of C-H functionalizations using CO<sub>2</sub> or ketones illustrate a broad applicability. The method opens new opportunities for late-stage C-H carboxylation and valorization of common arenes.

**Keywords:** Photocatalysis, carbon dioxide, carboxylation, C–H functionalization, radical anion, carboxylic acid, 9-anthrone, styrene, heteroarenes

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# INTRODUCTION

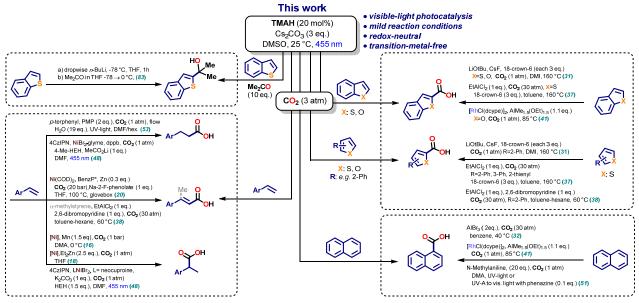
Photosynthesis, the most important photobiological process on our planet, allows photoautotrophs to store energy in form of chemical bonds by absorbing sunlight. Driven by that energy, CO<sub>2</sub> is captured from the atmosphere and serves as carbon feedstock for the organisms to build up sugars and biomass in the Calvin cycle.<sup>1</sup>

Electrochemical and catalytic dihydrogen reductions of carbon dioxide have been developed in the field of renewable energy storage.<sup>2-4</sup> However, the use of CO<sub>2</sub> as a C1 building block in organic synthesis has received far less attention despite resembling the principle of biological carbon fixing process the most.<sup>5</sup> The high thermodynamic stability and kinetic inertness of CO<sub>2</sub> require the use of stoichiometric amounts of reactive reaction partners such as Grignard reagents or organolithium compounds for chemical conversion.<sup>6</sup> Aiming for a better efficiency and an increased atom economy, a variety of catalytic carboxylation methods have been developed. These processes make use of readily available aryl bromides which undergo carboxylation with CO<sub>2</sub> in the presence of catalytic Pd(OAc)<sub>2</sub>, as reported by Martín and Correa.<sup>7</sup> Daugulis showed that Cu(I) catalyzes the carboxylation of aryl iodides. 8 Tsuji and co-workers applied NiCl<sub>2</sub>(PPh<sub>3</sub>)<sub>2</sub> to carboxylate more inert aryl and vinyl chlorides under 1 atm CO<sub>2</sub> at room temperature. The reaction scope was extended to sulfonates, set ester derivatives, 11,12 allylic alcohols, 13 benzylic ammonium salts, 14 arylsulfonium salts 15 and unsaturated hydrocarbons. 16-20 However, all these systems require stoichiometric reducing reagents based on Et<sub>2</sub>Zn, AlMe<sub>3</sub>, Mn and Zn powder or prefunctionalized starting materials.<sup>21</sup> Electrical current may also be used to drive reductive carboxylation chemistry. Buckley and co-workers reported the regioselective hydrocarboxylation of styrenes using a non-sacrificial electrode system.<sup>22</sup> Ackermann et al. showed that allyl chlorides derived from cinnamyl chloride are carboxylated in presence of a cobalt catalyst.<sup>23</sup> Concomitantly, difunctionalizations of alkenes via radical addition and subsequent reduction were reported affording thio-,24 carbo-25-27 phosphono-28 or silylcarboxylation<sup>27</sup> products.

Non-catalytic  $C(sp^2)$ –H carboxylations typically require stoichiometric amounts of either strong bases, such as NaH<sup>29</sup> or LiO'Bu,<sup>30,31</sup> or Lewis acids, like AlX<sub>3</sub> (X = Br, Cl),<sup>32–35</sup> Me<sub>2</sub>AlCl<sup>36</sup> and EtAlCl<sub>2</sub><sup>37,38</sup> to activate CO<sub>2</sub>. Transition metal-catalyzed directed  $C(sp^2)$ –H carboxylation reactions have been reported with Au,<sup>39</sup> Cu<sup>39,40</sup> and Rh<sup>41</sup> complexes.<sup>42</sup> Moreover, direct carboxylation of non-activated  $C(sp^2)$ –H was reported in molten alkali carbonate salts under elevated temperatures (>200 °C) and high CO<sub>2</sub> pressure.<sup>43,44</sup>

More recently, photoredox catalysis has been applied in the field of carboxylation chemistry. Photocatalytic carboxylation of aryl-<sup>45,46</sup> and alkyl-halides<sup>46</sup> were the first transformations to be reported, followed by direct C–H carboxylation of alkynes<sup>47</sup> and styrenes.<sup>48,49</sup> These methods utilize a dual catalytic approach consisting of a photocatalyst and an *in situ* generated low-valent transition metal complex enabled by an excess amount of a sacrificial electron donor. Visible-light mediated benzylic C–H carboxylation was recently reported by the use of 4CzIPN and an organo-silanethiol HAT reagent which allowed to generate carbanions.<sup>50</sup>

Direct UV-light excitation of polyaromatic hydrocarbons in presence of sacrificial amines and CO<sub>2</sub> was reported to yield the corresponding carboxylic acids.<sup>51,52</sup> Jamison employed *p*-terphenyl, which forms a radical anion upon UV-light excitation in the presence of amines.<sup>53</sup> The *p*-terphenyl radical anion is capable of the kinetically slow one-electron reduction of CO<sub>2</sub> to its radical anion, which is used in the hydrocarboxylation of styrenes (see Scheme 1). Murakami and co-workers employed UV-excited xanthone as hydrogen-atom-transfer (HAT) reagent in combination with a Cu complex or a Ni catalyst to carboxylate allylic<sup>54</sup> and benzylic<sup>55</sup> C(sp³)—H respectively.



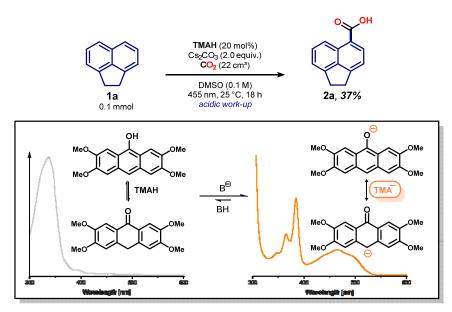
Scheme 1. Overview of representative literature procedures for the direct C-H carboxylation with CO<sub>2</sub> or hydroxyalkylation with acetone considering the herein discussed substrate classes.

However, despite the great progress achieved in thermal and photochemical carboxylation methods, the efficient redox-neutral carboxylation of  $C(sp^2)$ —H in arenes, heteroarenes and alkenes remains challenging. Here, we report a mechanistically different catalytic approach in which aromatic compounds are converted into their radical anions by photoinduced single-electron transfer (SET) from a visible-light excited anthrolate anion. The generated nucleophilic arene radical anions react with  $CO_2$  to provide (hetero)aromatic carboxylic- and cinnamic acids.

# **RESULTS & DISCUSSION**

Recently, we showed that upon photoexcitation, the anionic form of commercially available 9-anthrone and derivatives (Scheme 2 and Figure S6, Supplemental Information) readily generate strong reductants capable of activating aryl chlorides.<sup>56</sup> While comparing reported reduction potentials of various aromatic compounds, we noticed that many arenes lay within the range of the approximated excited state oxidation potential of the strongest

photo-reductant 2,3,6,7-tetramethoxyanthracen-9(10*H*)-one (**TMAH**) [ $E_{ox}$  (**TMA'/TMA**<sup>-\*</sup>) = -2.92 V vs. SCE] shown in that series. We thus envisioned a direct activation of arenes via radical anion formation, which may subsequently react with CO<sub>2</sub> to form aromatic carboxylic acids. Strong carbon nucleophiles (e.g. organolithium and -magnesium reagents)<sup>57-60</sup> or carbanions are well known to react with CO<sub>2</sub>. By contrast, aromatic radical anions formed in the presence of alkali metal have always been considered poor nucleophiles,<sup>61</sup> yet still showed reactivity towards CO<sub>2</sub>.  $^{62,63}$  With a strongly reducing photoredox catalyst in hand and inspired by early literature reports, we questioned if a similar reactivity of aromatic radical anions towards carbon dioxide can be obtained under much milder photocatalytic conditions.



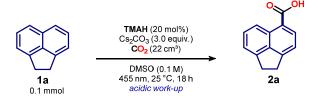
**Scheme 2.** (top) Photocatalytic C-H carboxylation of acenaphthene **1a**; (bottom) Base-promoted formation of TMA<sup>-</sup> and the influence on the absorption spectrum.

### Initial experiments and optimization

We chose acenaphthene (1a, Scheme 2) as a model substrate and applied the established combination of TMAH as the photocatalyst and cesium carbonate as the base. To our delight, after 18 hours of irradiation with blue LED light and acidic work-up, the desired carboxylic acid 2a could be isolated in 37% yield as a single regioisomer. Encouraged by this first result we run an intensive screening of the reaction conditions (Table 1). During the optimization studies, we observed that the reaction outcome was dependent on the amount of Cs<sub>2</sub>CO<sub>3</sub>. The use of less than 3 equivalents of base led to significantly lower product yield (entry 2) while more equivalents of Cs<sub>2</sub>CO<sub>3</sub> reduced the yield (entry 3). A lower catalyst loading reduced the overall amount of base required, while the carboxylated product 2a was still obtained in good yield (entry 4-5). K<sub>2</sub>CO<sub>3</sub>, although being scarcely soluble in DMSO, was also able to promote the carboxylation reaction and useful product yields were obtained in combination with crown-ether (entry 6). Monitoring the reaction progress over time (Figure S8, Supplemental

Information) showed that the reaction was not complete after 6 hours (entry 7). An overpressure of CO<sub>2</sub> was found to be beneficial for the reaction outcome (entries 8 and 9). The solubility of CO<sub>2</sub> is reported to be higher in DMF compared to DMSO, yet better yield was obtained in the latter (entry 10).<sup>64</sup> When using green light (535 nm), a reduced product yield was obtained (entry 11), which can be explained by both weaker catalyst absorption and LED radiant flux. Control experiments revealed that all reagents and light are crucial, as no product was detected in the absence of either photocatalyst, cesium carbonate, carbon dioxide or light (entry 12-15).

Table 1. Optimized reaction conditions and effects upon deviation.

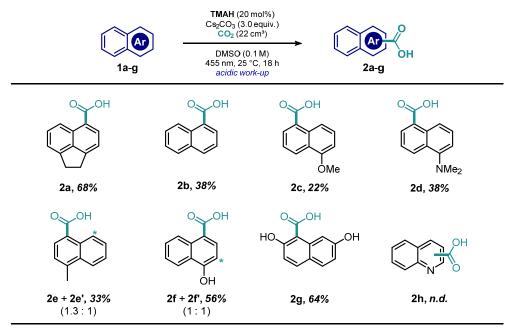


Entry	Deviations from optimized conditions	Yield 2a [%] <sup>a</sup>
1	none	68 <sup>b</sup>
2	Cs <sub>2</sub> CO <sub>3</sub> (2 equiv.)	$37^{\rm b}$
3	Cs <sub>2</sub> CO <sub>3</sub> (4 equiv.)	59
4	TMAH (5 mol%), Cs <sub>2</sub> CO <sub>3</sub> (2 equiv.)	54
5	TMAH (10 mol%), Cs <sub>2</sub> CO <sub>3</sub> (2 equiv.)	60
6°	TMAH (10 mol%), K <sub>2</sub> CO <sub>3</sub> instead of Cs <sub>2</sub> CO <sub>3</sub> , 18-crown-6	47
7	6 hours instead of 18 hours	44
8	no CO <sub>2</sub> pressure (1 atm)	37
9	11 cm <sup>3</sup> CO <sub>2</sub> instead of 22 cm <sup>3</sup>	48
10	DMF instead of DMSO	35
11 <sup>d</sup>	535 nm instead of 455 nm	29
12	no TMAH	n.d.
13	no Cs <sub>2</sub> CO <sub>3</sub>	n.d.
14	$N_2$ (1 atm) instead of $CO_2$	n.d.
15e	no light	n.d.

Optimized reaction conditions: 1a (0.1 mmol), TMAH (20 mol%) and  $Cs_2CO_3$  (0.3 mmol) were added to a 5 mL crimp top vial equipped with a stirring bar. The vial was sealed, evacuated and backfilled with  $CO_2$  (5×). Degassed, anhydrous DMSO (1 mL) was added *via* syringe. The septum was further sealed with Parafilm® and gaseous  $CO_2$  (22 cm³) was added to the headspace *via* syringe. While stirring, the reaction was irradiated from the bottom side (blue LED,  $455 \pm 15$  nm) and constant temperature was maintained by an aluminum cooling block and a water-cooling circuit. For complete optimization table, please see Table S1, Supplemental Information. n.d. = not detected. <sup>a</sup> Product yield was determined after acidic work-up by crude <sup>1</sup>H-NMR with an internal standard. <sup>b</sup> Combined isolated yield of four reactions. <sup>c</sup> Crown ether 18-crown-6 (1 equiv.) was added to the reaction. <sup>d</sup> Radiant flux is lowered by a factor of 8 compared to 455 nm LED (see Supplemental Information). <sup>c</sup> Reaction mixture was stirred in the dark.

# Substrate scope of the carboxylation reaction

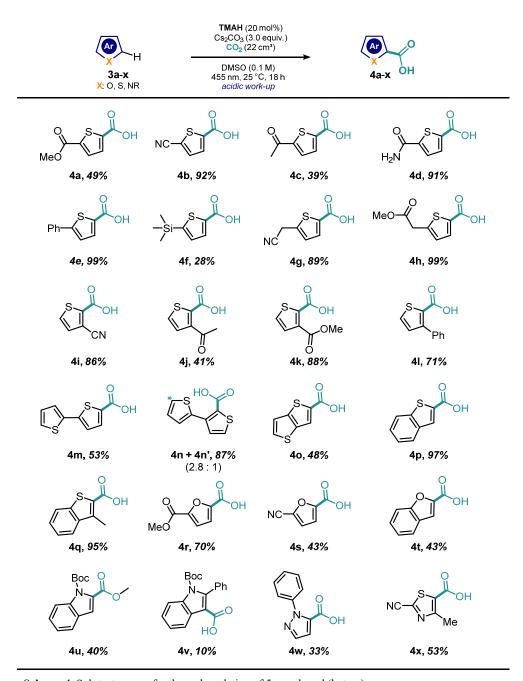
With the optimized reaction conditions in hand (*cf.*, Table 1), we explored the scope of this novel transformation. Naphthalene derivatives were investigated, as their reported potentials are in a feasible range (-2.49 up to -2.65 V vs. SCE)<sup>65</sup> for reduction by the photocatalyst (Scheme 3). We were pleased to see that unsubstituted as well as substituted naphthalene derivatives were converted to the corresponding aromatic carboxylic acids (**2b-g**) and could be isolated in useful yields. The regioselectivity of the reaction was found to be affected by strong electron-donating groups (-OMe **2c**, -NMe<sub>2</sub> **2d**) in the C1-position giving selectively 5-naphtoic acids as single regioisomers. In contrast, the directing effect of electronically neutral substituents (-Me, **2e**) was minor and led to a mixture of 4- and 5-naphtoic acid. Remarkably, carboxylation in the C8-position was not observed. Notably, unprotected hydroxyl groups (**2f**, **2g**) were tolerated. Utilizing 1-naphthol (**1f**) led to the formation of two regioisomers of the corresponding acid in 2- and 4-position. 2,7-Dihydroxynaphthalene (**1g**) reacted smoothly under our reaction conditions to yield the corresponding 1-naphtoic acid **2g** as a single regioisomer. Quinoline (**1h**), isoquinoline or quinazoline, although quenching the photoexcited state of the catalyst, failed to yield any product.



**Scheme 3.** Substrate scope for the carboxylation of naphthalene derivatives.

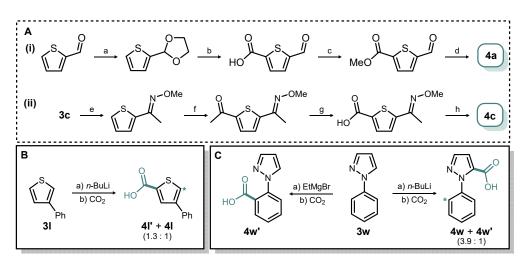
Pleasingly, many other heteroaromatic compounds were suitable substrates for our carboxylation method (Scheme 4). Thiophenes, bearing electron-deficient (3a-d, 3f, 3i-k) and -neutral (3e, 3g-h, 3l-n) substituents smoothly converted into the corresponding thiophenecarboxylic acids 4a-n in good to excellent yield. Remarkably, a broad range of functional groups including ketones, esters, amides, -CH<sub>2</sub>CO<sub>2</sub>Me, -CH<sub>2</sub>CN, phenyl-, trimethylsilyl-,

nitrile were tolerated. However, unfunctionalized thiophene could not be activated by the photocatalyst (see Figure S11b, Supplemental Information). Due to the mild nature of this reaction, we were pleased to see that regionselectivity was maintained even with substrates containing acidic C-H (4c, 4j) or active methylene groups (4g-h). This is in contrast to reported base promoted methods that are usually selective for the most acidic position of the substrates and that fail in the presence of sensitive functionalities.



Scheme 4. Substrate scope for the carboxylation of 5-membered (hetero)arenes.

Our procedure allowed for the conversion of methyl thiophene-2-carboxylate to 5-(methoxycarbonyl)thiophene-2-carboxylic acid (4a) in one step, providing a much shorter route than by using other reported methods [Scheme 5, A(i)]. 66 Moreover, our method allowed the synthesis of 5-carboxy-2-acetylthiophene (4c), an important building block for the synthesis of the alpha/beta blocker arotinolol, 67 in one step from 1-(thiophen-2-yl)ethan-1-one (3c). This is significantly shorter than well-established synthetic routes [Scheme 5, A(ii)]. 68 Remarkably, we also found that 3-substituted thiophenes were exclusively carboxylated in the 2-position (4i-1) which highlights the excellent regioselectivity of this reaction. In comparison, a previous literature report on the lithiation of 3-phenylthiophene (31) and subsequent carboxylation led to a mixture of 41 and 41' (Scheme 5, B). 69 Perfect regioselectivity was also observed for the photocatalyzed carboxylation of 1-phenylpyrazole where only 4w was obtained. In this case, due to the chelating effect of nitrogen, the use of organometallic reagents leads to product mixtures (*n*-BuLi)<sup>70</sup> or to an inverse regioselectivity (EtMgBr)<sup>71</sup> (Scheme 5, C). This photocatalyzed carboxylation method could also be extended to benzothiophenes (4p-q), furans (4r-s), benzofuran 4t, Boc-protected indoles (4u-v) and thiocarbazole 4x. Non-protected 1*H*-indoles however, were carboxylated at the nitrogen atom, as reported in literature. 30



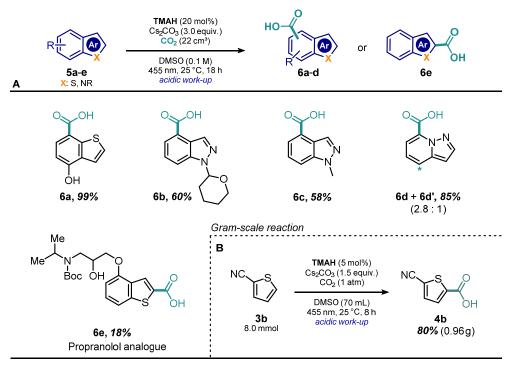
**Scheme 5.** Commonly applied synthetic routes to obtain products **4a** and **4c** (A). Regioselectivity of organometallic carboxylation (B+C).

(A) Reported four-step synthesis for compound 4a (i)<sup>66</sup> and 4c (ii)<sup>68</sup>; conditions: (a) ethane-1,2-diol, Al<sub>2</sub>O<sub>3</sub>, CCl<sub>4</sub>,  $\Delta$ , 48 h; (b) n-BuLi, THF, -78 °C, then CO<sub>2</sub> followed by H<sub>2</sub>SO<sub>4</sub> (10%); (c) MeI, Na<sub>2</sub>CO<sub>3</sub>, DMF, 20 °C, 48 h; (d) Jones reagent; (e) MeONH<sub>2</sub>·HCl, Na<sub>2</sub>CO<sub>3</sub>, H<sub>2</sub>O, MeOH, AcOH (pH 5),  $\Delta$ , 3 h; (f) ZnCl<sub>2</sub>, CHCl<sub>3</sub>, Ac<sub>2</sub>O, 100 °C, 12 h, r.t., aq. HCl (20%) (g) MeOH, aq. NaOCl (5.5%), 70 °C, 4 h, r.t., HCl conc.; (h) aq. HCl, 65 °C, 12 h.

- (B) *n*-BuLi promoted carboxylation of **31** leads to a mixture of regioisomers.
- (C) Organometallic methods for carboxylation of 1-phenylpyrazole. Due to the chelating effect of nitrogen, the use of organometallic reagents causes a mixture of regioisomers (*n*-BuLi) or leads to an inversion of regioselectivity (EtMgBr) yielding benzoic acid **4w'** as single product.

Noteworthy, modifying the conditions during the reaction work-up of **4r** allowed the formation of either 2,5-furandicarboxylic acid (FDCA) or dimethyl 2,5-furandicarboxylate (DMFDC, see Scheme S4, Supplemental Information). Both are important monomers for the manufacture of polyesters derived from biomass, including

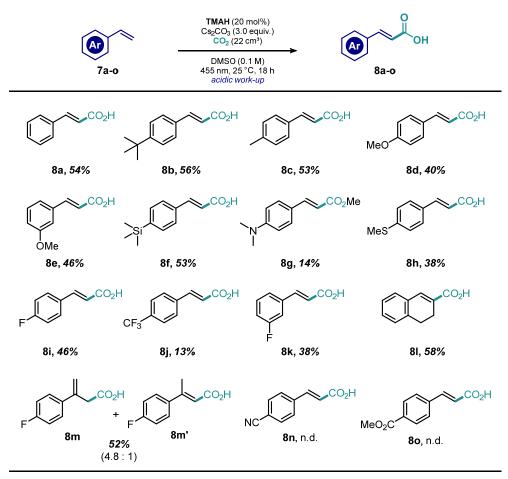
polyethylene furandicarboxylate (PEF), a potential large-scale substitute for fossil-based polyethylene terephthalate (PET).  $^{72,73}$  Lignocellulose is converted into furfural on an industrial scale  $^{74}$  and catalytic follow-up procedures have been reported to yield methyl furoate quantitatively.  $^{75,76}$  Starting from methyl furan-2-carboxylate  $3\mathbf{r}$ , our procedure offers a new one-step synthetic route to form lignocellulose-derived monomers. Remarkably, all examined heterocycles were carboxylated at the five-membered ring in  $\alpha$ -position (except  $4\mathbf{v}$ , where the  $\alpha$ -position is blocked) to the heteroatom. Contrarily, hydroxybenzothiophene  $6\mathbf{a}$  or condensed heterocycles such as indazoles ( $6\mathbf{b}$ - $\mathbf{c}$ ) and pyrazolo[1,5-a]pyridine ( $6\mathbf{d}$ ) showed exclusive selectivity for the six-membered ring (Scheme 6, A). With mild carboxylation conditions in hand, we postulated that this methodology could be applied for the late-stage functionalization of biologically active molecules. To this end, a Boc-protected thiophene analogue of propranolol, a well-established beta blocker bearing a free hydroxyl group in the side chain, was subjected to our reaction conditions. Pleasantly, regioselective carboxylation was successfully achieved to provide  $6\mathbf{e}$ , albeit in modest yield.



**Scheme 6.** (A) Substrate scope for the carboxylation of (hetero)arenes on the six-membered ring and example of late-stage functionalization. (B) Reaction conditions for the gram-scale carboxylation of **3b**.

To demonstrate the scalability of our reaction, we repeated the synthesis of **4b** on a gram-scale. 2-Cyanothiophene **3b** was reacted in a custom-built glass reactor (see Figure S2, Supplemental Information) with a reduced amount of both **TMAH** (5 mol%) and cesium carbonate (1.5 equiv.) in DMSO (Scheme 6B). Remarkably, as working with CO<sub>2</sub> overpressure was not possible with this reaction vessel, a gentle stream of CO<sub>2</sub> was sufficient to obtain the product in good yield.

We then questioned whether our photocatalytic system could be further utilized for the direct carboxylation of other stabilized sp<sup>2</sup>-hybridized carbon atoms. To this end, we examined styrene derivatives (Scheme 7) and we were delighted to see that under the presented redox-neutral conditions only the corresponding *trans*-cinnamic acids were obtained. Our protocol thus provides a complementary method to previous net-reductive approaches where excess of sacrificial reductant or electrical current yielded hydrocarboxylated products (*cf.*, Scheme 1).<sup>22,48,53</sup> Despite the limitation posed by competing polymerization reactions, a variety of vinyl benzenes could be converted into *trans*-cinnamic acid derivatives **8b-l** (Scheme 7), which find applications in the food industry, material science<sup>77</sup> and cosmetics.<sup>78</sup> While styrene derivatives bearing electron-donating substituents (**7d-f**, **7h**) reacted smoothly, the reaction with the electron-poor 4-(trifluoromethyl)styrene resulted in low product yield. 4-Cyanostyrene (**7n**) or methyl-4-vinylbenzoate (**7o**) were not suitable substrates under these conditions. As the electron transfer from the excited photocatalyst to an electron-poor styrene is thermodynamically favored, we postulated that electron-withdrawing groups stabilize the negative charge and reduce the nucleophilicity of the corresponding radical anion. When using  $\alpha$ -methylstyrene, we observed the formation of 3-aryl-3-butenoic acid **8m** in favor of the thermodynamically more stable  $\alpha$ , $\beta$ -unsaturated acid **8m'**. <sup>38</sup>



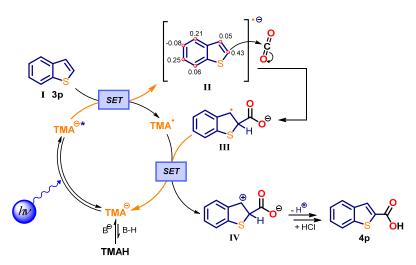
**Scheme 7.** Substrate scope for the carboxylation of styrene derivatives.

# **Mechanistic Insights**

We propose a photoinduced single-electron transfer (SET) from the excited TMA<sup>-\*</sup> to the substrate as the first step. The excited photocatalyst TMA<sup>\*</sup> is strongly emissive and its luminescence decay follows first-order kinetics (see Figure S4a-b and Figure S5, Supplemental Information). In presence of chosen (hetero)arenes (cf., Scheme 4) as well as styrenes (cf., Scheme 7) we observed a decrease in the luminescence lifetime of TMA<sup>-\*</sup>. Based on the obtained data, a Stern-Volmer plot was derived (see Figure S11a-b, Supplemental Information). The luminescence lifetime of TMA<sup>-\*</sup> in a CO<sub>2</sub>-saturated solution of DMSO remains almost unchanged, indicating that CO<sub>2</sub> is not reduced by the photocatalyst. Reduction of CO<sub>2</sub> by SET to achieve carbon-bond formation has been reported, but it requires reagents such as p-terphenyl radical anion<sup>53,79</sup>. Although a direct reduction of CO<sub>2</sub> (-2.21 V vs. SCE in DMF)<sup>80</sup> by **TMA**<sup>-\*</sup> is thermodynamically feasible, we assume a kinetic barrier preventing the formation of the bent CO<sub>2</sub> radical anion within the excited state lifetime of TMA<sup>-\*</sup>. Thus, we ascertained that the productive pathway is dominated by the formation of an aromatic radical anion, which subsequently reacts with CO<sub>2</sub> via nucleophilic addition. In order to get further insight into the reaction mechanism, we tested other electrophiles than CO<sub>2</sub>. We decided to use ketones for this study, as single-electron reduction from the excited photocatalyst to the ketone would give a ketyl radical anion. In sharp contrast to the radical anion of CO2, ketyl radical anions are considered as electron-rich species acting as single-electron reductants rather than forming C-C bonds via radical reactions. 81 When benzo[b]thiophene (3p, Scheme 8) was reacted with acetone (9a), the corresponding tertiary alcohol adduct **9pa** was formed in good yield. The luminescence lifetime of **TMA**<sup>-\*</sup> remains unchanged upon titration with acetone (-2.84 V vs. SCE in DMF)<sup>82</sup> and thus, the formation of a ketyl radical anion is unlikely (Figure S11d, Supplemental Information). Remarkably, similar transformations require very harsh reaction conditions (-78 °C, excess of *n*-BuLi) and are not viable in a one-pot procedure.<sup>83</sup> Moderate yield of the resulting tertiary alcohol were also obtained using cyclic ketone 9b and non-conjugated enone 9c as electrophiles.

**Scheme 8.** Mechanistic investigations: Probing the possibility of other electrophiles as viable candidates for reactivity with aromatic radical anions.

In addition, we performed deuterium-labeling experiments using D<sub>2</sub>O and 'BuOD respectively. Upon formation of the nucleophilic arene radical anion, we envisioned a fast acid-base reaction followed by reoxidation and deprotonation to yield a mixture of H and D in the substrate (see Scheme S3, Supplemental Information). However, as water in the reaction mixture was found to be detrimental and protic solvents were shown to inhibit the reaction (cf., Table S1, Supplemental Information), only small amounts of incorporated deuterium were detected using benzothiophene 3q (Table S3, Supplemental Information). Based on the aforementioned results we propose the following reaction mechanism for the photocatalytic C-H carboxylation of (hetero)arenes (Scheme 9). In the presence of a base, the pre-catalyst **TMAH** is chemically activated by deprotonation to form an anionic species TMA<sup>-</sup>, indicated by the solution colour change. Upon irradiation with visible light (455 nm) a strongly-reducing excited anion TMA<sup>-\*</sup> is formed. The excited state is then quenched by the arene I via SET to afford the radical of the photocatalyst TMA' and the electron rich radical anion of the arene (II). The Mulliken spin population for each aromatic sp<sup>2</sup>-hybridized carbon, indicative of reactivity towards CO<sub>2</sub>, is shown for the benzothiophene radical anion (II). In the bond-forming step CO2 is attacked by II to generate a radical carboxylate III as an intermediate. The catalytic cycle is closed via SET to recapture the active catalyst TMA followed by re-aromatization of the cationic arene IV upon deprotonation. Despite the weakly oxidizing nature of the photocatalyst, the electron transfer from III to TMA' may be driven by the stabilization energy gained upon re-aromatization of IV. It remains to be shown if this re-aromatization occurs via electron transfer and subsequent deprotonation as proposed, or via a direct H-atom abstraction by the oxidized photocatalyst.



**Scheme 9.** Proposed mechanism for the C-H carboxylation of (hetero)arenes exemplified by the carboxylation of benzothiophene.

During the exploration of the substrate scope, we found that several aromatic compounds that quench the excited state of the photocatalyst did not undergo carboxylation (see Figure S11c, Supplemental Information). While some functional groups are understandably not tolerated under our reaction conditions, such as aromatic halides

(these may undergo fast mesolytic bond-cleavage to form aryl radicals) or highly electrophilic moieties, we were surprised that only certain radical anions were reacting with CO<sub>2</sub>. In order to determine the required electronic characteristics for reactivity and to improve our insight regarding functional group tolerance, high-throughput screening (HTS) of various arenes containing a large array of functionalities and differing in complexity was carried out (see Figure S9, Supplemental Information). The analysis of the HTS outcome indicates that aldehydes, halides (with the exception of fluorine), aliphatic amines, 6-membered N-heterocycles and nitro groups are not compatible with the reaction conditions. Beyond functional group interference, the applicability of our methodology is related to the arene electron affinity and the nucleophilicity of the resulting radical anion. Arenes may be able to accept an electron with ease, but the radical anion formed may not be nucleophilic enough to add to CO<sub>2</sub>. Conversely, some aromatic radical anions may be highly nucleophilic, but their formation may be beyond the reductive capabilities of the photocatalyst or a functional group present may be reduced instead. The estimated electron affinities of the arenes, and the Mulliken spin population and charges for the arene radical anions were derived from DFT calculations. The importance of a correct balance of electron affinity of the arene and the nucleophilicity of the aromatic radical anion for a successful reaction is illustrated in Schemes S12a-d (Supplemental Information). The regioselectivity is generally well predicted by comparing the Mulliken spin population for the aromatic carbons in each substrate.

# Conclusion

We have developed a mild, direct, redox-neutral and transition-metal-free insertion of CO<sub>2</sub> into non-prefunctionalized C(sp<sup>2</sup>)–H bonds, leading to an efficient method for producing valuable aromatic carboxylic acids and *trans*-cinnamic acids in a single operation. A reaction performed on gram-scale demonstrated the scalability of this carboxylation method, while ketones could be used as alternative electrophiles to CO<sub>2</sub> yielding tertiary alcohols. The scope of the reaction can be predicted by DFT-estimated reduction potential of the substrates and nucleophilicity of the intermediate arene radical anions. These findings may open new opportunities for atom-economic and energy efficient use of CO<sub>2</sub> as a C1 building block in the chemical processing of aromatic hydrocarbons, as well as for developing new photocatalytic late-stage functionalizations of drug-like compounds.

## **Experimental Procedures**

Full experimental procedures are provided in the Supplemental Information

# **Supplemental Information**

Supplemental Information can be found online.

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### **Author Contributions**

M.S. developed the catalytic system, optimized the reaction, isolated the compounds 2a-d, 4a-f, 4i-t, 4v-w, 8a-m, 9pa-pc, designed and carried out the gram-scale reaction and conducted all experiments to investigate the reaction mechanism. T.D.S. designed and performed the high-throughput-screening and isolated compounds 2f-g, 4g-h, 4u, 4x, 6a-e. F.F. prepared compounds and isolated compound 2e. M.S. and T.D.S. wrote the manuscript and Supplemental Information with input from all of the authors. P.B. conducted the computational studies. M.J.J., G.B. and B.K. supervised the project.

### **Declaration of Interests**

The authors declare no competing interests. T.D.S., P.B., M.J.J., G.B. are employees and shareholders of AstraZeneca.

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# **Supplemental Information**

# Redox-neutral Photocatalytic C-H Carboxylation of Arenes and Styrenes with CO<sub>2</sub>

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# 1 - General Experimental Details

All required fine chemicals were purchased from commercial suppliers (abcr, Acros, Alfa Aesar, Fluka, Fluorochem, Merck, Sigma Aldrich, TCI) and were used directly without purification unless stated otherwise. All air and moisture sensitive reactions were carried out under nitrogen atmosphere using standard Schlenk manifold technique. Anhydrous DMSO was used directly from the bottle or dried using activated 4Å molecular sieves. <sup>1</sup>H and <sup>13</sup>C Nuclear Magnetic Resonance (NMR) spectra were acquired at room temperature with field strengths as indicated and were referenced to CDCl<sub>3</sub> (7.26 and 77.16 ppm for <sup>1</sup>H and <sup>13</sup>C respectively), DMSO-d6 (2.50 and 39.52 for <sup>1</sup>H and <sup>13</sup>C respectively) or CD<sub>3</sub>OD (3.31 and 49.0 ppm for <sup>1</sup>H and <sup>13</sup>C respectively). <sup>1</sup>H-NMR coupling constants are reported in Hertz and refer to apparent multiplicities and not true coupling constants. Data are reported as follows: chemical shift, multiplicity (s = singlet, bs = broad singlet, d = doublet, t = triplet, q = quartet, qi = quintet, sx = sextet, sp = septet, m = multiplet, dd = doublet of doublets, etc.), coupling constants, integration, proton assignment (determined by 2D NMR experiments: COSY, HSQC and HMBC) where possible. <sup>13</sup>C-NMR assignment was aided using DEPT 135 techniques (DEPT = distortion less enhancement by polarization transfer) to distinguish CH<sub>2</sub> groups from CH and CH<sub>3</sub> groups and to assign quaternary carbon atoms (C<sub>q</sub>). <sup>19</sup>F-NMR spectra were recorded for compounds containing fluorine atoms.

Analytical TLC was performed on silica gel coated aluminium sheets (Merck, TLC Silica gel 60 F<sub>254</sub>) Compounds were visualized by exposure to UV-light (254 or 366 nm) or by dipping the plates in staining solutions (permanganate stain, bromocresol green stain, ceric ammonium molybdate stain) followed by heating. Flash column chromatography was performed using Merck Silica Gel 60 (40-63 μm) & Medium pressure liquid chromatography (MPLC) was performed on a Grace Reveleris® X2 from Büchi with built-in UV-detector and fraction collector using Biotage® sfär silica HC D 20 μm column cartridges or on a Biotage® Isolera One flash purification system using flash silica gel. All mixed solvent eluents are reported as v/v solutions. High resolution mass spectrometry (HRMS) were performed at the Central Analytical Laboratory of the University of Regensburg. Mass spectra were recorded on a Finnigan MAT 95, ThermoQuest Finnigan TSQ 7000, Finnigan MAT SSQ 710 A or Agilent Q-TOF 6540 UHD instrument and a Waters Acquity UPLC system equipped with Waters PDA, sample manager, sample organiser, column oven and Waters Xevo QTOF mass spectrometer. Photoreactions in regular scale were irradiated with blue LEDs (OSRAM Oslon SSL 80 royal-blue,  $\lambda$  = 455 nm ( $\pm$  15), average radiant flux 232  $\pm$  23 mW, 2.9 V, 350 mA) or green LEDs ( $\lambda$  = 535 nm, average radiant flux,  $29 \pm 5$  mW) and were exposed to light from the flat bottom side of the vial. The temperature of the reaction mixtures was controlled by a water-cooling circuit consisting of an aluminium cooling block connected to a thermostat (Figure S1). An exemplary reaction in larger scale was carried out in a custom-built glass reactor which upon vigorous stirring generates a thin film of the reaction mixture between the reaction vessel and an attached cold finger. A hose which was dipped in the solution provided CO<sub>2</sub> gas from the cylinder. The reaction vessel was surrounded by blue LED arrays (OSRAM Oslon SSL 80 LT-2010,  $\lambda = 451$  nm, 700 mA) generating a total radiant flux of 12 W (Figure S2). For the high-throughput screening experiments, Kessil PR160L 456 nm LEDs were used. Cyclic voltammetry measurements were performed with a three-electrode system consisting of a glassy carbon working electrode, a platinum wire counter electrode and a silver wire as a reference electrode. Data was processed on a potentiostat PGSTAT302N from Metrohm Autolab. Prior to the measurement the solvent DMSO (dry) was degassed with argon and TBATFB (0.1M) was added as supporting electrolyte. All experiments were performed under argon atmosphere. Ferrocene was used as an internal reference. Measurements were performed at a scan rate of 0.05 Vs<sup>-1</sup>. Potentials are reported against saturated calomel electrode (SCE) as reference. UV-Vis measurements were performed on an Agilent Cary 4000 UV-Vis Spectrophotometer. Prior to measurements a solvent blank was recorded and subtracted. Precision cells (1×1 cm) made of quartz SUPRASIL® from Hellma® Analytics were used. Luminescence measurements were performed on a Horiba® Scientific FluoroMax-4 instrument using the above-mentioned quartz cells. Luminescence lifetime measurements were performed on a Horiba® Scientific DeltraPro<sup>TM</sup> fluorescence lifetime system using a 452 nm laser diode from Horiba® Scientific DeltaDiode<sup>TM</sup> as excitation source and above-mentioned quartz cells. The instrument response function (IRF) was determined prior to measurements by using colloidal silica (LUDOX®) in water.

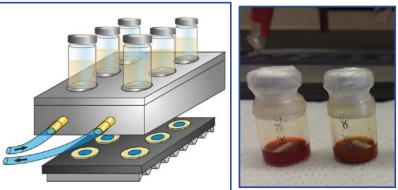


Figure S1. Schematic picture of the setup for photoreactions (left); crimp vials charged with stirring bar and reaction mixture and sealed with aluminium crimp seal with septum and Parafilm®

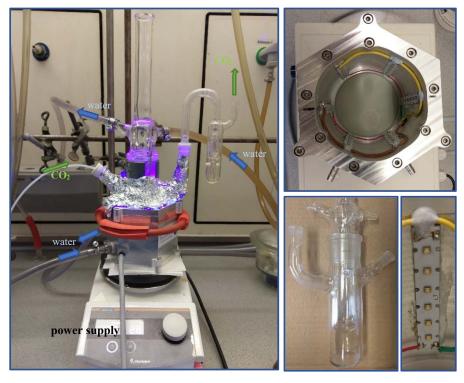


Figure S2. Custom-built glass reactor for upscaling of the photocatalytic carboxylation reaction.

# 2 - Catalyst Synthesis and Physicochemical Properties

# 2.1 - Synthesis of the Photocatalyst

2,3,6,7-tetramethoxyanthracen-9(10*H*)-one **TMAH** was synthesized in three steps with an overall yield of 33%. The catalyst in its neutral form is a bench stable compound and can be stored easily.

Scheme S1. Overview of the synthetic steps in the synthesis of the used photocatalyst TMAH; overall yield 33% (3 steps).

Synthesis of 9,10-diethyl-2,3,6,7-tetramethoxyanthracene: Referring to literature known procedures<sup>1,2</sup>, a 250 mL round bottom flask equipped with stirring bar was charged with  $H_2SO_4$  (70 mL, 70% v/v in  $H_2O$ ) and veratrole (12.8 mL, 0.1 mol, 1 equiv.) and the resulting mixture was cooled to -10 °C. Under vigorous stirring, propanal (14.3 mL, 0.2 mol, 2 equiv.) was added dropwise *via* a syringe pump within 2 hrs. Care was taken, that the reaction temperature during addition of aldehyde was kept below 0 °C. The reaction mixture was poured into ice water (ca. 500 mL) and the resulting precipitate was filtered off and washed with water. The filter cake was dried over night by lyophilization and was washed in boiling EtOH. The precipitate was filtered off, washed with EtOH and dried under vacuo to give the title compound (8.06 g, 23 mmol, 46%) as pale-yellow powder. <sup>1</sup>H-NMR (400 MHz, Chloroform-*d*)  $\delta$  7.41 (s, 4H), 4.07 (s, 12H), 3.47 (q, J = 7.6 Hz, 4H), 1.44 (t, J = 7.6 Hz, 6H). <sup>13</sup>C-NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  149.0 (C<sub>q</sub>), 130.7 (C<sub>q</sub>), 125.1 (C<sub>q</sub>), 102.4, 55.8, 22.0 (CH<sub>2</sub>), 14.6.

**Synthesis of 2,3,6,7- tetramethoxy-9,10-anthraquinone**: According to a literature known procedure<sup>3</sup> a 500 mL round bottom flask was charged with 9,10-diethyl-2,3,6,7-tetramethoxyanhtracene (8.0 g, 22.6 mmol, 1 equiv.) and K<sub>2</sub>Cr<sub>2</sub>O<sub>7</sub> (33.2 g, 113 mmol, 5 equiv.) and the solids were suspended in glacial acetic acid (270 mL). The resulting mixture was heated to 90 °C for 3 hrs. After the mixture was cooled

to ambient temperature the yellow precipitate was filtered off and washed several times with water to remove excess of  $K_2Cr_2O_7$ . The filter cake was freeze-dried and finally washed with  $Et_2O$  and dried in vacuo to afford the title compound as yellow powder (5.67 g, 17.3 mmol, 76%), which was used without further purification for the next step. <sup>1</sup>H-NMR (300 MHz, Chloroform-d)  $\delta$  7.68 (s, 4H), 4.07 (s, 12H). <sup>13</sup>C-NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  182.1(C<sub>q</sub>), 153.6 (C<sub>q</sub>), 128.6 (C<sub>q</sub>), 108.5, 56.7.

Synthesis of 2,3,6,7-tetramethoxyanthracen-9(10H)-one (TMAH): Referring to a literature known procedure<sup>4</sup> a 500 mL Schlenk flask equipped with stirring bar and condenser was charged with 2,3,6,7-tetramethoxy-9,10-anthraquinone (5.60 g, 17.1 mmol, 1 equiv) and zinc dust (3.40 g, 52.0 mmol, 3.1 equiv.). The flask was set under  $N_2$  atmosphere and a mixture of aq. ammonia solution (135 mL, 25%), EtOH (135 mL) and water (135 mL) was added. The resulting mixture was refluxed for 5 hrs under  $N_2$  atmosphere and vigorous stirring. The mixture was allowed to cool to ambient temperature and was poured in ice water (ca. 1 L). Conc. HCl (150 mL) was added to dissolve excess zinc and the mixture was stirred overnight. The turbid solution was filtered and the residue was washed several times with water and was freeze-dried to yield **TMAH** as pale-yellow powder (5.10 g, 16.2 mmol, 95%). <sup>1</sup>H-NMR (400 MHz, Chloroform-d)  $\delta$  7.79 (s, 2H), 6.82 (s, 2H), 4.15 (s, 2H), 3.98 (d, J = 7.3 Hz, 12H). <sup>13</sup>C-NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  182.4 ( $C_q$ ), 153.1 ( $C_q$ ), 148.6 ( $C_q$ ), 135.1 ( $C_q$ ), 125.4 ( $C_q$ ), 109.7, 108.5, 56.2, 32.0 (CH<sub>2</sub>). Data in accordance with the literature.<sup>3</sup>

# 2.2 - Spectroscopic and Photochemical Characteristics

The properties of the used photocatalyst (PC) were investigated in various spectroscopic experiments.

# 2.2.1 - UV-Vis absorption

The absorption of the photocatalyst was recorded in dry, degassed DMSO ( $50\mu M$ ) by using a quartz cuvette ( $1\times1$  cm) with septum screw cap. The cuvette was degassed *in vacuo* and backfilled with N<sub>2</sub> ( $5\times$ ) before the solvent and the catalyst solution were added *via* syringe. In presence of cesium carbonate, a distinct absorption band arises in the visible range of the spectrum (Figure S3). This process can also be followed by naked eye, as the solution turns from colorless into yellow.

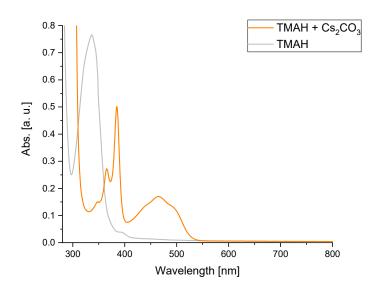


Figure S3. UV-vis spectra of (PC) in DMSO and in presence and absence of cesium carbonate.

# 2.2.2 - Emission spectra

The emission spectrum of the photocatalyst was recorded in dry, degassed DMSO in presence of cesium carbonate by using a quartz cuvette (1×1 cm) with septum screw cap. The cuvette was degassed *in vacuo* and backfilled with  $N_2$  (5×) before the solvent and the catalyst solution were added *via* syringe. The excitation wavelength was set to 420 nm (entrance-/exit slit 1 nm) and the emission was measured starting from 450 nm to 800 nm (Increment 1 nm, entrance-/exit slit 2 nm). Relative intensities are plotted for absorption and emission (Figure S4a). To determine the intersection between normalized symmetrical absorption- and emission spectra, relative intensities for the lowest energy absorption band ( $\lambda > 400$  nm) were calculated and plotted (Figure S4b).

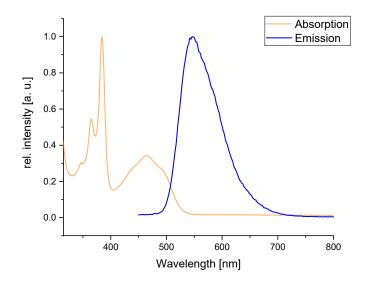


Figure S4a: Superimposed absorption and emission spectra of the photocatalyst in DMSO and in presence of cesium carbonate

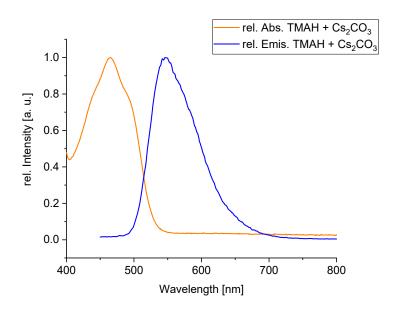
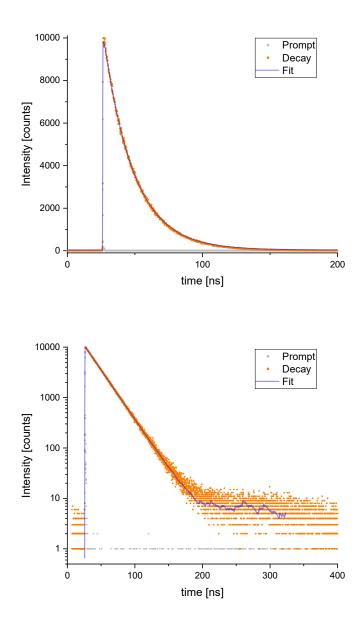


Figure S5b: Superimposed normalized absorption- and emission spectra of the photocatalyst in DMSO and in presence of cesium carbonate with an intersection at  $\lambda_{isec} = 514$  nm.

# 2.2.3 - Excited state lifetime

The luminescence lifetime of the PC was recorded in dry, degassed DMSO in presence of cesium carbonate by using a quartz cuvette ( $1\times1$  cm) with septum screw cap. The cuvette was degassed *in vacuo* and backfilled with  $N_2$  ( $5\times$ ) before the solvent and the catalyst solution were added *via* syringe. For excitation of the sample, a 452 nm laser diode was used and an optical longpass filter (cut-on wavelength

500 nm) was installed before the detection unit. The time range for the measurement was set to 400 ns. The experimental data were fitted with a mono-exponential function.

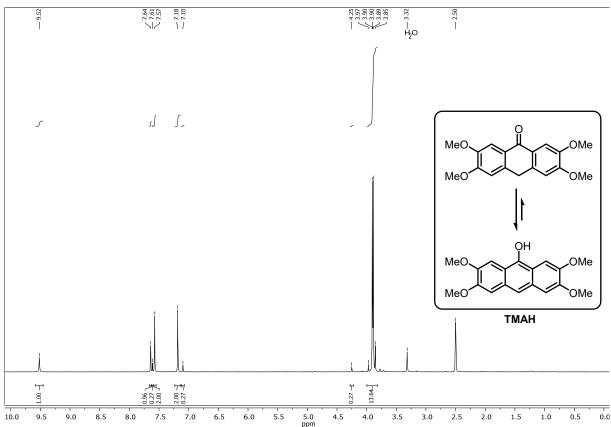


**Figure S6.** Luminescence decay of the excited photocatalyst with fit function in a linear plot (top) and logarithmic plot (bottom).

According to the parameters of the single-exponential fit function, a luminescence lifetime of 22.08 ns (CHISQ = 1.416859) was found.

# 2.2.4 - <sup>1</sup>H-NMR spectroscopy of TMAH

Proton NMR spectra of **TMAH** were recorded in absence and presence of Cs<sub>2</sub>CO<sub>3</sub> (Figure S6a-b) in dried, degassed DMSO-d6. For the measurement in presence of base, a NMR tube with septum and screw-cap was used and the spectra was recorded under N<sub>2</sub> atmosphere. Integration over the NMR signals in presence of Cs<sub>2</sub>CO<sub>3</sub> confirms the quantitative formation of the anionic species **TMA**<sup>-</sup>.



**Figure S7a.** <sup>1</sup>H-NMR of **TMAH** in DMSO-d6. The keto-enol tautomerism causes two sets of signals; The peak at 3.32 ppm is caused by residual water in the sample.

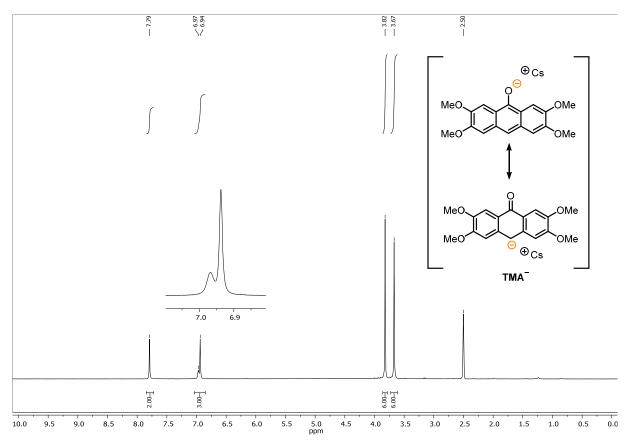


Figure S6b. <sup>1</sup>H-NMR of TMAH in presence of Cs<sub>2</sub>CO<sub>3</sub> (6 eq.) in DMSO-d6.

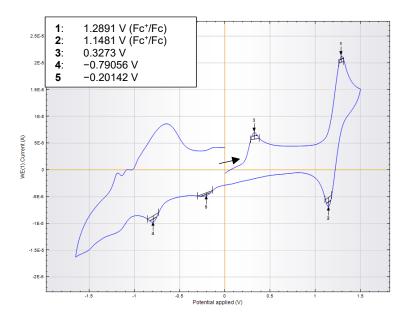
# 2.2.5 - Ground state and excited state potential

The ground state potential of the photocatalyst was investigated by cyclic voltammetry. In presence of 1,1,3,3-tetramethylguanidine the anionic photocatalyst was oxidized (Figure S7) upon sweeping to positive potentials. Obtained potentials vs. Fc<sup>+</sup>/Fc were converted to potentials against SCE.<sup>5</sup> The estimated excited state oxidation potential ( $E^*_{ox} = -2.92 \text{ V vs. SCE}$ ) of the photocatalyst was determined according to the free enthalpy change of PET (neglecting the solvent-dependent electrostatic work term) as described in literature<sup>6,7</sup> by taking the excited state energy ( $E_{0,0} = 2.41 \text{ eV}$ ,  $\lambda_{isec} = 514 \text{ nm}$ ) and the converted ground state potential ( $E_{p,ox} = -0.51 \text{ V vs. SCE}$ ) into account.

$$E_{ox}^* = E_{p,ox} - E_{0,0} + \omega$$

The obtained value for  $E^*_{ox}$  is based on following approximations: (a) As reported in literature, <sup>8,9</sup> the single electron oxidation of an organic anion causes an irreversible peak in the cyclic voltammogram and an accurate value for the ground state oxidation potential is not accessible. Thus, for the anionic photocatalyst the peak potential  $E_{p,ox}$  obtained for this irreversible process (Figure S7) was used to determine the excited state potential. (b) The excited state energy  $E_{0,0}$  can be estimated in a number of ways. When using the wavelength at the luminescence maximum  $\lambda_{emis,max}$  (546 nm) an underestimation of  $E_{0,0}$  is likely. <sup>6</sup> Furthermore, it is possible to use the midpoint between the absorption maximum of the

most red shifted absorption band and the emission maximum (506 nm). The most common way to determine  $E_{0,0}$  is by taking the intersection between symmetric normalized absorption- and emission spectra (Figure S4b,  $\lambda_{isec} = 514$  nm) which was used for the calculation herein. (c) The solvent-dependent electrostatic work term  $\omega$  contributes little to the free enthalpy change of PET when working in polar solvents like DMSO and hence was omitted in the calculation.



**Figure S7.** Cyclic voltammetry of the photocatalyst was recorded in anhydrous, degassed DMSO, in presence of 1,1,3,3-tetramethylguandine as base and ferrocene (peaks 1, 2) as internal reference.

# 3 - Substrate Synthesis

# Methyl 2-(thiophen-2-yl)acetate (3h)

To a solution of 2-thiopheneacetic acid (123 mg, 0.2 mmol) in methanol (2 mL) was added conc. H<sub>2</sub>SO<sub>4</sub> (2 drops) and the reaction was heated at reflux for 4 h. The solution was cooled, diluted with water (20 mL) and extracted with diethyl ether (3×20 mL). The combined organics were washed with brine (2×50 mL), dried via a phase separator and concentrated *in vacuo*. Purification by column chromatography on silica gel eluting with heptane:EtOAc (9:1) gave the title compound as a colorless oil (125 mg, 92%). <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.22 (dd, J = 4.9, 1.4 Hz, 1H), 6.98–6.95 (m, 2H), 3.85 (s, 2H), 3.73 (s, 3H); <sup>13</sup>C-NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  171.2, 135.3, 127.2, 127.1, 125.4, 52.6, 35.5. Data in accordance with the literature. <sup>10</sup>

# tert-Butyl (3-(benzo[b]thiophen-4-yloxy)-2-hydroxypropyl)(isopropyl)carbamate (5e)

1-(Benzo[*b*]thiophen-4-yloxy)-3-(isopropylamino)propan-2-ol (239 mg, 0.9 mmol), triethylamine (0.377 ml, 2.70 mmol), and di-*tert*-butyl dicarbonate (0.236 g, 1.08 mmol) were added to a 50 mL round bottom flask. DCM (25 mL) was added and the mixture stirred at room temperature for 2 h. Water (25 mL) was subsequently added and the layers were separated. The organic layer was washed brine (2 x 25 mL) with then dried *via* a phase separator and concentrated *in vacuo*. Purification by column chromatography on silica gel eluting with heptane:EtOAc (9:1) gave the title compound as a yellow/orange oil (151 mg, 45%).  $R_f$  0.56 [petrol–EtOAc (9:1)]; <sup>1</sup>**H-NMR** (400 MHz, DMSO) δ 7.66 (d, J = 5.4 Hz, 1H), 7.53 (dd, J = 10.5, 2.3 Hz, 2H), 7.30 (t, J = 8.0 Hz, 1H), 6.84 (d, J = 7.9 Hz, 1H), 5.76 (s, 1H), 5.17 (d, J = 5.2 Hz, 1H), 4.23 – 3.89 (m, 4H), 3.54 – 3.22 (m, 1H), 1.38 (s, 9H), 1.24 – 1.01 (m, 6H). <sup>13</sup>C-NMR (125 MHz, DMSO) δ 156.4, 155.2, 141.5, 131.1, 130.0, 128.9, 123.2, 121.1, 109.1, 80.3, 71.5, 68.2, 49.0, 45.5, 28.3, 19.5.

# tert-Butyl 2-phenyl-1H-indole-1-carboxylate (3v)

The compound was synthesized according to a literature known procedure. <sup>11</sup> In a flame dried 100 mL Schlenk flask under N<sub>2</sub> atmosphere equipped with stirring bar, (Boc)<sub>2</sub>O (1.20 g, 5.50 mmol, 1.1 equiv.) was added to a solution of 2-phenylindole (0.966 g, 5.0 mmol, 1 equiv.) and 4-(*N*,*N*-dimethylamino)pyridine in dry MeCN (30 mL). The resulting mixture was stirred at room temperature for 24 h and was then concentrated *in vacuo*. After the addition of water, the mixture was extracted with EtOAc (3×). The combined organic layers were washed with brine, dried over Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated under reduced pressure. The crude material was purified by flash silica gel column chromatography using a mixture of hexanes/EtOAc to provide the title compound as white solid (1.40 g, 4.8 mmol, 96%). <sup>1</sup>H-NMR (400 MHz, Chloroform-*d*)  $\delta$  8.27 – 8.20 (m, 1H), 7.59 – 7.54 (m, 1H), 7.46 – 7.31 (m, 6H), 7.27 (td, J = 7.6, 0.9 Hz, 1H), 6.57 (s, 1H), 1.32 (s, 9H). <sup>13</sup>C-NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  150.3 (C<sub>q</sub>), 140.6 (C<sub>q</sub>), 137.6 (C<sub>q</sub>), 135.1 (C<sub>q</sub>), 129.3 (C<sub>q</sub>), 128.9, 127.9, 127.7, 124.4, 123.0, 120.6, 115.3, 110.0, 83.5 (C<sub>q</sub>), 27.7.

# Thieno[2,3-d|pyrimidine (3y)

Based on a literature reported procedure for dehalogenation of aromatic compounds<sup>12</sup>, a 5 mL crimp vial equipped with stirring bar was charged with 4-chlorothieno[2,3-d]pyrimidine (17.1 mg, 0.1 mmol, 1 equiv.) and 10-phenylphenothiazine (2.8 mg, 0.01 mmol, 10 mol%) and sealed with an aluminium crimp seal with septum. The vial was degassed and flushed with N<sub>2</sub> and tributylamine (119 μL, 0.5 mmol, 5 equiv.), formic acid (18.9 μL, 0.5 mmol, 5 equiv.) and dry MeCN (1 mL) were added. The reaction mixture was degassed by freeze-pump-thaw cycles (3×) and backfilled with N<sub>2</sub>. The crimp vial was irradiated from the bottom side with 365 nm LED light for 22 hrs and a constant reaction temperature (25°C) was maintained by employing a water-cooling circuit connected to a thermostat. For isolation of the compound, 10 reactions were combined. The reactions were quenched by adding water and brine and the resulting mixture was extracted with EtOAc (3×). The combined organic layers were dried over Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated *in vacuo*. Purification was accomplished by flash silica gel chromatography using a mixture of hexanes/EtOAc as eluents and subsequent recrystallization from hexanes to afford the title compound as pale-yellow needles (31.8 mg, 0.23 mmol, 23%). <sup>1</sup>H-NMR (400

MHz, Chloroform-d)  $\delta$  9.16 (s, 1H), 9.10 (s, 1H), 7.57 (d, J = 6.0 Hz, 1H), 7.36 (d, J = 6.0 Hz, 1H). Data in accordance with the literature.<sup>13</sup>

# 2-(Thiophen-3-yl)pyridine (3z)

Following a literature known procedure<sup>14</sup> a 10 mL crimp vial was charged with 3-thienylboronic acid (130 mg, 1.01 mmol, 1.2 equiv.),  $K_2CO_3$  (326 mg, 2.36 mmol, 2.8 equiv.),  $[Pd(PPh_3)_2Cl_2]$  (29.6 mg, 0.042 mmol, 0.05 equiv.), DME (2.5 mL) and water (1.17 mL). The vial was sealed with an aluminium crimp seal with septum and argon was bubbled through the solution for 10 minutes. Bromopyridine (81  $\mu$ L, 0.842 mmol, 1 equiv.) was added *via* syringe and the reaction was stirred in a pre-heated heating block for 18 hrs at 80 °C. The reaction was allowed to cool to ambient temperature, was quenched by adding water and was extracted with EtOAc (3×). The combined organic layers were washed with brine, dried over Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated *in vacuo*. The crude product was purified by flash silica gel column chromatography using a mixture of hexanes/EtOAc and was obtained as colorless oil (127 mg, 0.79 mmol, 94%). <sup>1</sup>H-NMR (400 MHz, Chloroform-*d*)  $\delta$  8.62 (ddd, J = 4.9, 1.8, 0.9 Hz, 1H), 7.90 (dd, J = 3.0, 1.3 Hz, 1H), 7.70 (td, J = 7.7, 1.8 Hz, 1H), 7.66 (dd, J = 5.0, 1.3 Hz, 1H), 7.62 (dt, J = 8.0, 1.1 Hz, 1H), 7.40 (dd, J = 5.0, 3.0 Hz, 1H), 7.17 (ddd, J = 7.4, 4.9, 1.2 Hz, 1H). <sup>13</sup>C-NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  153.7 ( $C_q$ ), 149.8, 142.3 ( $C_q$ ), 136.8, 126.4, 126.3, 123.6, 121.9, 120.4.

# *N*,*N*-Dimethyl-4-vinylaniline (7g)

According to a literature known procedure<sup>15</sup> a flame-dried 250 mL Schlenk flask was charged under N<sub>2</sub> atmosphere with methyltriphenylphosphonium bromide (14.4 g, 40.3 mmol, 1 equiv.) and dry THF (60 mL). The suspension was cooled to 0 °C and *n*-BuLi (1.6M in hexane, 25.2 mL, 40.3 mmol, 1 equiv.) was slowly added *via* syringe and the resulting mixture was stirred for 1 h. A solution of 4-(*N*,*N*-dimethylamino)benzaldehyde (6.02 g, 40.3 mmol, 1 equiv.) in dry THF (20 mL) was added dropwise and the reaction was further stirred at 0 °C for 1 h and at ambient temperature for 18 hrs. The reaction was quenched by adding sat. aq. NH<sub>4</sub>Cl (30 mL) and the resulting mixture was extracted with DCM (3×20 mL) and the combined organic layers were dried over Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated *in vacuo*. Purification by vacuum distillation (0.8 mbar, 75 °C) afforded the title compound as yellowish oil

(4.70 g, 31.9 mmol, 79%). <sup>1</sup>**H-NMR** (300 MHz, Chloroform-*d*)  $\delta$  7.37 – 7.27 (m, 2H), 6.75 – 6.58 (m, 3H), 5.55 (dd, J = 17.6, 1.1 Hz, 1H), 5.03 (dd, J = 10.9, 1.1 Hz, 1H), 2.97 (s, 6H). <sup>13</sup>**C-NMR** (75 MHz, CDCl<sub>3</sub>)  $\delta$  150.4 (C<sub>q</sub>), 136.7, 127.3, 126.3 (C<sub>q</sub>), 112.5, 109.5 (CH<sub>2</sub>), 40.7.

# Methyl(4-vinylphenyl)sulfane (7h)

According to a literature known procedure<sup>15</sup> a flame-dried 250 mL Schlenk flask was charged under  $N_2$  atmosphere with methyltriphenylphosphonium bromide (14.1 g, 39.5 mmol, 1 equiv.) and dry THF (60 mL). The suspension was cooled to 0 °C and n-BuLi (1.6M in hexane, 30 mL, 48 mmol, 1.2 equiv.) was slowly added via syringe and the resulting mixture was stirred for 1 h. 4-(Methylthio)benzaldehyde (5.24 mL, 38.4 mmol, 1 equiv.) was added dropwise via syringe and the reaction mixture was stirred for further 2.5 h at 0 °C. After dilution with THF (20 mL) the reaction was stirred at ambient temperature overnight and was quenched by adding sat. aq. NH<sub>4</sub>Cl (20 mL). The crude mixture was extracted with DCM (3×20 mL) and the combined organic layers were dried over Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated in vacuo. Purification by flash silica gel chromatography (hexanes/MTBE) afforded the title compound as colorless liquid (4.04 g, 26.9 mmol, 68%).  $^{1}$ H-NMR (400 MHz, Chloroform-d)  $\delta$  7.37 – 7.32 (m, 2H), 7.25 – 7.20 (m, 2H), 6.69 (dd, J = 17.6, 10.9 Hz, 1H), 5.73 (dt, J = 17.6, 0.9 Hz, 1H), 5.23 (dt, J = 10.8, 0.9 Hz, 1H), 2.50 (s, 3H).  $^{13}$ C-NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  138.1 (C<sub>q</sub>), 136.3, 134.6 (C<sub>q</sub>), 126.7, 126.7, 113.3 (CH<sub>2</sub>), 15.9.

# 4 - Carboxylation Reactions

# 4.1 - Reaction Optimization

# 4.1.1 - General Procedure for the Reaction Optimization – GP1

To a dry flat-bottomed crimp vial (5 mL) equipped with stirring bar, was added acenaphthene (1a, 0.1-0.2 mmol, 1 equiv.) and photocatalyst (5-20 mol%, Scheme S2). Base (if solid) was quickly added and the vial was sealed with a Supelco aluminium crimp seal with septum (PTFE/butyl). The vial was then evacuated and refilled with CO<sub>2</sub> (5×) *via* syringe needle. The reaction mixture was dissolved in the solvent (dry and degassed by bubbling with N<sub>2</sub>) and base (if liquid) was added *via* syringe. The vial was sealed with two layers of Parafilm® and then had gaseous CO<sub>2</sub> added *via* a Luer Lock Monoject<sup>TM</sup> (20 ccm) syringe, into the head space. The vial was then stirred and irradiated from the bottom side and a constant reaction temperature (0 °C or 25 °C) was maintained by employing a cooling circuit connected to a thermostat. After 18 hrs the reaction was transferred with aq. NaOH (0.1M) into a centrifuge tube and was washed with Et<sub>2</sub>O (2×) to remove left over starting material or non-polar side products. The aqueous layer was acidified by adding aq. HCl (2M) and was extracted with EtOAc (3×). The combined organic layers were dried over Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated *in vacuo*. As internal standard a stock solution of 1,3,5-trimethoxybenzene in DMSO-d6 (0.7 mL, 42.9 mM) was added and the mixture was analyzed by <sup>1</sup>H-NMR spectroscopy. The <sup>1</sup>H-NMR yield was determined by integration over product signals and internal standard signals.

**Scheme S2.** Tested 9-anthrone based derivatives for the optimization of the carboxylation reaction; 9-anthrone (**Ant**), 2,6-dimethoxyanthracen-9(10*H*)-one (**DMA**), 2,3,6,7-tetramethoxyanthracen-9(10*H*)-one (**TMAH**), bianthronyl (**BA**), 10-benzhydryl-anthrone (**BHA**).

Table S1. Optimization of the photocatalyzed aromatic C-H carboxylation.

entry	PC (mol%)	base (eq.)	solvent	λ [nm]	V(CO <sub>2</sub> ) added [ccm]	NMR yield [%]
1	Ant (20)	Cs <sub>2</sub> CO <sub>3</sub> (2)	DMSO	455	22	n.d.a
2	<b>DMA</b> (20)	$Cs_2CO_3(2)$	DMSO	455	22	n.d.
3	TMA (20)	$Cs_2CO_3(2)$	DMSO	455	22	$37^{b}$
4	<b>BA</b> (20)	$Cs_2CO_3(2)$	DMSO	455	22	trace
5	BHA (20)	$Cs_2CO_3(2)$	DMSO	455	22	n.d.
6	TMAH (20)	$Cs_2CO_3(1)$	DMSO	455	22	1
7	TMAH (20)	$Cs_2CO_3(3)$	DMSO	455	22	$68^{b}$
8	TMAH (20)	$Cs_2CO_3(4)$	DMSO	455	22	59
9	<b>TMAH</b> (5)	$Cs_2CO_3(2)$	DMSO	455	22	54
10	TMAH (10)	$Cs_2CO_3(2)$	DMSO	455	22	60
11	TMAH (10)	$Cs_2CO_3(3)$	DMSO	455	22	56
$12^c$	TMAH (20)	$Cs_2CO_3(3)$	DMSO	455	22	52
13	TMAH (20)	$Na_2CO_3(2)$	DMSO	455	22	1
14	TMAH (20)	$K_2CO_3(2)$	DMSO	455	22	17
15	TMAH (10)	$K_2CO_3(3)$	DMSO	455	22	25
$16^{c}$	TMAH (10)	$K_2CO_3(3)$	DMSO	455	22	47
$17^c$	TMAH (20)	$K_2CO_3(3)$	DMSO	455	22	38
18	TMAH (20)	$(NH_4)_2CO_3(3)$	DMSO	455	22	2
19	TMAH (20)	Cs pivalate (3)	DMSO	455	22	6
20	TMAH (20)	$K_3PO_4(3)$	DMSO	455	22	trace
21	TMAH (20)	$(NBu_4)H_2PO_4(3)$	DMSO	455	22	7
22	TMAH (20)	DBU (3)	DMSO	455	22	28
23	TMAH (20)	TMG (3)	DMSO	455	22	22
24	TMAH (20)	BTMG (3)	DMSO	455	22	25
25	TMAH (20)	$Cs_2CO_3(3)$	DMF	455	22	35
26	TMAH (20)	$Cs_2CO_3(3)$	DMA	455	22	12
27	TMAH (20)	$Cs_2CO_3(3)$	NMP	455	22	14
28	TMAH (20)	$Cs_2CO_3(3)$	$^{i}$ PrOH	455	22	n.d.
29	TMAH (20)	$Cs_2CO_3(3)$	DCM	455	22	n.d.
30	TMAH (20)	$Cs_2CO_3(3)$	acetone	455	22	trace
31	TMAH (20)	$Cs_2CO_3(3)$	DMSO	400	22	14
32	TMAH (20)	$Cs_2CO_3(3)$	DMSO	535	22	29
33	TMAH (20)	$Cs_2CO_3(3)$	DMSO	$white^d$	22	27
34	TMAH (20)	$Cs_2CO_3(3)$	DMSO	455e	22	23
35	TMAH (20)	$Cs_2CO_3(3)$	DMSO	$455^{f}$	22	56
$36^g$	TMAH (20)	$Cs_2CO_3(3)$	DMSO/DMF (1:1)	455	22	17
37	TMAH (20)	$Cs_2CO_3(3)$	DMSO	455	- h	37
38	TMAH (20)	$Cs_2CO_3(3)$	DMSO	455	11	48
39 <sup>i</sup>	TMAH (20)	$Cs_2CO_3(3)$	DMSO	455	22	39
$40^{j}$	TMAH (20)	$Cs_2CO_3(3)$	DMSO	455	22	51
$41^k$	TMAH (20)	$Cs_2CO_3(3)$	DMSO	455	22	13
42 <sup>l</sup>	TMAH (20)	$Cs_2CO_3(3)$	DMSO	455	22	34
43 <sup>m</sup>	TMAH (20)	$Cs_2CO_3(3)$	DMSO	455	22	49
$44^{n}$	TMAH (20)	$Cs_2CO_3(3)$	DMSO	455	22	57
45°	TMAH (20)	$Cs_2CO_3(3)$	DMSO	455	22	52

All reactions, if not otherwise stated, were run following GP1: <sup>a</sup> product was not detected; <sup>b</sup> isolated yield following GP2a; <sup>c</sup> reaction was run with 18-crown-6 (1 equiv.); <sup>d</sup> cold white LED; <sup>e</sup> fan-cooled high-power LED setup (7 W) was used. Due to inefficient cooling, the reaction temperature was significantly elevated; <sup>f</sup> water-cooled high-power LED setup (1.4 W) was used. DMSO was saturated with CO<sub>2</sub> by bubbling gas through the solvent; <sup>g</sup> reaction was run at 0 °C; <sup>h</sup> reaction was run without pressure (1 atm) of CO<sub>2</sub>; <sup>i</sup> reaction was run with 0.2 mmol of substrate; <sup>j</sup> concentration was changed to 0.05 M (0.1 mmol substrate in 2 mL DMSO); <sup>k</sup> reaction with chloro(pyridine)bis(dimethylglyoximato)cobalt(III) (5 mol%); <sup>l</sup> reaction with p-terphenyl (5 mol%); <sup>m</sup> reaction with p-quaterphenyl (5 mol%); <sup>n</sup> reaction with (iPr)<sub>3</sub>SiSH (10 mol%); <sup>e</sup> reaction with 1,4-cyclohexadiene (20 mol%).

# 4.1.2 - Kinetic profile of the aromatic carboxylation reaction

Under the optimized conditions the kinetic profile of the reaction was monitored within the first six hours of the reaction (Figure S8). Reactions were run following GP1.

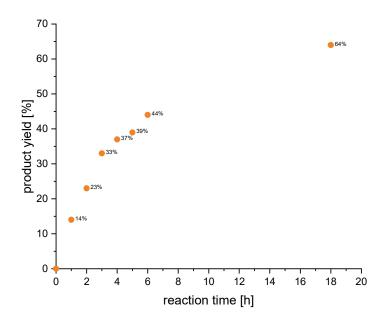


Figure S8. The progress of the carboxylation reaction was monitored within the first six hours.

#### 4.1.3 - Control reactions

Conducted control experiments revealed that all compounds and light are crucial for product formation (Table S2). To exclude a base promoted carboxylation reaction, substrates 3c & 3j possessing acidic C-H bonds were tested in absence of catalyst and light and upon work-up, the respective products 4c & 4j could not be detected. Substrates 3b and 3p gave excellent yields of the respective carboxylation products following GP2a. No product was formed in absence of light and catalyst, demonstrating the photocatalytic nature of this reaction.

Table S2. Control reactions

entry	PC (mol%)	Substrate	base (eq.)	λ [nm]	V(CO <sub>2</sub> ) added [ccm]	Product	NMR yield [%]
1	-	1a	$Cs_2CO_3(3)$	455	22	2a	n.d.a
2	TMAH (20)	1a	-	455	22	2a	n.d.
3	TMAH (20)	1a	$Cs_2CO_3(3)$	455	no CO <sub>2</sub> <sup>b</sup>	2a	n.d.
4	TMAH (20)	1a	$Cs_2CO_3(3)$	$\operatorname{dark}^c$	22	2a	n.d.
5	-	$3\mathbf{b}^d$	$Cs_2CO_3(3)$	dark	22	4b	n.d.
6	-	$3c^e$	$Cs_2CO_3(3)$	dark	22	4c	n.d.
7	-	$3\mathbf{j}^f$	$Cs_2CO_3(3)$	dark	22	4j	n.d.
8	-	$3p^g$	$Cs_2CO_3(3)$	dark	22	4p	n.d.

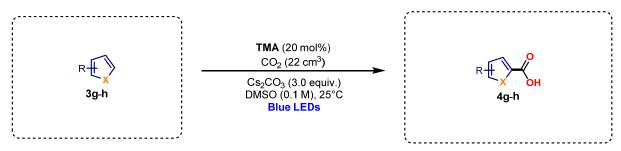
All reactions, if not otherwise stated, were run following GP1: "product was not detected; "reaction was run under  $N_2$  atmosphere; "reaction was stirred in the dark; "reaction was run following GP1 using **3b** (0.1 mmol) as substrate; "reaction was run following GP1 using **3c** (0.1 mmol) as substrate; "reaction was run following GP1 using **3j** (0.1 mmol) as substrate; "reaction was run following GP1 using **3p** (0.1 mmol) as substrate.

# 4.2 - Substrate Scope

#### General Procedure for Carboxylation of Hetero(arenes) and Styrenes-GP2a

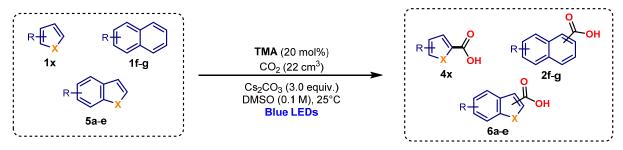
To a dry flat-bottomed crimp vial (5 mL) equipped with stirring bar, was added the arene (if solid) (0.1 mmol) and 2,3,6,7-tetramethoxyanthracen-9(10*H*)-one (6.3 mg, 0.02 mmol, 20 mol%). Cs<sub>2</sub>CO<sub>3</sub> (98 mg, 3 equiv.) was quickly added and the vial was sealed with a Supelco aluminium crimp seal with septum (PTFE/butyl). The vial was then evacuated and refilled with CO<sub>2</sub> (5×) *via* syringe needle. The reaction mixture was dissolved in DMSO (1 mL, dry and degassed by bubbling with N<sub>2</sub>) and the arene (0.1 mmol) (if liquid) was added *via* syringe. The vial was sealed with two layers of Parafilm® and then had gaseous CO<sub>2</sub> added *via* a Luer Lock Monoject<sup>TM</sup> (20 ccm) syringe, into the head space. The vial was then irradiated from the bottom side with blue LED light and a constant reaction temperature (25°C) was maintained by employing a water-cooling circuit connected to a thermostat. After 18 hrs of reaction time the pressure was released. For product isolation, the reaction mixtures of 4 reactions run in parallel were combined and transferred with water and Et<sub>2</sub>O into a separating funnel. The ether layer was extracted with water (3×) and the combined aqueous layers were acidified with aq. HCl (2M) to adjust to an acidic pH. The aqueous layer was extracted with EtOAc (3×) and the combined EtOAc layers were dried over Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated *in vacuo*. The crude material was purified by silica flash column chromatography using mixtures of hexanes and ethyl acetate with 0.5% HOAc (v/v) as eluents.

# General Procedure for Carboxylation of Arenes – GP2b (no glovebox)



To a dry flat-bottomed crimp vial (5 mL) equipped with stirring bar, was added the arene (if solid) (0.1 mmol) and 2,3,6,7-tetramethoxyanthracen-9(10*H*)-one (6.3 mg, 0.02 mmol, 20 mol%). Cs<sub>2</sub>CO<sub>3</sub> (98 mg, 3 equiv.) was quickly added and the vial was sealed with a Supelco aluminium crimp seal with septum (PTFE/butyl). The vial was then evacuated and refilled with N<sub>2</sub> (3×) *via* syringe needle. The reaction mixture was dissolved in DMSO (1 mL, dry and degassed by bubbling with N<sub>2</sub>) and the arene (0.1 mmol) (if liquid) was added *via* syringe. The reaction mixture then had gaseous CO<sub>2</sub> (22 cm<sup>3</sup>) added *via* a gastight Hamilton® syringe, into the head space of the vial. The vial was then irradiated from the bottom side with blue light and a constant reaction temperature (25°C) was maintained by employing a water-cooling circuit connected to a thermostat. After the designated time the reactions were extracted *via* an acid base wash. The combined organic layers were dried over Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated *in vacuo*.

# General Procedure for Carboxylation of Arenes – GP2c (glovebox)



To a dry microwave vial (5 mL) equipped with stirring bar, was added the arene (if solid) (0.1 mmol) and 2,3,6,7-tetramethoxyanthracen-9(10H)-one (6.3 mg, 0.02 mmol, 20 mol%). The vials were sealed and transferred to a glovebox after the vial had evacuated and refilled with  $N_2$  (×3) within the antechamber via syringe.  $Cs_2CO_3$  (98 mg, 3 equiv.) and DMSO (1 mL) was added and the vial was sealed with a Supelco aluminium crimp seal with septum (PTFE/butyl). The vials were removed from the glovebox and the arene (0.1 mmol) (if liquid) was added via syringe. The reaction mixture then had gaseous  $CO_2(22 \text{ cm}^3)$  added via a gastight Hamilton® syringe, into the head space of the vial. The vial was then irradiated from two sides by two kessil lamps (vials approximately 6 cm away from the light source) with fans placed in front of the vials for cooling (temperatures measured via IR ranged between 25-30 °C). After 18 hrs. the irradiation was stopped and the vials were decapped quenched with aq. HCl (1 mL, 0.3M) and monitored by LCMS and <sup>1</sup>H NMR. Reactions were filtered of any solids and purified directly *via* prep-HPLC.

# General Procedure for the substitution reaction of benzo[b]thiophene with ketones-GP3

To a dry flat-bottomed crimp vial (5 mL) equipped with stirring bar, was added the benzo[b]thiophene (13.4 mg, 0.1 mmol, 1 equiv.) and 2,3,6,7-tetramethoxyanthracen-9(10H)-one (6.3 mg, 0.02 mmol, 20 mol%). Cs<sub>2</sub>CO<sub>3</sub> (98 mg, 0.3 mmol, 3 equiv.) was quickly added and the vial was sealed with a Supelco aluminium crimp seal with septum (PTFE/butyl). The vial was then evacuated and refilled with N<sub>2</sub> (5×) via syringe needle. The reaction mixture was dissolved in DMSO (1mL, dry and degassed by bubbling with N<sub>2</sub>) and the ketone (1.0 mmol, 10 equiv.) was added via syringe. The vial was then irradiated from the bottom side with blue LED light and a constant reaction temperature (25°C) was maintained by employing a water-cooling circuit connected to a thermostat. After 18 hrs the reaction was quenched by adding water. For product isolation, the reaction mixtures of 4 reactions run in parallel were combined and transferred with water and EtOAc into a separating funnel. The reaction mixture was extracted with EtOAc (3×) and the combined organic layers were dried over Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated *in vacuo*. The crude material was purified by silica flash column chromatography using mixtures of hexanes and ethyl acetate as eluents.

#### 1,2-Dihydroacenaphthylene-5-carboxylic acid (2a)

Following GP2a, acenaphthene (0.4 mmol) gave **2a** (68%) as a pale orange solid; <sup>1</sup>**H-NMR** (400 MHz, Chloroform-d)  $\delta$  8.74 (d, J = 8.5 Hz, 1H), 8.45 (d, J = 7.3 Hz, 1H), 7.63 (dd, J = 8.6, 6.9 Hz, 1H), 7.37 (dd, J = 12.2, 7.1 Hz, 2H), 3.45 (s, 4H). <sup>1</sup>**H-NMR** (400 MHz, DMSO- $d_6$ )  $\delta$  12.85 (bs, 1H), 8.60 (d, J = 8.5 Hz, 1H), 8.22 (d, J = 7.3 Hz, 1H), 7.57 (dd, J = 8.6, 6.9 Hz, 1H), 7.36 (t, J = 6.6 Hz, 2H), 3.35 (s, 4H). <sup>13</sup>**C-NMR** (101 MHz, DMSO)  $\delta$  168.3 (C $_q$ ), 152.5 (C $_q$ ), 146.2 (C $_q$ ), 139.1 (C $_q$ ), 132.9, 129.8 (C $_q$ ), 129.6, 122.3 (C $_q$ ), 121.6, 119.9, 118.6, 29.9. **HRMS** (EI+): calculated m/z for C<sub>13</sub>H<sub>10</sub>O<sub>2</sub> [M<sup>++</sup>] 198.06753; found 198.06722. Data in accordance with the literature. <sup>16</sup>

#### 1-Naphthoic acid (2b)

Following GP2a, naphthalene (0.3 mmol) gave **2b** (38%) as a white solid; <sup>1</sup>**H-NMR** (400 MHz, DMSO- $d_6$ )  $\delta$  13.13 (bs, 1H), 8.86 (d, J = 8.5 Hz, 1H), 8.20 – 8.10 (m, 2H), 8.02 (d, J = 7.7 Hz, 1H), 7.70 – 7.52 (m, 3H). <sup>13</sup>C-NMR (101 MHz, DMSO)  $\delta$  168.6 (C<sub>q</sub>), 133.5 (C<sub>q</sub>), 132.9, 130.7 (C<sub>q</sub>), 129.8, 128.6, 127.7 (C<sub>q</sub>), 127.5, 126.2, 125.5, 124.9. **HRMS** (EI+): calculated m/z for C<sub>11</sub>H<sub>8</sub>O<sub>2</sub> [M<sup>++</sup>] 172.05188; found 172.05143. Data in accordance with the literature. <sup>17</sup>

#### 5-Methoxy-1-naphthoic acid (2c)

Following GP2a, 1-methoxynaphthalene (0.4 mmol) gave **2c** (22%) as a pale yellow solid; <sup>1</sup>**H-NMR** (400 MHz, DMSO- $d_6$ )  $\delta$  12.95 (bs, 1H), 8.45 – 8.35 (m, 2H), 8.13 (dd, J = 7.2, 1.3 Hz, 1H), 7.60 – 7.51 (m, 2H), 7.05 (d, J = 7.8 Hz, 1H), 3.99 (s, 3H). <sup>13</sup>**C-NMR** (101 MHz, DMSO)  $\delta$  168.8 (C<sub>q</sub>), 154.9 (C<sub>q</sub>),

131.6 ( $C_q$ ), 130.0, 127.8, 127.7 ( $C_q$ ), 126.1, 125.3 ( $C_q$ ), 124.2, 117.5, 104.7, 55.7. **HRMS** (EI+): calculated m/z for  $C_{12}H_{10}O_3$  [ $M^{++}$ ] 202.06245; found 202.06283. Data in accordance with the literature.<sup>18</sup>

#### 5-(Dimethylamino)-1-naphthoic acid (2d)

Following GP2a, 1-dimethylaminonaphthalen (0.4 mmol) gave **2d** (38%) as a yellow solid; <sup>1</sup>**H-NMR** (400 MHz, DMSO- $d_6$ )  $\delta$  13.04 (bs, 1H), 8.43 (dd, J = 15.9, 8.6 Hz, 2H), 8.08 (dd, J = 7.1, 1.0 Hz, 1H), 7.54 (ddd, J = 18.0, 8.6, 7.3 Hz, 2H), 7.19 (d, J = 7.3 Hz, 1H), 2.82 (s, 6H). <sup>13</sup>**C-NMR** (101 MHz, DMSO)  $\delta$  168.9 (C<sub>q</sub>), 151.0 (C<sub>q</sub>), 132.0 (C<sub>q</sub>), 129.3, 128.6 (C<sub>q</sub>), 128.5 (C<sub>q</sub>), 128.4, 127.4, 124.0, 120.0, 114.4, 45.0. **HRMS** (ESI+): calculated m/z for C<sub>13</sub>H<sub>14</sub>NO<sub>2</sub> [(M+H)<sup>+</sup>] 216.1019; found 216.1022.

#### 4-Methyl-1-naphtoic acid (2e) & 5-Methyl-1-naphtoic acid (2e')

Following GP2a, 1-methylnaphthalene (0.1 mmol) gave **2e** (19%) and **2e'** (14%) as a beige solid. **2e:2e'** = 1.3:1. The ratio of products was determined by  ${}^{1}$ H-NMR analysis. **HRMS** (EI+): calculated m/z for  $C_{12}H_{10}O_{2}$  [M<sup>++</sup>] 186.06753; found 186.06731.

Data for **2e**: <sup>1</sup>**H-NMR** (300 MHz, Chloroform-d)  $\delta$  9.22 – 9.13 (m, 1H), 8.34 (d, J = 7.5 Hz, 1H), 8.13 – 8.05 (m, 1H), 7.73 – 7.50 (m, 2H), 7.41 (d, J = 7.5 Hz, 1H), 2.78 (s, 3H). <sup>1</sup>**H-NMR** (400 MHz, DMSO- $d_6$ )  $\delta$  13.06 (bs, 1H), 9.00 – 8.88 (m, 1H), 8.11 (m, 1H), 8.06 (d, J = 7.4 Hz, 1H), 7.68 – 7.58 (m, 2H), 7.44 (d, J = 7.0 Hz, 1H), 2.70 (s, 3H). <sup>13</sup>C-NMR (101 MHz, DMSO)  $\delta$  168.7, 139.7, 132.4, 130.9, 129.8, 127.2, 126.1, 126.0, 125.9, 125.7, 124.7, 19.6. Data in accordance with literature. <sup>19,20</sup>

Data for **2e'**: <sup>1</sup>**H-NMR** (300 MHz, CD<sub>3</sub>OD)  $\delta$  8.95 (d, J = 8.8 Hz, 1H), 8.40 (dd, J = 7.3, 1.3 Hz, 1H), 8.30 (dt, J = 8.5, 1.1 Hz, 1H), 7.71 – 7.51 (m, 2H), 7.41 (d, J = 7.5 Hz, 1H), 2.75 (s, 3H). <sup>1</sup>**H-NMR** (400 MHz, DMSO- $d_6$ )  $\delta$  13.06 (bs, 1H), 8.67 (d, J = 8.6 Hz, 1H), 8.25 (d, J = 8.5 Hz, 1H), 8.14 – 8.08 (m, 1H), 7.68 – 7.58 (m, 1H), 7.54 – 7.48 (m, 1H), 7.39 – 7.47 (m, 1H), 2.68 (s, 3H). <sup>13</sup>**C-NMR** (101 MHz,

DMSO)  $\delta$  169.0, 134.5, 132.5, 130.8, 129.1, 128.7, 128.6, 127.1, 126.9, 124.8, 123.7, 19.5. Data in accordance with the literature.<sup>21</sup>

#### 1-Hydroxy-4-naphthoic acid (2f) & 1-hydroxy-2-naphthoic acid (2f')

Following GP2c, 1-naphthol (0.1 mmol) gave **2f & 2f'** (56%) as an inseparable mixture (1:1), as a waxy solid.  ${}^{1}$ H-NMR (500 MHz, DMSO)  $\delta$  12.58 (s, 2H), 11.02 (s, 2H), 9.03 (d, J = 8.7 Hz, 1H), 8.23 (d, J = 7.7 Hz, 1H), 8.13 (d, J = 8.1 Hz, 1H), 7.87 (dd, J = 8.3, 1.0 Hz, 1H), 7.61 (ddd, J = 8.5, 6.8, 1.4 Hz, 1H), 7.50 (ddd, J = 8.2, 6.8, 1.2 Hz, 1H), 7.48 – 7.42 (m, 1H), 7.41 – 7.32 (m, 3H), 6.91 (d, J = 8.2 Hz, 1H), 6.87 (dd, J = 7.2, 1.3 Hz, 1H).  ${}^{13}$ C-NMR (126 MHz, DMSO)  $\delta$  171.88, 168.30, 157.80, 134.78, 133.04, 132.85, 128.74, 127.86, 126.98, 125.54, 125.08, 124.75, 124.56, 123.59, 122.43, 118.63, 116.69, 109.60, 106.95, 104.40. Data in accordance with the literature.  ${}^{18,22}$ 

# 2,7-dihydroxy-1-naphthoic acid (2g)

Following GP2c, naphthalene-2,7-diol (0.1 mmol) gave **2g** (64%) as a yellow solid. <sup>1</sup>**H-NMR** (500 MHz, DMSO-d6)  $\delta$  12.43 (bs, 1H), 9.82 (s, 2H), 7.95 (s, 1H), 7.84 (d, J = 8.8 Hz, 1H), 7.68 (d, J = 8.8 Hz, 1H), 6.95-6.77 (m, 2H); <sup>13</sup>**C-NMR** (126 MHz, DMSO-d6)  $\delta$  173.0, 161.8, 157.5, 135.1, 133.6, 130.5, 122.5, 115.1, 115.1 107.5, 105.8; **HRMS** (ESI+): calculated m/z for C<sub>11</sub>H<sub>9</sub>O<sub>4</sub> [(M+H)<sup>+</sup>] 205.0495; found 205.0497.

#### 5-(methoxycarbonyl)thiophene-2-carboxylic acid (4a)

Following GP2a, methyl thiophene-2-carboxylate (0.4 mmol) gave **4a** (49%) as a white solid;  ${}^{1}$ **H-NMR** (400 MHz, DMSO-d6)  $\delta$  13.65 (bs, 1H), 7.78 (d, J = 3.9 Hz, 1H), 7.72 (d, J = 3.9 Hz, 1H), 3.85 (s, 3H).

<sup>13</sup>C-NMR (101 MHz, DMSO) δ 162.3 ( $C_q$ ), 161.4 ( $C_q$ ), 140.4 ( $C_q$ ), 137.5 ( $C_q$ ), 133.7, 133.2, 52.7. **HRMS** (EI+): calculated m/z for  $C_7H_6O_4S$  [ $M^{++}$ ] 185.99813; found 185.99833. Data in accordance with the literature.<sup>23</sup>

# 5-Cyanothiophene-2-carboxylic acid (4b)

Following GP2a, thiophene-2-carbonitrile (0.4 mmol) gave **4b** (92%) as a pale yellow solid; <sup>1</sup>**H-NMR** (400 MHz, DMSO- $d_6$ )  $\delta$  13.98 (bs, 1H), 8.00 (d, J = 4.0 Hz, 1H), 7.79 (d, J = 4.0 Hz, 1H). <sup>13</sup>**C-NMR** (101 MHz, DMSO)  $\delta$  161.5 (C<sub>q</sub>), 141.7 (C<sub>q</sub>), 139.5 (C<sub>q</sub>), 132.9 (C<sub>q</sub>), 113.6, 113.3. **HRMS** (EI+): calculated m/z for C<sub>6</sub>H<sub>3</sub>NO<sub>2</sub>S [M<sup>++</sup>] 152.98790; found 152.98804. Data in accordance with the literature.<sup>24</sup>

#### 5-Acetylthiophene-2-carboxylic acid (4c)

Following GP2a, 2-acetylthiophene (0.4 mmol) gave **4c** (39%) as a pale yellow solid; <sup>1</sup>**H-NMR** (300 MHz, DMSO- $d_6$ )  $\delta$  13.55 (bs, 1H), 7.92 (d, J = 4.0 Hz, 1H), 7.76 (d, J = 3.9 Hz, 1H), 2.58 (s, 3H). <sup>13</sup>**C-NMR** (75 MHz, DMSO)  $\delta$  191.5 (C $_q$ ), 162.5 (C $_q$ ), 148.2 (C $_q$ ), 141.0 (C $_q$ ), 133.7, 133.6, 26.8. **HRMS** (EI+): calculated m/z for C $_7$ H $_6$ O $_3$ S [M $^{++}$ ] 170.00322; found 170.00333. Data in accordance with literature.<sup>24</sup>

# 5-Carbamoylthiophene-2-carboxylic acid (4d)

The title compound was prepared according to GP2a. Isolation of the compound was accomplished by employing C18 reversed-phase silica gel column chromatography using a mixture of water and acetonitrile as eluents. Thiophene-2-carboxamide (0.4 mmol) gave **4d** (91%) as a white solid; <sup>1</sup>**H-NMR** (400 MHz, DMSO- $d_6$ )  $\delta$  13.40 (bs, 1H), 8.17 (bs, 1H), 7.73 (d, J = 3.9 Hz, 1H), 7.69 (d, J = 3.9 Hz, 1H), 7.64 (bs, 1H). <sup>13</sup>C-NMR (101 MHz, DMSO)  $\delta$  162.7 (C $_q$ ), 162.2 (C $_q$ ), 145.7 (C $_q$ ), 137.9 (C $_q$ ), 133.3, 128.9. **HRMS** (ESI+): calculated m/z for C $_6$ H $_6$ O $_3$ S [(M+H) $^+$ ] 172.0063; found 172.0065.

#### 5-Phenylthiophene-2-carboxylic acid (4e)

Following GP2a, 3-phenylthiophene (0.4 mmol) gave 4e (99%) as a white solid;  ${}^{1}$ H-NMR (300 MHz, DMSO-d6)  $\delta$  13.18 (bs, 1H), 7.77 – 7.68 (m, 3H), 7.55 (d, J = 4.0 Hz, 1H), 7.49 – 7.33 (m, 3H).  ${}^{13}$ C-NMR (75 MHz, DMSO)  $\delta$  162.9 (C<sub>q</sub>), 149.8 (C<sub>q</sub>), 134.4, 133.4 (C<sub>q</sub>), 132.9 (C<sub>q</sub>), 129.3, 128.9, 125.9, 124.6. HRMS (ESI+): calculated m/z for C<sub>11</sub>H<sub>9</sub>O<sub>2</sub>S [(M+H)<sup>+</sup>] 205.0318; found 205.0321. Data in accordance with the literature.  ${}^{25}$ 

#### 5-(Trimethylsilyl)thiophene-2-carboxylic acid (4f)

Following GP2a, trimethyl(thiophen-2-yl)silane (0.4 mmol) gave **4f** (28%) as a white solid; <sup>1</sup>**H-NMR** (400 MHz, DMSO- $d_6$ )  $\delta$  13.02 (bs, 1H), 7.75 (d, J = 3.5 Hz, 1H), 7.34 (d, J = 3.5 Hz, 1H), 0.31 (s, 9H). <sup>13</sup>**C-NMR** (101 MHz, DMSO)  $\delta$  162.6 (C<sub>q</sub>), 148.1 (C<sub>q</sub>), 139.5 (C<sub>q</sub>), 135.0, 133.9, -0.4. **HRMS** (EI+): calculated m/z for C<sub>8</sub>H<sub>12</sub>O<sub>2</sub>SiS [M<sup>++</sup>] 200.03218; found 200.03162.

#### 5-(cyanomethyl)thiophene-2-carboxylic acid (4g)

Following GP2b, 2-(thiophen-2-yl)acetonitrile (0.1 mmol) gave **4g** (89%) as a white soild. <sup>1</sup>**H-NMR** (500 MHz,CDCl<sub>3</sub>)  $\delta$  7.78 (d, J = 3.8 Hz, 1H), 7.13 (d, J =3.8 Hz, 1H), 3.96 (s, 2H); <sup>13</sup>**C-NMR** (126 MHz, CDCl<sub>3</sub>)  $\delta$  165.6 (C<sub>q</sub>), 139.8 (C<sub>q</sub>), 135.3, 133.2 (C<sub>q</sub>), 128.3, 115.9 (C<sub>q</sub>), 19.3 (CH<sub>2</sub>); **HRMS** (ESI): calculated m/z for C<sub>7</sub>H<sub>4</sub>NO<sub>2</sub>S [(M-H)<sup>+</sup>] 165.9968; found 165.9967.

#### 5-(2-methoxy-2-oxoethyl)thiophene-2-carboxylic acid (4h)

Following GP2b, methyl 2-(thiophen-2-yl)acetate (0.1 mmol) gave **4h** (99%) as a waxy solid. <sup>1</sup>**H-NMR** (500 MHz, CDCl<sub>3</sub>)  $\delta$  7.76 (d, J = 3.8 Hz, 1H), 6.99 (d, J = 3.8 Hz, 1H), 3.8 (s, 2H), 3.76 (s, 3H); <sup>13</sup>**C-NMR** (126 MHz, CDCl<sub>3</sub>)  $\delta$  170.0, 167.4, 144.5, 135.2, 132.2, 128.2, 52.7, 35.9; **HRMS** (ESI): calculated m/z for  $C_8H_7O_4S$  [(M-H)<sup>+</sup>] 199.0065; found 199.0065.

#### 3-Cyanothiophene-2-carboxylic acid (4i)

Following GP2a, thiophene-3-carbonitrile (0.4 mmol) gave **4i** (86%) as a pale yellow solid; <sup>1</sup>**H-NMR** (300 MHz, DMSO- $d_6$ )  $\delta$  14.12 (bs, 1H), 8.06 (d, J = 5.2 Hz, 1H), 7.62 (d, J = 5.1 Hz, 1H). <sup>13</sup>**C-NMR** (75 MHz, DMSO)  $\delta$  160.7 (C<sub>q</sub>), 141.9 (C<sub>q</sub>), 133.9, 131.6, 114.1 (C<sub>q</sub>), 113.0 (C<sub>q</sub>). **HRMS** (ESI+): calculated m/z for C<sub>6</sub>H<sub>4</sub>NO<sub>2</sub>S [(M+H)<sup>+</sup>] 153.9957; found 153.9957. Data in accordance with the literature.<sup>26</sup>

#### 3-Acetylthiophene-2-carboxylic acid (4j)

Following GP2a, 3-acetylthiophene (0.4 mmol) gave **4j** (41%) as a pale yellow solid; <sup>1</sup>**H-NMR** (300 MHz, Acetonitrile- $d_3$ )  $\delta$  7.78 (d, J = 5.4 Hz, 1H), 7.71 (d, J = 5.4 Hz, 1H), 2.73 (s, 3H). <sup>1</sup>**H-NMR** (400 MHz, DMSO- $d_6$ )  $\delta$  13.59 (bs, 1H), 7.85 (d, J = 5.0 Hz, 1H), 7.24 (d, J = 5.0 Hz, 1H), 2.50 (s, 3H). <sup>13</sup>C-**NMR** (101 MHz, DMSO)  $\delta$  199.3 (C<sub>q</sub>), 162.3 (C<sub>q</sub>), 146.9 (C<sub>q</sub>), 132.2 (C<sub>q</sub>), 132.1, 128.1, 30.9. **HRMS** (EI+): calculated m/z for C<sub>7</sub>H<sub>6</sub>O<sub>3</sub>S [M<sup>++</sup>] 170.00322; found 170.00304.

# 3-(Methoxycarbonyl)thiophene-2-carboxylic acid (4k)

Following GP2a, methyl thiophene-2-carboxylate (0.4 mmol) gave **4k** (88%) as a white solid; <sup>1</sup>**H-NMR** (400 MHz, DMSO- $d_6$ )  $\delta$  13.53 (bs, 1H), 7.86 (d, J = 5.1 Hz, 1H), 7.31 (d, J = 5.1 Hz, 1H), 3.80 (s, 3H). <sup>13</sup>**C-NMR** (101 MHz, DMSO)  $\delta$  164.8 (C<sub>q</sub>), 161.9 (C<sub>q</sub>), 136.8 (C<sub>q</sub>), 134.5 (C<sub>q</sub>), 131.8, 128.3, 52.5. **HRMS** (ESI+): calculated m/z for C<sub>7</sub>H<sub>7</sub>O<sub>4</sub>S [(M+H)<sup>+</sup>] 187.0060; found 187.0059.

#### 3-Phenylthiophene-2-carboxylic acid (4l)

Following GP2a, 3-phenylthiophene (0.4 mmol) gave **4l** (71%) as a pale yellow solid; <sup>1</sup>**H-NMR** (400 MHz, DMSO- $d_6$ )  $\delta$  12.86 (bs, 1H), 7.85 (d, J = 5.0 Hz, 1H), 7.49 – 7.42 (m, 2H), 7.42 – 7.32 (m, 3H), 7.17 (d, J = 5.1 Hz, 1H). <sup>13</sup>C-NMR (101 MHz, DMSO)  $\delta$  162.8 (C<sub>q</sub>), 147.1 (C<sub>q</sub>), 135.5 (C<sub>q</sub>), 131.7, 131.0, 129.3, 128.1 (C<sub>q</sub>), 127.7, 127.6. **HRMS** (ESI+): calculated m/z for C<sub>11</sub>H<sub>9</sub>O<sub>2</sub>S [(M+H)<sup>+</sup>] 205.0318; found 205.0319. Data in accordance with the literature.<sup>27</sup>

#### [2,2'-Bithiophene]-5-carboxylic acid (4m)

Following GP2a, 2,2'-bithiophene (0.4 mmol) gave **4m** (53%) as a pale yellow solid; <sup>1</sup>**H-NMR** (400 MHz, DMSO- $d_6$ )  $\delta$  13.15 (bs, 1H), 7.66 (d, J = 3.9 Hz, 1H), 7.62 (dd, J = 5.1, 1.1 Hz, 1H), 7.48 (dd, J = 3.7, 1.1 Hz, 1H), 7.34 (d, J = 3.9 Hz, 1H), 7.13 (dd, J = 5.1, 3.6 Hz, 1H). <sup>13</sup>**C-NMR** (101 MHz, DMSO)  $\delta$  162.6 (C<sub>q</sub>), 142.9 (C<sub>q</sub>), 135.4 (C<sub>q</sub>), 134.2, 132.5 (C<sub>q</sub>), 128.6, 127.2, 125.9, 124.5. **HRMS** (ESI+): calculated m/z for C<sub>9</sub>H<sub>7</sub>O<sub>2</sub>S<sub>2</sub> [(M+H)<sup>+</sup>] 210.9882; found 210.9884. Data in accordance with the literature.<sup>28</sup>

#### [2,3'-Bithiophene]-2'-carboxylic acid (4n) & [2,3'-bithiophene]-5-carboxylic acid (4n')

Following GP2a, 2,3'-bithiophene (0.4 mmol) gave 4n (64%) as a white solid and 4n' (23%) as a pale yellow solid. 4n:4n'=2.8:1.

Data for **4n**: <sup>1</sup>**H-NMR** (400 MHz, Chloroform-d)  $\delta$  7.60 (dd, J = 3.7, 1.2 Hz, 1H), 7.56 (d, J = 5.2 Hz, 1H), 7.40 (dd, J = 5.1, 1.2 Hz, 1H), 7.27 (d, J = 5.2 Hz, 1H), 7.10 (dd, J = 5.1, 3.7 Hz, 1H). <sup>1</sup>**H-NMR** (400 MHz, DMSO- $d_6$ )  $\delta$  13.07 (bs, 1H), 7.84 (d, J = 5.2 Hz, 1H), 7.65 – 7.59 (m, 2H), 7.36 (d, J = 5.2

Hz, 1H), 7.11 (dd, J = 5.1, 3.7 Hz, 1H). <sup>13</sup>C-NMR (101 MHz, DMSO)  $\delta$  162.8 (C<sub>q</sub>), 138.7 (C<sub>q</sub>), 135.9 (C<sub>q</sub>), 131.3, 131.2, 129.0, 127.3, 127.2, 126.7 (C<sub>q</sub>). HRMS (ESI+): calculated m/z for C<sub>9</sub>H<sub>7</sub>O<sub>2</sub>S<sub>2</sub> [(M+H)<sup>+</sup>] 210.9882; found 210.9883. Data in accordance with the literature.<sup>29</sup>

Data for **4n'**: <sup>1</sup>**H-NMR** (400 MHz, DMSO- $d_6$ )  $\delta$  7.93 (dd, J = 2.9, 1.4 Hz, 1H), 7.69 – 7.64 (m, 2H), 7.49 (dd, J = 5.0, 1.4 Hz, 1H), 7.44 (d, J = 3.8 Hz, 1H). <sup>13</sup>**C-NMR** (101 MHz, DMSO)  $\delta$  162.9 (C<sub>q</sub>), 144.7 (C<sub>q</sub>), 134.3 (C<sub>q</sub>), 134.0, 132.6 (C<sub>q</sub>), 128.0, 126.0, 124.5, 122.4. Data in accordance with the literature.<sup>30</sup>

#### Thieno[3,2-b]thiophene-2-carboxylic acid (40)

Following GP2a, thieno[3,2-*b*]thiophene (0.4 mmol) gave **4o** (48%) as a pale green solid; <sup>1</sup>**H-NMR** (400 MHz, DMSO- $d_6$ )  $\delta$  13.20 (bs, 1H), 8.11 (s, 1H), 7.92 (d, J = 5.3 Hz, 1H), 7.51 (d, J = 5.3 Hz, 1H). <sup>13</sup>**C-NMR** (101 MHz, DMSO)  $\delta$  163.4 (C<sub>q</sub>), 143.2 (C<sub>q</sub>), 138.6 (C<sub>q</sub>), 135.7 (C<sub>q</sub>), 133.0, 126.1, 120.3. **HRMS** (ESI+): calculated m/z for C<sub>7</sub>H<sub>5</sub>O<sub>2</sub>S<sub>2</sub> [(M+H)<sup>+</sup>] 184.9725; found 184.9728. Data in accordance with the literature.<sup>31</sup>

#### Benzo[b]thiophene-2-carboxylic acid (4p)

Following GP2a, benzo[*b*]thiophene (0.4 mmol) gave **4p** (97%) as a white solid; <sup>1</sup>**H-NMR** (400 MHz, DMSO- $d_6$ )  $\delta$  13.45 (bs, 1H), 8.11 (s, 1H), 8.06 – 7.96 (m, 2H), 7.54 –7.41 (m, 2H). <sup>13</sup>**C-NMR** (101 MHz, DMSO)  $\delta$  163.5 (C<sub>q</sub>), 141.3 (C<sub>q</sub>), 138.7 (C<sub>q</sub>), 134.8 (C<sub>q</sub>), 130.2, 127.0, 125.7, 125.1, 123.0. **HRMS** (ESI+): calculated m/z for C<sub>9</sub>H<sub>7</sub>O<sub>2</sub>S [(M+H)<sup>+</sup>] 179.0161; found 179.0163. Data in accordance with the literature.<sup>32</sup>

#### 3-Methylbenzo[b]thiophene-2-carboxylic acid (4q)

Following GP2a, 3-methylbenzo[b]thiophene (0.4 mmol) gave 4q (95%) as a white solid; <sup>1</sup>H-NMR (300 MHz, DMSO- $d_6$ )  $\delta$  13.37 (bs, 1H), 8.01 – 7.88 (m, 2H), 7.55 – 7.41 (m, 2H), 2.70 (s, 3H). <sup>13</sup>C-NMR (75 MHz, DMSO)  $\delta$  164.4 (C<sub>q</sub>), 140.0 (C<sub>q</sub>), 139.8 (C<sub>q</sub>), 139.4 (C<sub>q</sub>), 127.9 (C<sub>q</sub>), 127.3, 124.7, 123.9, 122.8, 12.8. HRMS (ESI+): calculated m/z for C<sub>10</sub>H<sub>9</sub>O<sub>2</sub>S [(M+H)<sup>+</sup>] 193.0318; found 193.0319. Data in accordance with the literature.<sup>33</sup>

#### 5-(Methoxycarbonyl)furan-2-carboxylic acid (4r)

$$HO_2C$$
  $O$   $CO_2Me$ 

Following GP2a, methyl furan-2-carboxylate (0.4 mmol) gave **4r** (70%) as a pale yellow solid. <sup>1</sup>**H-NMR** (400 MHz, DMSO-d6)  $\delta$  13.72 (bs, 1H), 7.39 (d, J = 3.6 Hz, 1H), 7.32 (d, J = 3.6 Hz, 1H), 3.85 (s, 3H). <sup>13</sup>**C-NMR** (101 MHz, DMSO)  $\delta$  158.8 (C<sub>q</sub>), 157.9 (C<sub>q</sub>), 147.4 (C<sub>q</sub>), 145.6 (C<sub>q</sub>), 119.0, 118.4, 52.3. **HRMS** (ESI+): calculated m/z for C<sub>7</sub>H<sub>7</sub>O<sub>5</sub> [(M+H)<sup>+</sup>] 171.0288; found 171.0289. Data in accordance with the literature.<sup>34</sup>

#### 5-Cyanofuran-2-carboxylic acid (4s)

Following GP2a, furan-2-carbonitrile (0.4 mmol) gave **4s** (43%) as a pale yellow solid; <sup>1</sup>**H-NMR** (400 MHz, DMSO- $d_6$ )  $\delta$  13.98 (bs, 1H), 7.72 (d, J = 3.8 Hz, 1H), 7.40 (d, J = 3.8 Hz, 1H). <sup>13</sup>**C-NMR** (101 MHz, DMSO)  $\delta$  158.0 (C<sub>q</sub>), 149.0 (C<sub>q</sub>), 126.9 (C<sub>q</sub>), 124.6, 117.8, 111.0 (C<sub>q</sub>). **HRMS** (EI+): calculated m/z for C<sub>6</sub>H<sub>3</sub>NO<sub>3</sub> [M<sup>++</sup>] 137.01074; found 137.01037.

# Benzofuran-2-carboxylic acid (4t)

Following GP2a, benzofuran (0.4 mmol) gave **4t** (47%) as a pale yellow solid; <sup>1</sup>**H-NMR** (400 MHz, DMSO- $d_6$ )  $\delta$  13.55 (bs, 1H), 7.78 (d, J = 7.8 Hz, 1H), 7.69 (d, J = 8.4 Hz, 1H), 7.67 – 7.63 (m, 1H), 7.49 (ddd, J = 8.4, 7.1, 1.3 Hz, 1H), 7.38 – 7.32 (m, 1H). <sup>13</sup>**C-NMR** (101 MHz, DMSO)  $\delta$  160.1 (C<sub>q</sub>), 155.0 (C<sub>q</sub>), 146.3 (C<sub>q</sub>), 127.5, 126.9 (C<sub>q</sub>), 123.8, 123.1, 113.4, 112.1. **HRMS** (EI+): calculated m/z for C<sub>9</sub>H<sub>6</sub>O<sub>3</sub> [M<sup>++</sup>] 162.03115; found 162.03138. Data in accordance with the literature. <sup>17</sup>

#### 1-(tert-butyl) 2-methyl 1H-indole-1,2-dicarboxylate (4u)

A dry microwave vial (5 mL) equipped with stirring bar, was charged with tert-butyl 1H-indole-1carboxylate (0.1 mmol) and 2,3,6,7-tetramethoxyanthracen-9(10H)-one (20 mol%). The vial was sealed and transferred to a glovebox after the vial had evacuated and refilled with N2 (3×) within the antechamber via syringe. Cs<sub>2</sub>CO<sub>3</sub> (3.0 equiv.) and dry DMSO (1 mL) was added and the vial was sealed with a Supelco aluminium crimp seal with septum (PTFE/butyl). The vial was removed from the glovebox. The reaction mixture then had gaseous CO<sub>2</sub> (22 cm<sup>3</sup>) added via a gastight Hamilton® syringe, into the head space of the vial. The vial was then irradiated from two sides by two kessil lamps (vials approximately 6 cm away from the light source) with fans placed in front of the vials for cooling (temperatures measured via IR ranged between 25-30 °C). After 18 hrs the irradiation was stopped, the pressure was vented via syringe and MeI (31 µL, 0.5 mmol) was added. The reaction mixture was left to stir for 4 hrs. The vial was decapped and quenched with aq. HCl (1 mL, 0.3M). The reaction was diluted with brine (10 mL) extracted with EtOAc (3×5 mL). The organic layers were combined and washed with brine (3×10 mL) and then dried via phase separator and concentrated in vacuo. Purification by column chromatography on silica gel eluting with heptane:EtOAc (9:1) gave the title compound 4u (40%) as a white waxy solid.  $R_f 0.51$  [Heptane–EtOAc (9:1)]; 8.10 (dq, J = 8.5, 0.9 Hz, 1H), 7.60 (ddd, J = 7.8, 1.2, 0.8 Hz, 1H), 7.42 (ddd, J = 8.5, 7.2, 1.3 Hz, 1H), 7.27 (ddd, J = 8.1, 7.2, 1.0 Hz, 1H), 7.11 (d, J = 0.8 Hz, 1H), 3.93 (s, 3H), 1.63 (s, 9H).  $^{13}$ C-NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  162.5, 149.4, 137.9, 130.5, 127.6, 126.9, 123.4, 122.3, 115.0, 115.0, 84.7, 52.5, 27.9. Data in accordance with the literature.<sup>35</sup>

#### 1-(tert-Butoxycarbonyl)-2-phenyl-1H-indole-3-carboxylic acid (4v)

Following GP2a, tert-butyl 2-phenyl-1H-indole-1-carboxylate (0.4 mmol) gave 4v (10%) as a white solid;  ${}^{1}H$ -NMR (400 MHz, DMSO- $d_6$ )  $\delta$  12.40 (bs, 1H), 8.17 – 8.12 (m, 1H), 8.12 – 8.07 (m, 1H), 7.48 – 7.32 (m, 7H), 1.16 (s, 9H).  ${}^{13}C$ -NMR (101 MHz, DMSO)  $\delta$  164.9 (C<sub>q</sub>), 148.9 (C<sub>q</sub>), 143.8 (C<sub>q</sub>), 135.3 (C<sub>q</sub>), 133.1 (C<sub>q</sub>), 129.7, 128.1, 127.4, 126.9 (C<sub>q</sub>), 124.9, 123.7, 121.6, 114.2, 111.4 (C<sub>q</sub>), 84.3 (C<sub>q</sub>), 26.8. HRMS (ESI+): calculated m/z for C<sub>20</sub>H<sub>20</sub>NO<sub>4</sub> [(M+H)<sup>+</sup>] 338.1387; found 338.1391.

#### 1-Phenyl-1*H*-pyrazole-5-carboxylic acid (4w)

Following GP2a, 1-phenylpyrazole (0.4 mmol) gave **4w** (33%) as a white solid; <sup>1</sup>**H-NMR** (300 MHz, Methanol- $d_4$ )  $\delta$  7.72 (d, J = 2.0 Hz, 1H), 7.52 – 7.39 (m, 5H), 7.05 (d, J = 2.0 Hz, 1H). <sup>1</sup>**H-NMR** (400 MHz, DMSO- $d_6$ )  $\delta$  13.29 (bs, 1H), 7.77 (d, J = 1.9 Hz, 1H), 7.52 – 7.41 (m, 5H), 7.03 (d, J = 1.9 Hz, 1H). <sup>13</sup>**C-NMR** (101 MHz, DMSO)  $\delta$  160.0 (C<sub>q</sub>), 140.2 (C<sub>q</sub>), 139.7, 134.1 (C<sub>q</sub>), 128.5, 128.2, 125.7, 112.5. **HRMS** (ESI+): calculated m/z for C<sub>10</sub>H<sub>9</sub>N<sub>2</sub>O<sub>2</sub> [(M+H)<sup>+</sup>] 189.0659; found 189.0661. Data in accordance with the literature.<sup>36</sup>

#### 2-cyano-4-methylthiazole-5-carboxylic acid (4x)

Following GP2c, 4-methylthiazole-2-carbonitrile (0.1 mmol) gave 4x (53%) as a white soild. <sup>1</sup>H-NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  2.83 (3H, s); <sup>13</sup>C-NMR (126 MHz, CDCl<sub>3</sub>)  $\delta$  165.4 (Cq), 163.3 (Cq), 139.3 (Cq), 126.6 (Cq), 112.1 (Cq), 17.7; HRMS (EI): calculated m/z for C<sub>6</sub>H<sub>4</sub>N<sub>2</sub>O<sub>2</sub>S [M<sup>++</sup>] 167.9992; found 168.0003.

# 4-hydroxybenzo[b]thiophene-7-carboxylic acid (6a)

Following GP2c, 4-hydroxybenzo[b]thiophene (0.1 mmol) gave **6a** (99%) as a white solid. <sup>1</sup>**H-NMR** (500 MHz, DMSO-d<sub>6</sub>):  $\delta$  7.91 (d, J = 8.2 Hz, 1H), 7.66 (d, J = 5.6 Hz, 1H), 7.51 (d, J = 5.6 Hz, 1H), 6.85 (d, J = 8.2 Hz, 1H); <sup>13</sup>**C-NMR** (126 MHz, DMSO-d<sub>6</sub>)  $\delta$  166.9 (C<sub>q</sub>), 157.0 (C<sub>q</sub>), 141.8 (C<sub>q</sub>), 129.8 (C<sub>q</sub>), 129.3, 127.7, 119.8, 115.6 (C<sub>q</sub>), 108.9; **HRMS** (ESI): calculated m/z for C<sub>9</sub>H<sub>5</sub>O<sub>3</sub>S [(M-H)<sup>+</sup>] 192.9965; found 192.9965.

#### 1-(Tetrahydro-2*H*-pyran-2-yl)-1*H*-indazole-4-carboxylic acid (6b)

Following GP2c, 1-(tetrahydro-2*H*-pyran-2-yl)-1*H*-indazole (0.1 mmol) gave **6b** (60%) as a white solid. **¹H-NMR** (500 MHz, DMSO-d6):  $\delta$  12.88 (bs, 1H), 8.42 (s, 1H), 8.03 (d, J = 8.4 Hz, 1H), 7.83 (d, J = 7.1 Hz, 1H), 7.63-7.43 (m, 1H), 5.92 (dd, J = 9.6, 2.1 Hz, 1H), 3.88 (d, J = 11.2 Hz, 1H), 3.80-3.71 (m, 1H), 2.42 (qd, J = 3.7, 13.0 Hz, 1H), 2.15-1.94 (m, 2H), 1.88-1.67 (m, 1H), 1.65-1.52 (m, 2H); <sup>13</sup>C-NMR (126 MHz, DMSO-d6):  $\delta$  167.0, 139.8, 133.6, 125.9, 124.4, 123.3, 122.5, 115.3, 84.0, 66.5, 28.9, 24.7, 22.1; **HRMS** (ESI): calculated m/z for C<sub>13</sub>H<sub>15</sub>N<sub>2</sub>O<sub>3</sub> [(M+H)<sup>†</sup>] 247.1077; found 247.1066.

#### 1-Methyl-1*H*-indazole-4-carboxylic acid (6c)

Following GP2c, 1-methyl-1*H*-indazole (0.1 mmol) gave **6c** (58%) as a white solid. <sup>1</sup>**H-NMR** (500 MHz, CDCl<sub>3</sub>):  $\delta$  8.58 (s, 1H), 8.05 (d, J = 6.9 Hz, 1H) 7.69 (d, J = 8.8 Hz, 1H), 7.44–7.56 (m, 1H), 4.16 (s, 3H); <sup>13</sup>**C-NMR** (126 MHz, CDCl<sub>3</sub>): 171.3, 140.5, 133.9, 125.7, 125.3, 122.8, 122.3, 114.9, 35.9; **HRMS** (ESI): calculated m/z for C<sub>9</sub>H<sub>9</sub>N<sub>2</sub>O<sub>2</sub> [(M+H)<sup>+</sup>] 177.0664; found 177.0669.

#### Pyrazolo[1,5-a]pyridine-7-carboxylic acid (6d) & pyrazolo[1,5-a]pyridine-4-carboxylic acid (6d')

Following GP2c, pyrazolo[1,5-a]pyridine (0.1 mmol) gave **6d** (63%) as a white solid & **6d'** (22%) as a white solid. **6d:6d'** = 2.8:1.

**Data for 6d:** <sup>1</sup>**H-NMR** (500 MHz, DMSO-d<sub>6</sub>)  $\delta$  8.20 (d, J = 2.3 Hz, 1H), 8.14–7.88 (m, 1H), 7.73–7.58 (m, 1H), 7.38 (dd, J = 7.1, 8.8 Hz, 1H), 6.87 (d, J = 2.3 Hz, 1H); <sup>13</sup>**C-NMR** (126 MHz, DMSO-d<sub>6</sub>)  $\delta$  161.8, 140.8, 140.6, 129.3, 123.6, 122.3, 116.9, 98.5. Data in accordance with the literature.<sup>37</sup>

**Data for 6d'**: <sup>1</sup>**H-NMR** (500 MHz, DMSO-d<sub>6</sub>)  $\delta$  13.34 (bs, 1H), 8.94 (d, J = 6.8 Hz, 1H), 8.12 (d, J = 2.1 Hz, 1H), 7.95 (d, J = 6.4 Hz, 1H), 7.03–6.98 (m, 2H); <sup>13</sup>**C-NMR** (126 MHz, DMSO-d<sub>6</sub>)  $\delta$  165.5, 142.9, 137.9, 133.0, 128.6, 120.7, 110.9, 98.6. Data in accordance with the literature.<sup>37</sup>

# 4-(3-((tert-butoxycarbonyl)(isopropyl)amino)-2-hydroxypropoxy) benzo[b] thiophene-2-carboxylic acid (6e)

Following GP2c, tert-butyl (3-(benzo[b]thiophen-4-yloxy)-2-hydroxypropyl)(isopropyl)carbamate (0.1 mmol) gave **6e** (18%) as a yellow oil.  ${}^{1}$ **H-NMR** (500 MHz, CDCl<sub>3</sub>):  $\delta$  8.26 (s, 1H), 7.32–7.49 (m, 2H), 6.77 (d, J = 7.5 Hz, 1H), 4.28–4.11 (m, 3H), 4.10–3.99 (m, 1H), 3.47 (s, 2H), 2.05 (s, 1H), 1.50 (s, 9H), 1.18 (dd, J = 6.5, 31.4 Hz, 6H);  ${}^{13}$ **C-NMR** (126 MHz, CDCl<sub>3</sub>)  $\delta$   ${}^{13}$ C NMR (126 MHz, CDCl<sub>3</sub>)  $\delta$  165.6, 154.3, 143.4, 131.0, 129.1, 127.7, 127.3, 114.4, 104.2, 80.0, 71.0, 69.1, 48.0, 46.1, 27.6, 19.7 **HRMS** (ESI): calculated m/z for C<sub>20</sub>H<sub>28</sub>NO<sub>6</sub>S [(M+H)<sup>+</sup>] 410.1637; found 410.1655.

#### Cinnamic acid (8a)

Following GP2a, styrene (0.4 mmol) gave **8a** (54%) as a white solid;  ${}^{1}$ **H-NMR** (400 MHz, DMSO- $d_6$ )  $\delta$  12.40 (bs, 1H), 7.72 – 7.64 (m, 2H), 7.59 (d, J = 16.0 Hz, 1H), 7.45 – 7.37 (m, 3H), 6.53 (d, J = 16.0 Hz, 1H).  ${}^{13}$ **C-NMR** (101 MHz, DMSO)  $\delta$  167.6 (C<sub>q</sub>), 143.9, 134.2 (C<sub>q</sub>), 130.2, 128.9, 128.2, 119.3. **HRMS** (EI+): calculated m/z for C<sub>9</sub>H<sub>8</sub>O<sub>2</sub> [M<sup>++</sup>] 148.05188; found 148.05161. Data in accordance with the literature.  ${}^{38}$ 

# (E)-3-(4-(tert-Butyl)phenyl)acrylic acid (8b)

Following GP2a, 1-(*tert*-butyl)-4-vinylbenzene (0.4 mmol) gave **8b** (56%) as a white solid; <sup>1</sup>**H-NMR** (400 MHz, DMSO- $d_6$ )  $\delta$  12.33 (bs, 1H), 7.62 – 7.53 (m, 3H), 7.46 – 7.38 (m, 2H), 6.47 (d, J = 16.0 Hz, 1H), 1.27 (s, 9H). <sup>13</sup>**C-NMR** (101 MHz, DMSO)  $\delta$  167.7 (C<sub>q</sub>), 153.1 (C<sub>q</sub>), 143.8, 131.5 (C<sub>q</sub>), 128.0, 125.7, 118.3, 34.6 (C<sub>q</sub>), 30.9. **HRMS** (EI+): calculated m/z for C<sub>13</sub>H<sub>16</sub>O<sub>2</sub> [M<sup>++</sup>] 204.11448; found 204.11415.

# (E)-3-(p-Tolyl)acrylic acid (8c)

Following GP2a, 4-methylstyrene (0.4 mmol) gave **8c** (53%) as a white solid; <sup>1</sup>**H-NMR** (400 MHz, DMSO- $d_6$ )  $\delta$  12.31 (bs, 1H), 7.60 – 7.51 (m, 3H), 7.22 (d, J = 7.9 Hz, 2H), 6.46 (d, J = 16.0 Hz, 1H), 2.32 (s, 3H). <sup>13</sup>**C-NMR** (101 MHz, DMSO)  $\delta$  167.7 (C<sub>q</sub>), 143.9, 140.1 (C<sub>q</sub>), 131.5 (C<sub>q</sub>), 129.5, 128.2, 118.1, 21.0. **HRMS** (EI+): calculated m/z for C<sub>10</sub>H<sub>10</sub>O<sub>2</sub> [M<sup>++</sup>] 162.06753; found 162.06783.

# (E)-3-(4-Methoxyphenyl)acrylic acid (8d)

Following GP2a, 4-methoxystyrene (0.4 mmol) gave **8d** (40%) as a pale yellow solid;  ${}^{1}$ **H-NMR** (400 MHz, DMSO- $d_6$ )  $\delta$  12.21 (bs, 1H), 7.66 – 7.60 (m, 2H), 7.54 (d, J = 16.0 Hz, 1H), 7.00 – 6.93 (m, 2H), 6.37 (d, J = 15.9 Hz, 1H), 3.79 (s, 3H).  ${}^{13}$ **C-NMR** (101 MHz, DMSO)  $\delta$  167.8 ( $C_q$ ), 160.9 ( $C_q$ ), 143.7, 129.9, 126.8 ( $C_q$ ), 116.5, 114.3, 55.3. **HRMS** (EI+): calculated m/z for  $C_{10}$ H<sub>10</sub>O<sub>3</sub> [ $M^{+*}$ ] 178.06245; found 178.06194.

# (E)-3-(3-Methoxyphenyl)acrylic acid (8e):

Following GP2a, 3-methoxystyrene (0.4 mmol) gave **8e** (46%) as a white solid; <sup>1</sup>**H-NMR** (400 MHz, DMSO- $d_6$ )  $\delta$  12.40 (bs, 1H), 7.56 (d, J = 16.0 Hz, 1H), 7.36 – 7.20 (m, 3H), 7.03 – 6.92 (m, 1H), 6.55 (d, J = 16.0 Hz, 1H), 3.79 (s, 3H). <sup>13</sup>**C-NMR** (101 MHz, DMSO)  $\delta$  167.6 (C<sub>q</sub>), 159.6 (C<sub>q</sub>), 143.9, 135.7 (C<sub>q</sub>), 129.9, 120.8, 119.6, 116.2, 112.9, 55.2. **HRMS** (EI+): calculated m/z for C<sub>10</sub>H<sub>10</sub>O<sub>3</sub> [M<sup>++</sup>] 178.06245; found 178.06210.

#### (E)-3-(4-(Trimethylsilyl)phenyl)acrylic acid (8f)

Following GP2a, trimethyl(4-vinylphenyl)silane (0.4 mmol) gave **8f** (53%) as a pale yellow solid; <sup>1</sup>**H-NMR** (400 MHz, DMSO- $d_6$ )  $\delta$  12.38 (bs, 1H), 7.67 – 7.62 (m, 2H), 7.61 – 7.52 (m, 3H), 6.54 (d, J = 16.0 Hz, 1H), 0.24 (s, 9H). <sup>13</sup>C-NMR (101 MHz, DMSO)  $\delta$  167.5 (C<sub>q</sub>), 143.9, 142.6 (C<sub>q</sub>), 134.6 (C<sub>q</sub>), 133.7, 127.3, 119.4, –1.3. **HRMS** (EI+): calculated m/z for C<sub>12</sub>H<sub>16</sub>O<sub>2</sub>Si [M<sup>++</sup>] 220.09141; found 220.09148.

#### Methyl (E)-3-(4-(dimethylamino)phenyl)acrylate (8g)

The title compound was prepared according to GP2a with *N*,*N*-dimethyl-4-vinylaniline (0.4 mmol). After irradiating the mixture for 18 hrs the reaction vial was vented and MeI (18.7  $\mu$ L, 0.3 mmol, 3 equiv.) was added *via* syringe. The resulting mixture was stirred for 2 hrs at 35 °C and was quenched by adding water. The crude mixture was extracted with DCM (3×) and the combined organic layers were dried over Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated *in vacuo*. Flash silica gel column chromatography with a mixture of hexanes+NEt<sub>3</sub> (1% v/v) and EtOAc provided the title compound **8g** (14%) as a pale brown solid; <sup>1</sup>**H-NMR** (400 MHz, Chloroform-*d*)  $\delta$  7.63 (d, J = 15.9 Hz, 1H), 7.45 – 7.38 (m, 2H), 6.70 – 6.62 (m, 2H), 6.22 (d, J = 15.8 Hz, 1H), 3.78 (s, 3H), 3.01 (s, 6H). <sup>13</sup>**C-NMR** (101 MHz, CDCl<sub>3</sub>)  $\delta$ 

168.5 ( $C_q$ ), 151.9 ( $C_q$ ), 145.5, 129.9, 122.3 ( $C_q$ ), 112.2, 112.0, 51.5, 40.3. **HRMS** (EI+): calculated m/z for  $C_{12}H_{15}NO_2$  [ $M^{+*}$ ] 205.10973; found 205.10937.

#### (E)-3-(4-(Methylthio)phenyl)acrylic acid (8h)

Following GP2a, methyl(4-vinylphenyl)sulfane (0.4 mmol) gave **8h** (38%) as a white solid; <sup>1</sup>**H-NMR** (400 MHz, DMSO- $d_6$ )  $\delta$  12.32 (bs, 1H), 7.65 – 7.60 (m, 2H), 7.55 (d, J = 16.0 Hz, 1H), 7.31 – 7.24 (m, 2H), 6.48 (d, J = 16.0 Hz, 1H), 2.51 (s, 3H). <sup>13</sup>C-NMR (101 MHz, DMSO)  $\delta$  167.7 (C<sub>q</sub>), 143.4, 141.3 (C<sub>q</sub>), 130.6 (C<sub>q</sub>), 128.7, 125.6, 118.0, 14.2. **HRMS** (EI+): calculated m/z for C<sub>10</sub>H<sub>10</sub>O<sub>2</sub>S [M<sup>++</sup>] 194.03960; found 194.03936.

#### (E)-3-(4-Fluorophenyl)acrylic acid (8i)

Following GP2a, 4-fluorostyrene (0.4 mmol) gave **8i** (46%) as a white solid; <sup>1</sup>**H-NMR** (400 MHz, DMSO- $d_6$ )  $\delta$  12.39 (bs, 1H), 7.80 – 7.71 (m, 2H), 7.59 (d, J = 16.0 Hz, 1H), 7.29 – 7.19 (m, 2H), 6.49 (d, J = 16.0 Hz, 1H). <sup>13</sup>**C-NMR** (101 MHz, DMSO- $d_6$ )  $\delta$  167.5 (C<sub>q</sub>), 163.1 (d, J = 248.3 Hz, C<sub>q</sub>), 142.7, 130.9 (d, J = 3.2 Hz, C<sub>q</sub>), 130.5 (d, J = 8.6 Hz), 119.1 (d, J = 2.2 Hz), 115.9 (d, J = 21.7 Hz). <sup>19</sup>**F-NMR** (376 MHz, DMSO)  $\delta$  -110.0. **HRMS** (EI+): calculated m/z for C<sub>9</sub>H<sub>7</sub>FO<sub>2</sub> [M<sup>++</sup>] 166.04246; found 166.04203.

# (E)-3-(4-(Trifluoromethyl)phenyl)acrylic acid (8j)

Following GP2a, 1-(trifluoromethyl)-4-vinylbenzene (0.4 mmol) gave **8j** (13%) as a white solid; <sup>1</sup>**H-NMR** (400 MHz, DMSO- $d_6$ )  $\delta$  12.58 (bs, 1H), 7.92 (d, J = 8.1 Hz, 2H), 7.76 (d, J = 8.1 Hz, 2H), 7.66 (d, J = 16.1 Hz, 1H), 6.68 (d, J = 16.1 Hz, 1H). <sup>13</sup>**C-NMR** (101 MHz, DMSO)  $\delta$  167.2 (C<sub>q</sub>), 142.0, 138.3 (d, J = 1.3 Hz, C<sub>q</sub>), 129.8 (q, J = 31.8 Hz, C<sub>q</sub>), 128.8, 125.7 (q, J = 3.7 Hz), 124.0 (q, J = 272.1 Hz, C<sub>q</sub>),

122.3. <sup>19</sup>F-NMR (376 MHz, DMSO)  $\delta$  -60.8. HRMS (ESI+): calculated m/z for  $C_{10}H_8F_3O_2$  [(M+H)<sup>+</sup>] 217.0471; found 217.0472. Data in accordance with the literature.<sup>38</sup>

#### (E)-3-(3-Fluorophenyl)acrylic acid (8k)

Following GP2a, 3-fluorostyrene (0.4 mmol) gave **8k** (38%) as a white solid; <sup>1</sup>**H-NMR** (400 MHz, DMSO- $d_6$ )  $\delta$  12.49 (bs, 1H), 7.64 – 7.49 (m, 3H), 7.49 – 7.41 (m, 1H), 7.29 – 7.19 (m, 1H), 6.61 (d, J = 16.0 Hz, 1H). <sup>13</sup>**C-NMR** (101 MHz, DMSO- $d_6$ )  $\delta$  167.4 (C<sub>q</sub>), 162.42 (d, J = 243.7 Hz, C<sub>q</sub>), 142.51 (d, J = 2.6 Hz), 136.82 (d, J = 8.1 Hz, C<sub>q</sub>), 130.81 (d, J = 8.4 Hz), 124.63 (d, J = 2.6 Hz), 120.9, 116.86 (d, J = 21.3 Hz), 114.38 (d, J = 22.0 Hz). <sup>19</sup>**F-NMR** (377 MHz, DMSO)  $\delta$  -112.4. **HRMS** (ESI+): calculated m/z for C<sub>9</sub>H<sub>8</sub>FO<sub>2</sub> [(M+H)<sup>+</sup>] 167.0503; found 167.0502.

#### 3,4-Dihydronaphthalene-2-carboxylic acid (81)

Following GP2a, 1,2-dihydronaphthalene (0.4 mmol) gave **8l** (58%) as a white solid; <sup>1</sup>**H-NMR** (400 MHz, DMSO- $d_6$ )  $\delta$  12.43 (bs, 1H), 7.47 (s, 1H), 7.34 – 7.18 (m, 4H), 2.81 (t, J = 8.3 Hz, 2H), 2.49 – 2.43 (m, 2H). <sup>13</sup>C-NMR (101 MHz, DMSO)  $\delta$  168.1 (C<sub>q</sub>), 136.5 (C<sub>q</sub>), 135.3, 132.3 (C<sub>q</sub>), 130.0 (C<sub>q</sub>), 129.3, 128.3, 127.5, 126.7, 26.9 (CH<sub>2</sub>), 21.9 (CH<sub>2</sub>). **HRMS** (EI+): calculated m/z for C<sub>11</sub>H<sub>10</sub>O<sub>2</sub> [M<sup>++</sup>] 174.06753; found 174.06707. Data in accordance with the literature.<sup>39</sup>

#### 3-(4-Fluorophenyl)but-3-enoic acid (8m) & (E)-3-(4-fluorophenyl)but-2-enoic acid (8m')

Following GP2a, 4-fluoro- $\alpha$ -methylstyrene (0.4 mmol) gave **8m** (43%) as a white solid and **8m'** (9%) as a pale yellow solid. **8m:8m'** = 4.8:1. **HRMS** (EI+): calculated m/z for  $C_{10}H_9FO_2$  [M<sup>++</sup>] 180.05811; found 180.05831.

S-40

**Data for 8m**: <sup>1</sup>**H-NMR** (400 MHz, Chloroform-*d*) δ 9.83 (bs, 1H), 7.45 – 7.35 (m, 2H), 7.06 – 6.97 (m, 2H), 5.52 (s, 1H), 5.24 (s, 1H), 3.52 (s, 2H). <sup>13</sup>**C-NMR** (101 MHz, Chloroform-*d*) δ 177.5 (C<sub>q</sub>), 162.7 (d, J = 247.3 Hz, C<sub>q</sub>), 139.3 (C<sub>q</sub>), 135.7 (d, J = 3.3 Hz, C<sub>q</sub>), 127.61 (d, J = 8.1 Hz), 116.98 (d, J = 1.3 Hz, CH<sub>2</sub>), 115.49 (d, J = 21.5 Hz), 41.1 (CH<sub>2</sub>). <sup>19</sup>**F-NMR** (376 MHz, CDCl<sub>3</sub>) δ -114.8.

**Data for 8m'**: <sup>1</sup>**H-NMR** (400 MHz, Chloroform-*d*)  $\delta$  7.51 – 7.45 (m, 2H), 7.11 – 7.04 (m, 2H), 6.13 (s, 1H), 2.58 (d, J = 0.7 Hz, 3H). <sup>19</sup>**F-NMR** (376 MHz, CDCl<sub>3</sub>)  $\delta$  -112.4. Data in accordance with the literature. <sup>40</sup>

#### 2-(Benzo[b]thiophen-2-yl)propan-2-ol (9pa)

According to GP3 benzo[b]thiophene and acetone gave  $\mathbf{9pa}$  (54%) as a white solid;  ${}^{1}\mathbf{H}$ -NMR (400 MHz, Chloroform-d)  $\delta$  7.80 (d, J = 7.8 Hz, 1H), 7.71 (d, J = 7.7 Hz, 1H), 7.31 (dt, J = 15.1, 7.4 Hz, 2H), 7.16 (s, 1H), 2.26 (s, 1H), 1.73 (s, 6H).  ${}^{13}\mathbf{C}$ -NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  155.1 (C $_q$ ), 139.9 (C $_q$ ), 139.3 (C $_q$ ), 124.3, 124.1, 123.5, 122.4, 118.5, 71.8 (C $_q$ ), 32.1. **HRMS** (ESI+): calculated m/z for C<sub>11</sub>H<sub>13</sub>O<sub>2</sub>S [(M+H)<sup>+</sup>] 175.0576; found 175.0577.

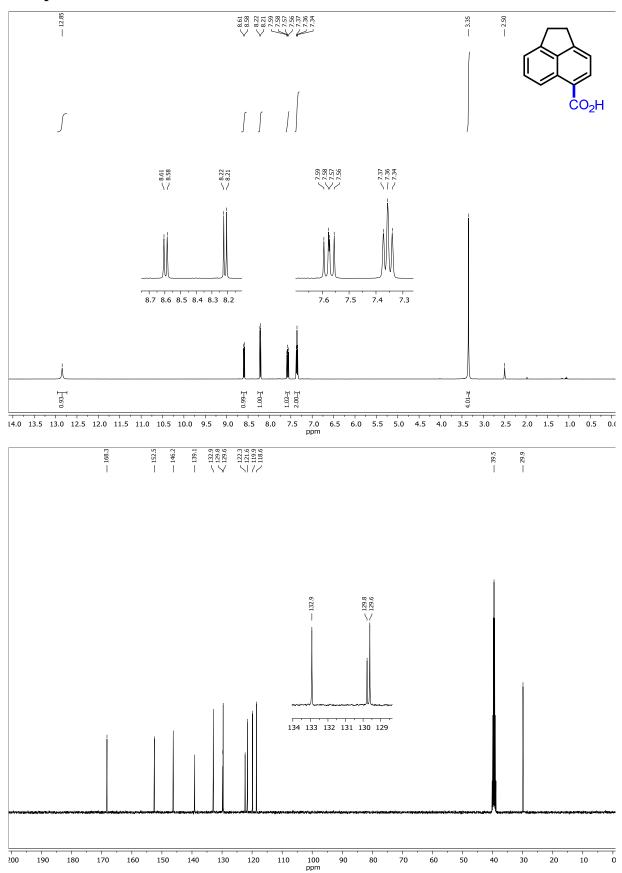
# 1-(Benzo[b]thiophen-2-yl)cyclobutan-1-ol (9pb)

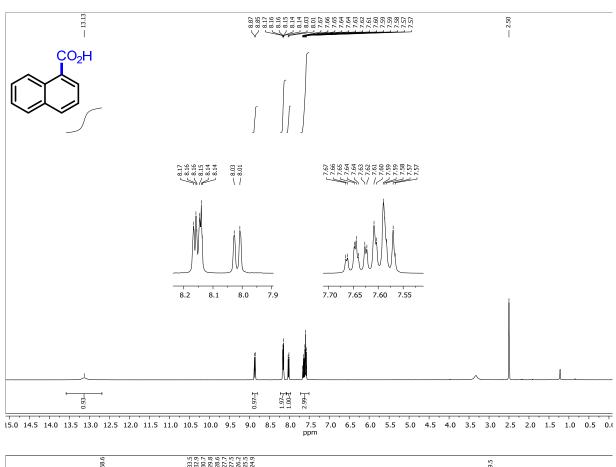
According to GP3 benzo[b]thiophene and cyclobutanone gave **9pb** (30%) as a pale yellow oil; <sup>1</sup>**H-NMR** (300 MHz, Methanol- $d_4$ )  $\delta$  7.84 – 7.73 (m, 1H), 7.76 – 7.70 (m, 1H), 7.35 – 7.22 (m, 3H), 2.65 – 2.52 (m, 2H), 2.52 – 2.38 (m, 2H), 2.06 – 1.77 (m, 2H). <sup>13</sup>**C-NMR** (75 MHz, MeOD)  $\delta$  154.1 (C $_q$ ), 141.3 (C $_q$ ), 141.0 (C $_q$ ), 125.1, 125.0, 124.4, 123.2, 119.8, 75.7 (C $_q$ ), 39.0 (CH $_2$ ), 13.7 (CH $_2$ ). **HRMS** (ESI+): calculated m/z for C $_{12}$ H $_{13}$ OS [(M+H) $^+$ ] 187.0576; found 187.0577.

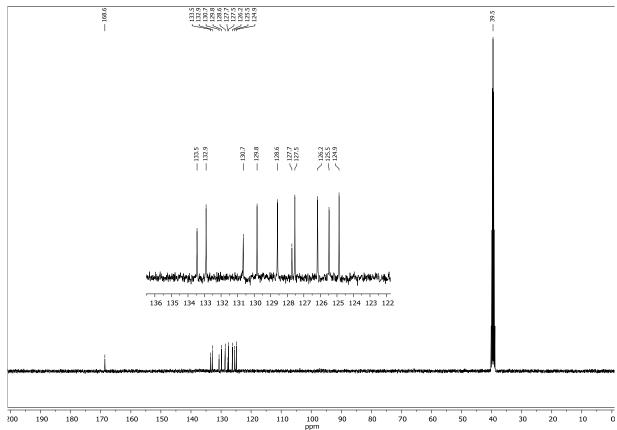
#### 2-(Benzo[b]thiophen-2-yl)hex-5-en-2-ol (9pc)

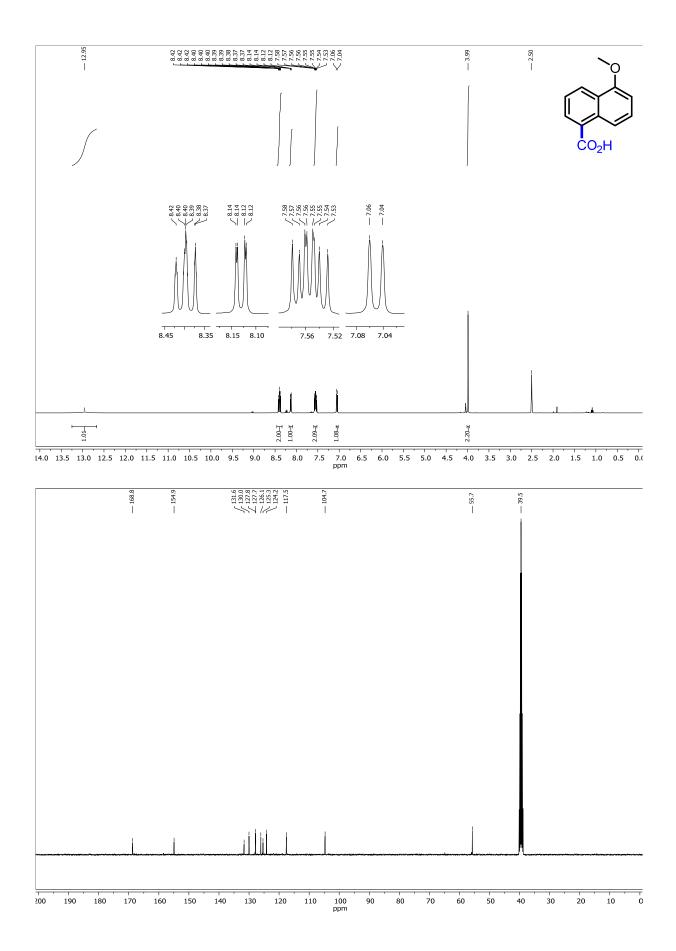
According to GP3 benzo[*b*]thiophene and hex-5-en-2-one gave **9pc** (19%) as a colorless oil; <sup>1</sup>**H-NMR** (400 MHz, Chloroform-*d*)  $\delta$  7.83 – 7.77 (m, 1H), 7.74 – 7.68 (m, 1H), 7.37 – 7.26 (m, 2H), 7.14 (s, 1H), 5.84 (ddt, J = 16.8, 10.2, 6.4 Hz, 1H), 5.02 (dq, J = 17.1, 1.7 Hz, 1H), 4.96 (dq, J = 9.9, 1.3 Hz, 1H), 2.24 – 2.07 (m, 3H), 2.06 – 1.99 (m, 2H), 1.70 (s, 3H). <sup>13</sup>C-NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  153.9 (C<sub>q</sub>), 140.0 (C<sub>q</sub>), 139.4 (C<sub>q</sub>), 138.4, 124.4, 124.1, 123.4, 122.4, 119.1, 115.1 (CH<sub>2</sub>), 74.4 (C<sub>q</sub>), 43.5 (CH<sub>2</sub>), 30.7, 28.8 (CH<sub>2</sub>). **HRMS** (EI+): calculated m/z for C<sub>14</sub>H<sub>16</sub>OS [M<sup>++</sup>] 232.09164; found 232.09159.

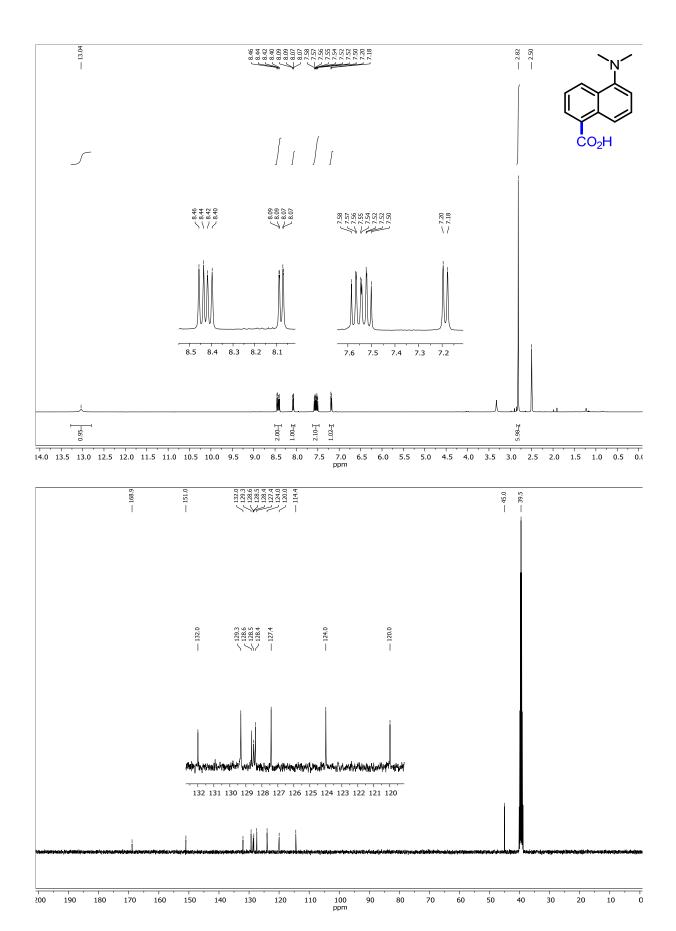
# 5 - Spectra

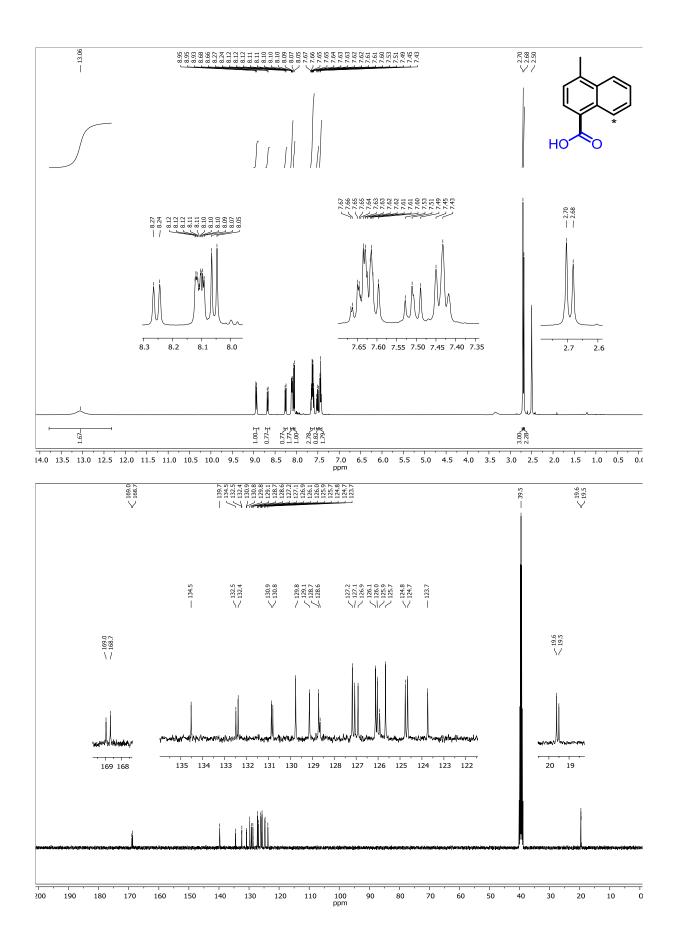


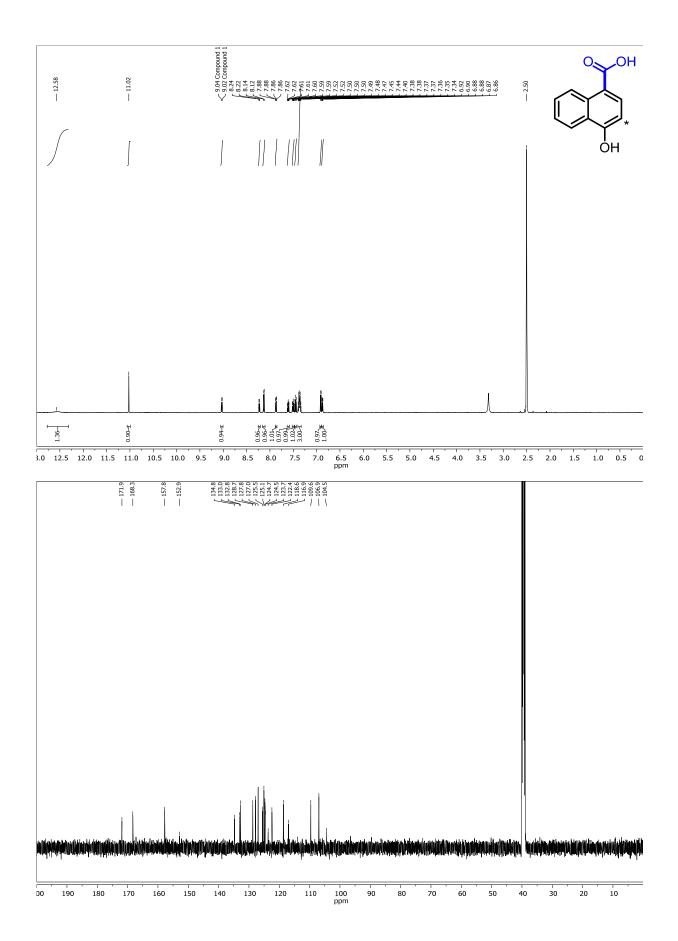


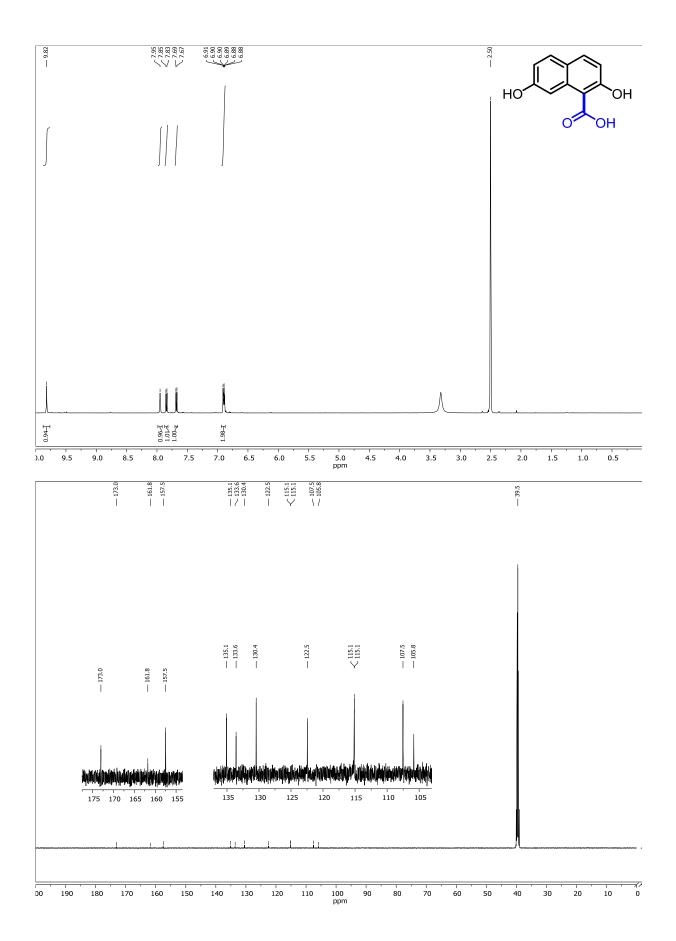


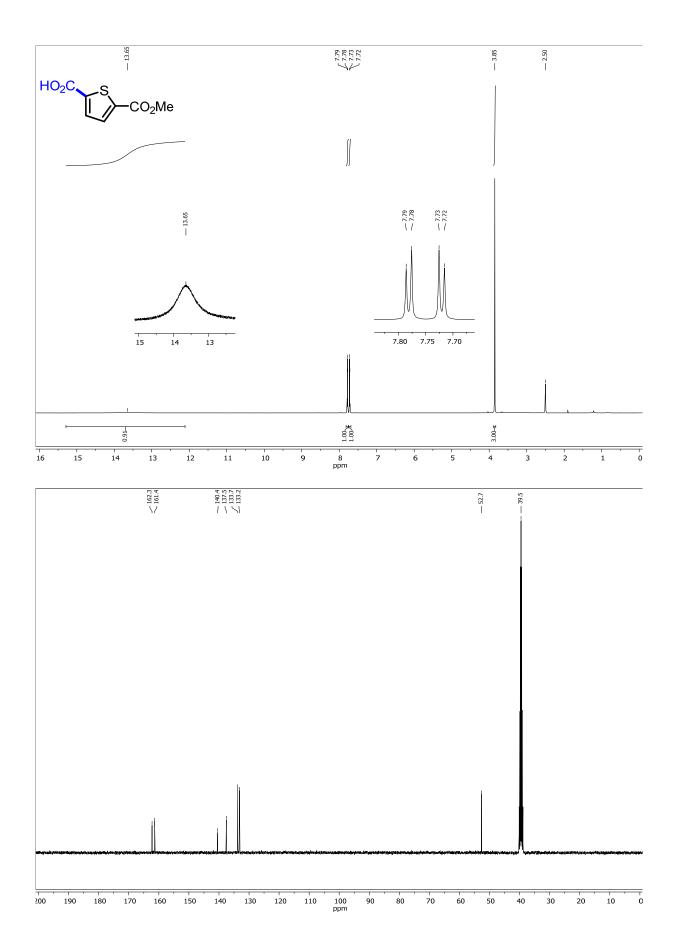


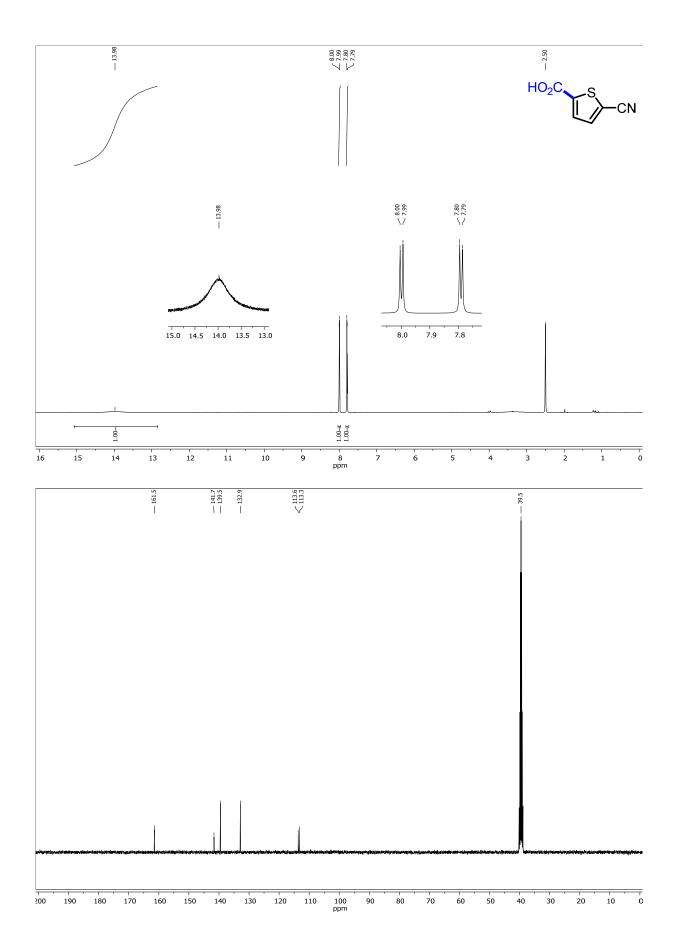


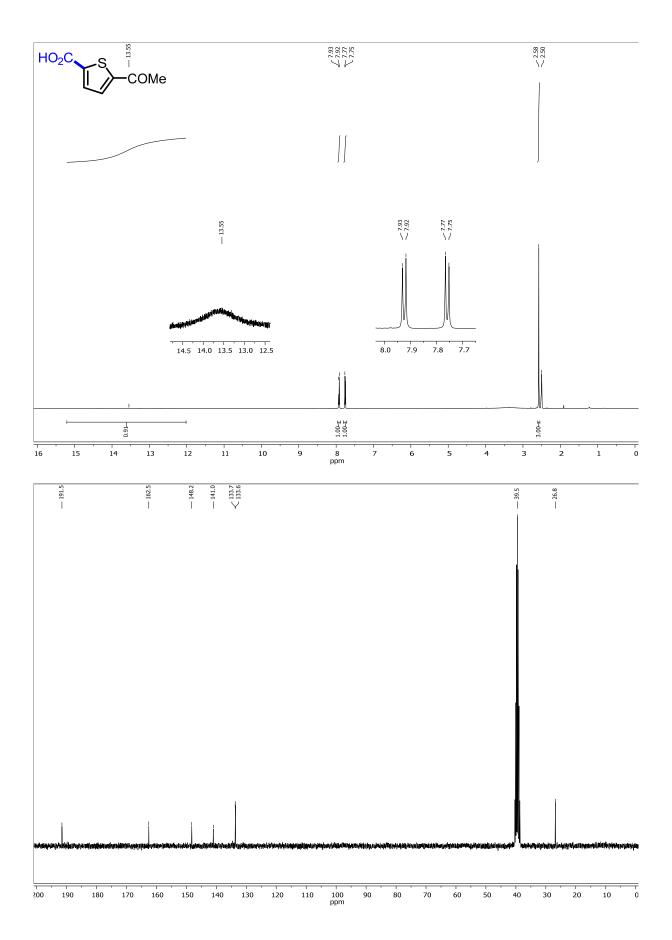


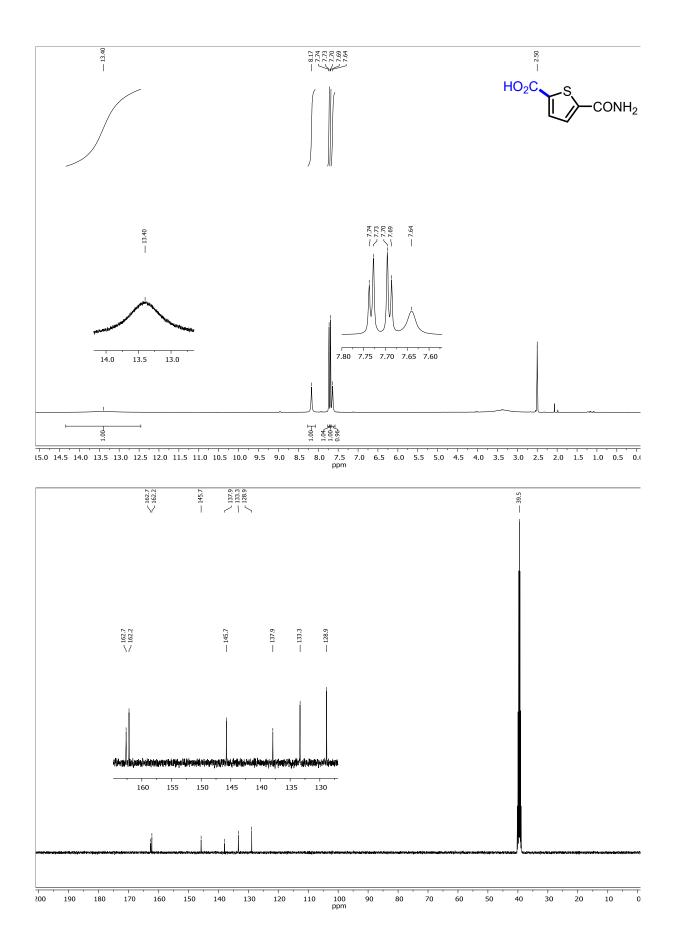


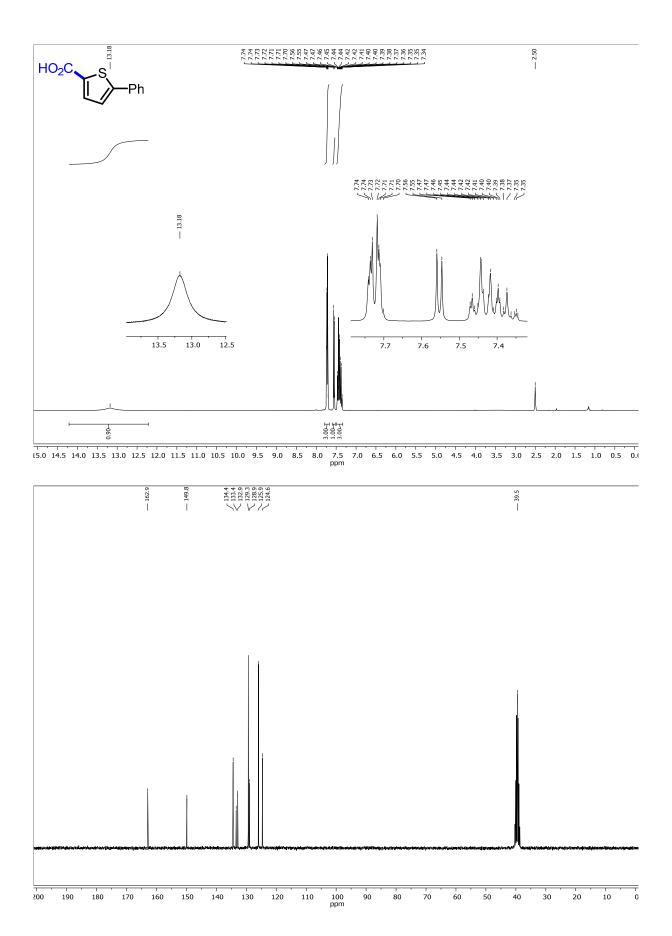


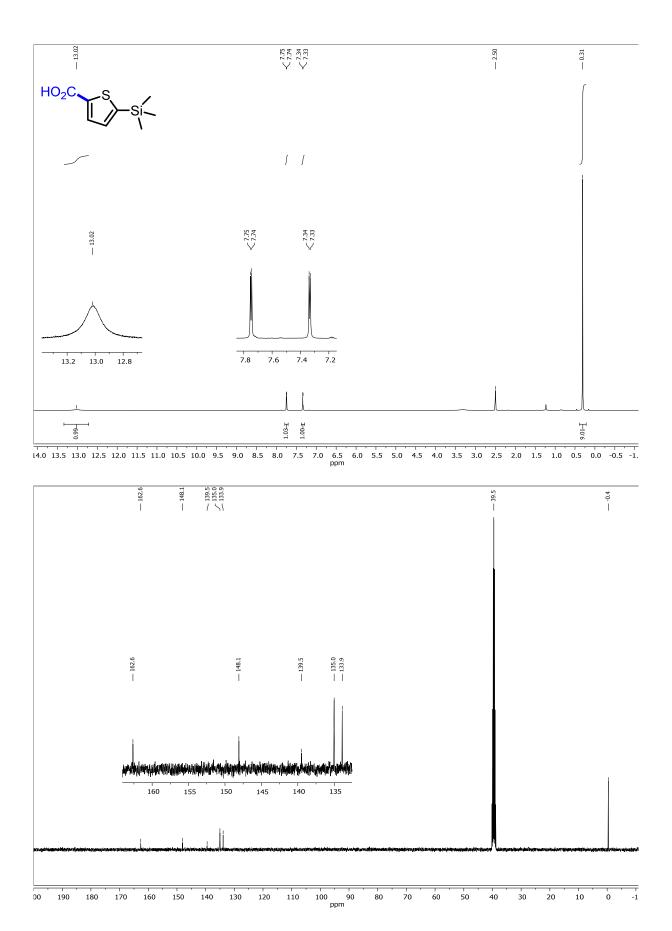


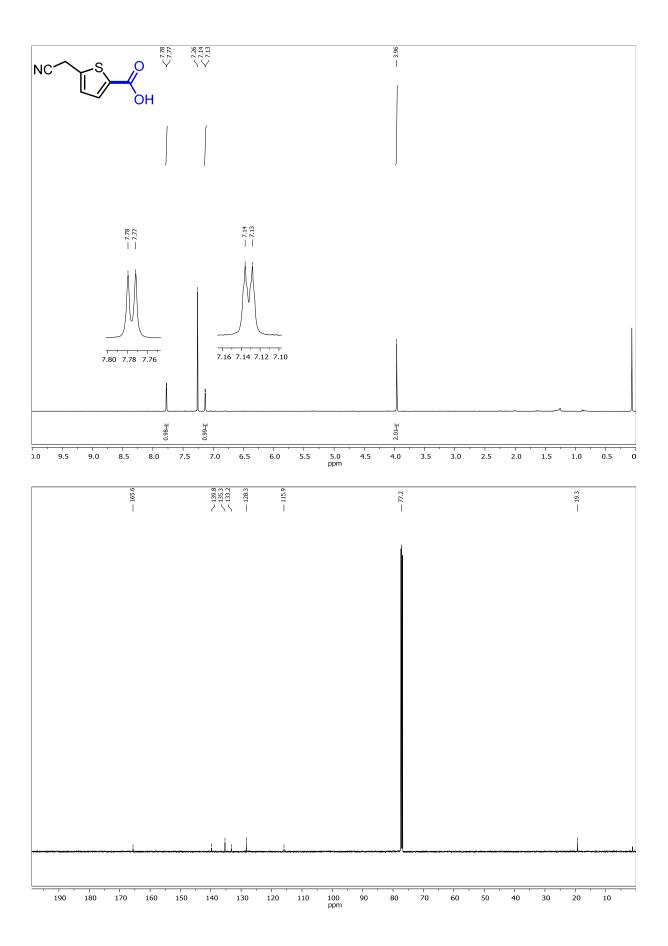


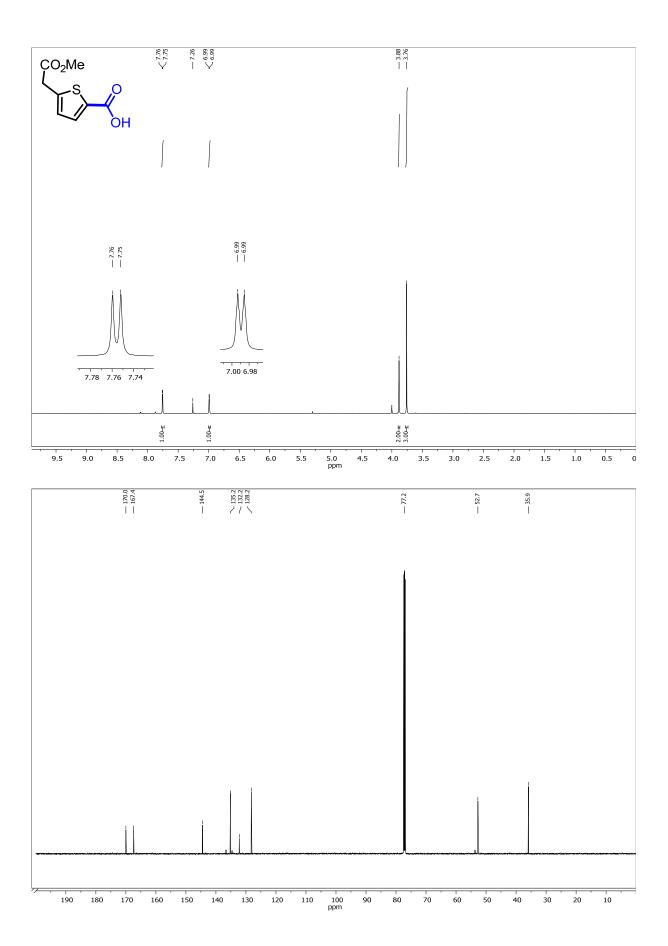


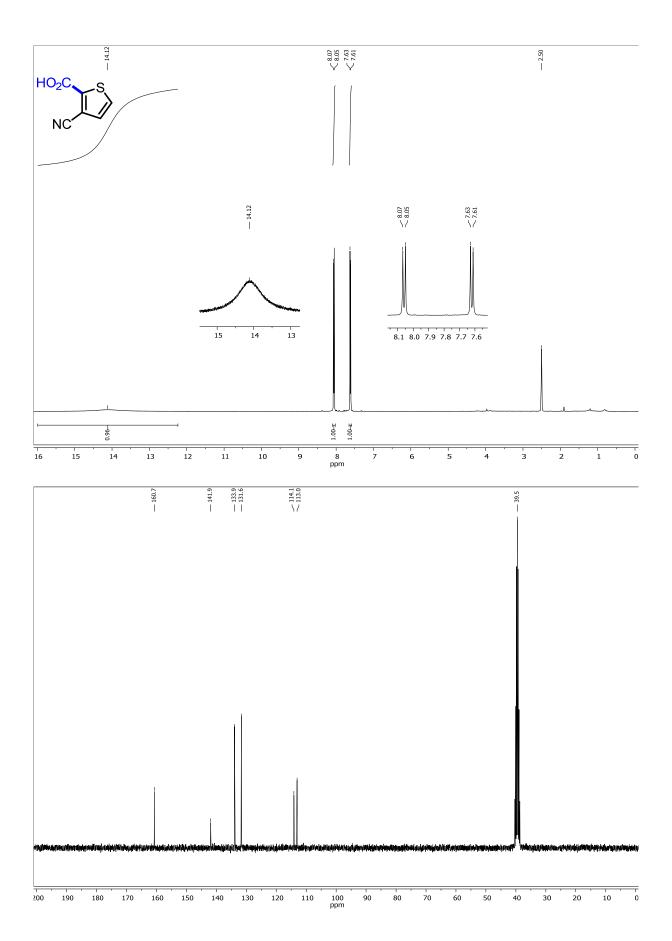


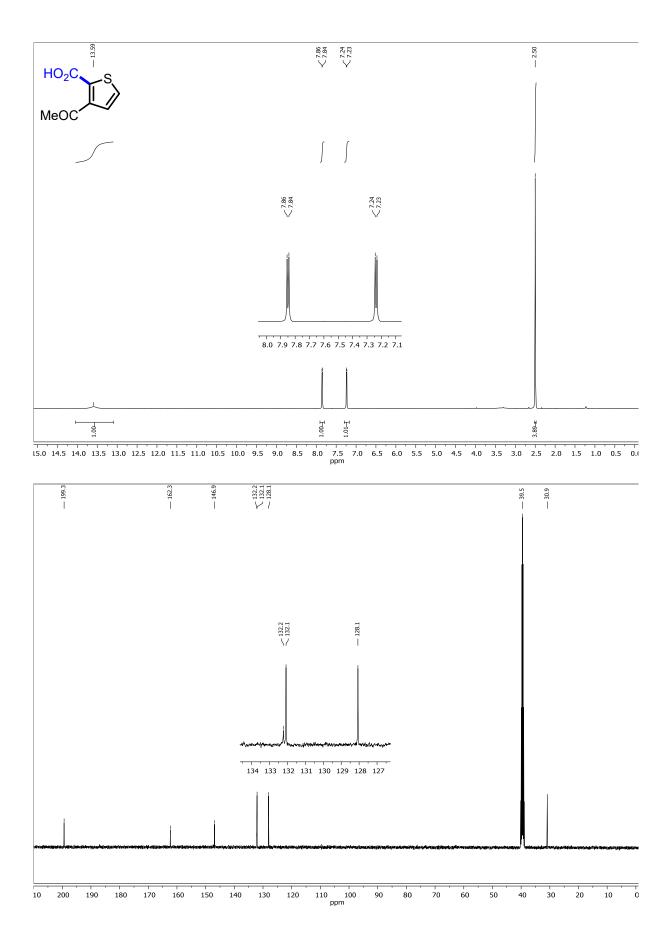


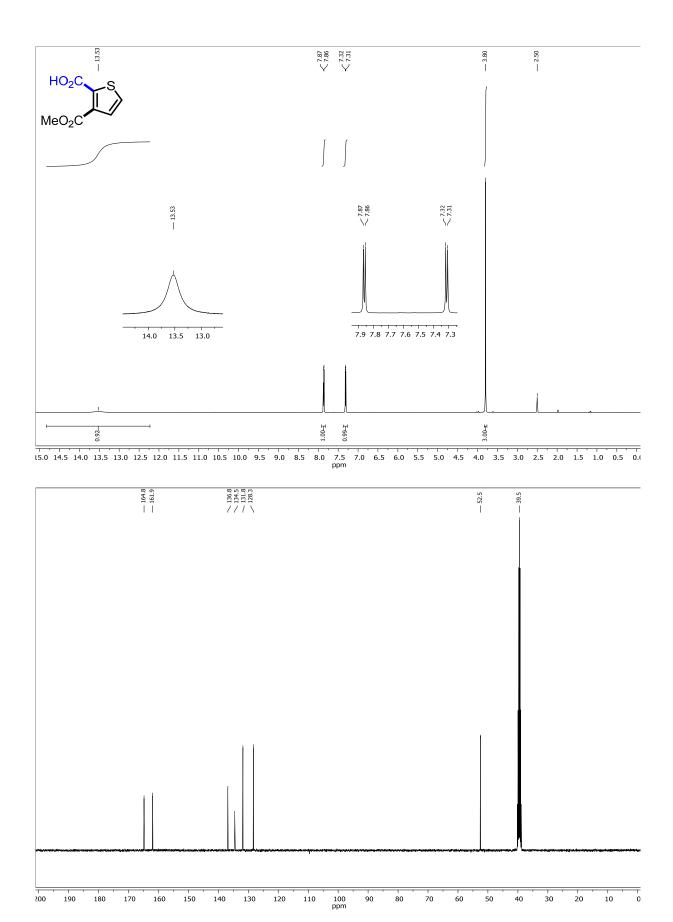


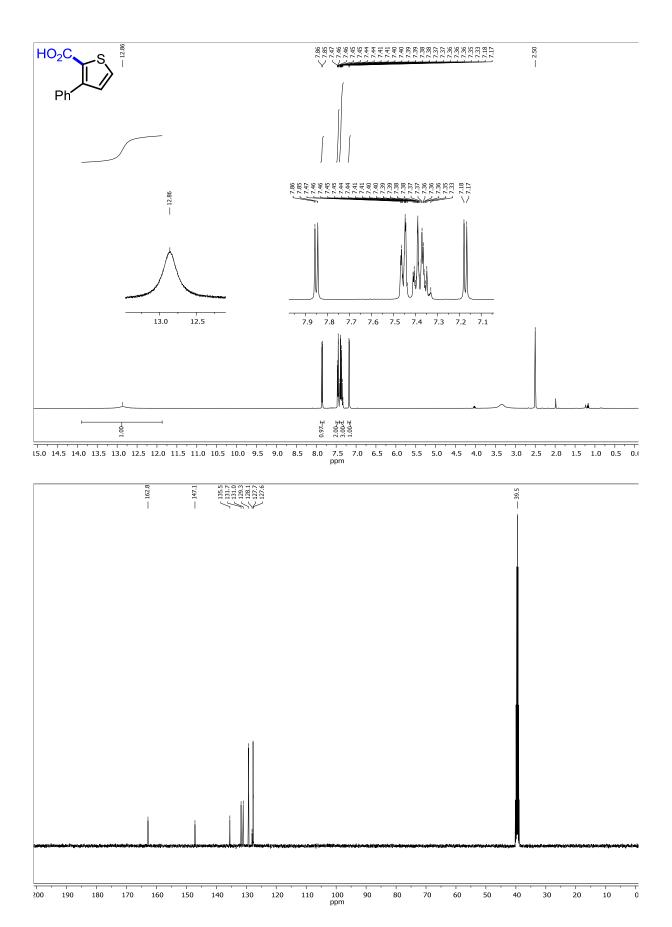


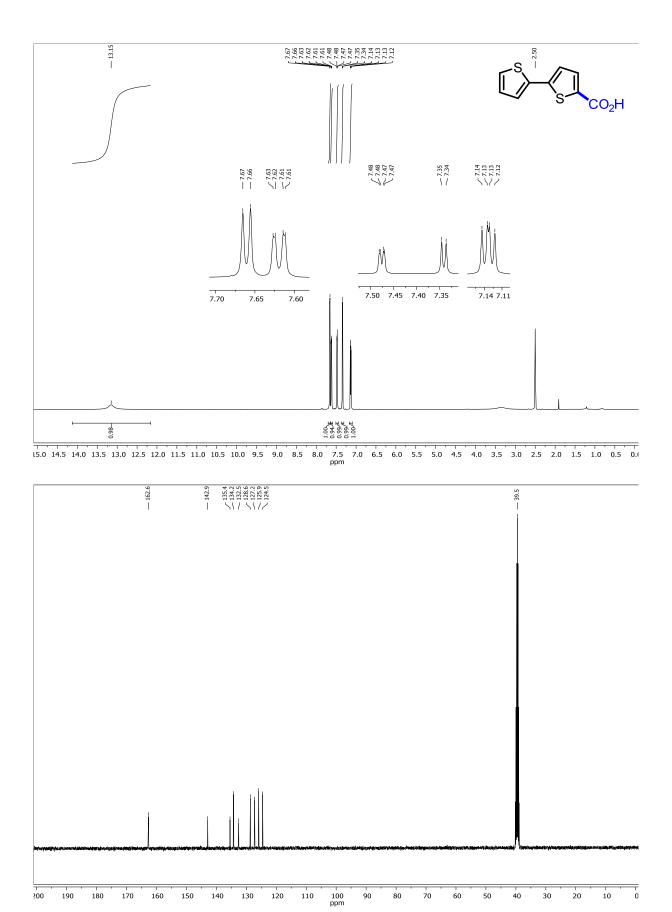


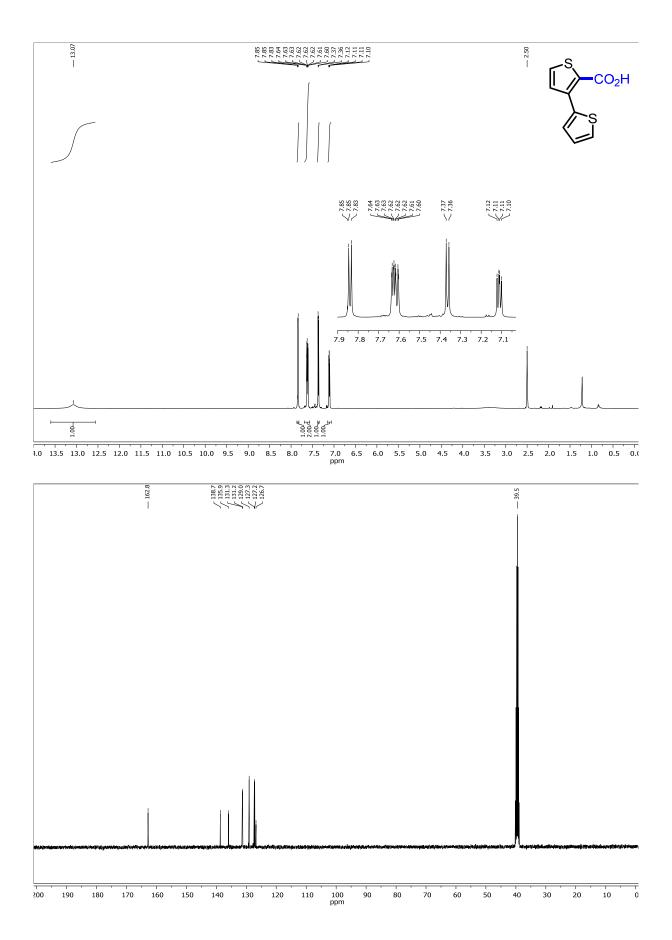


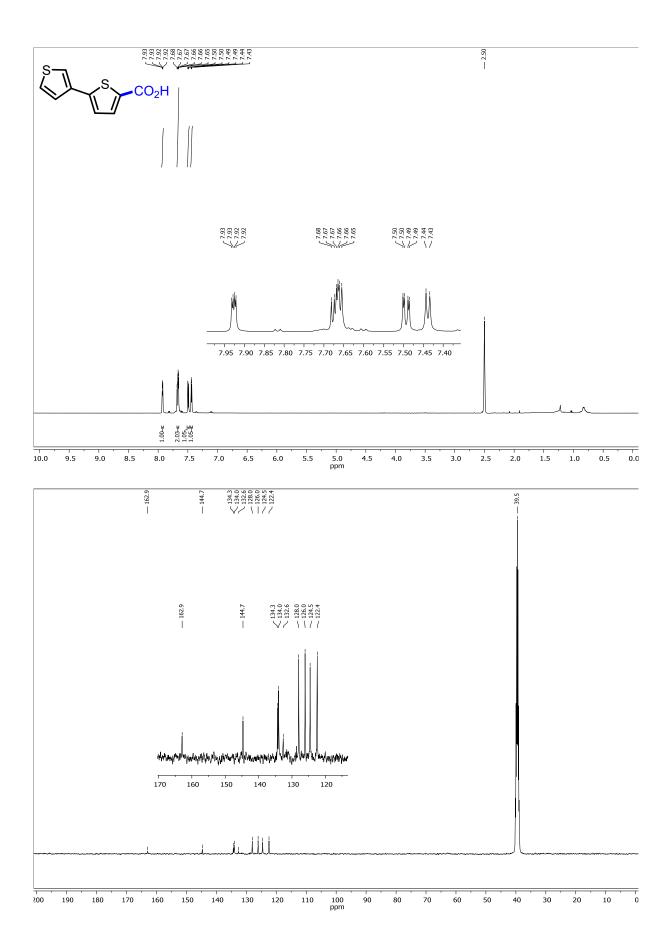


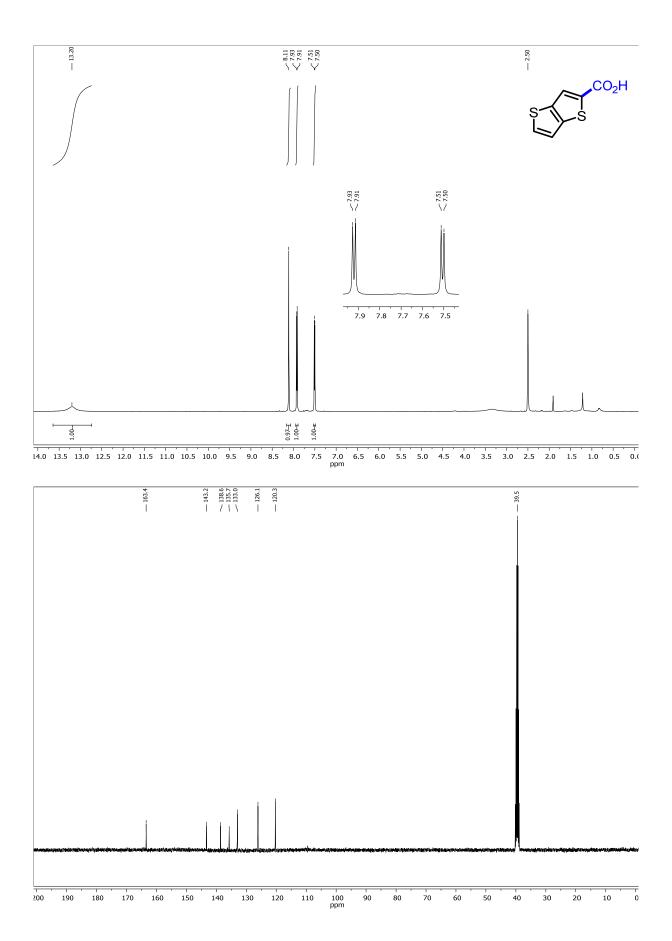


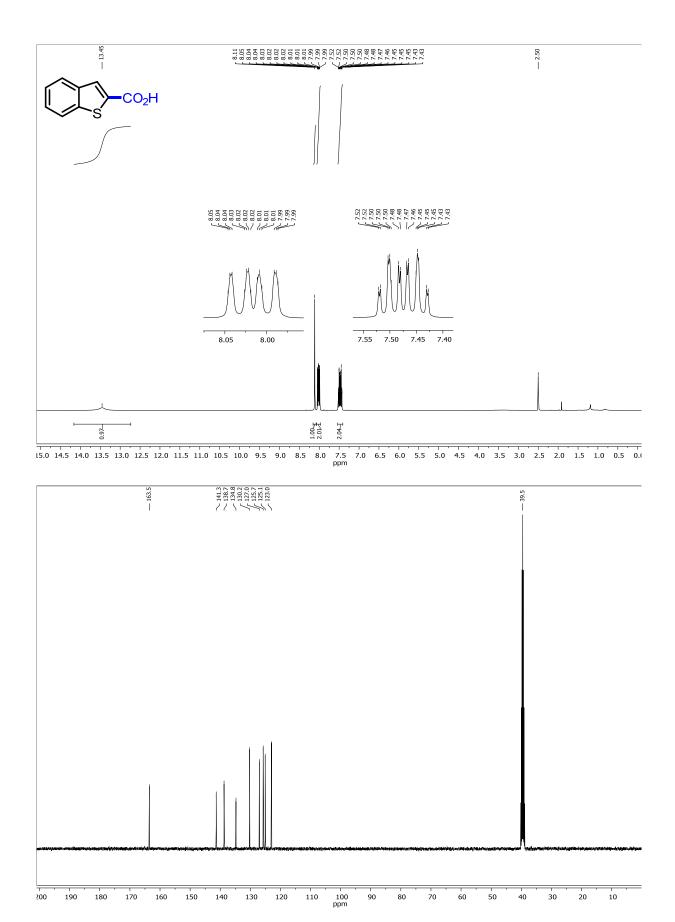


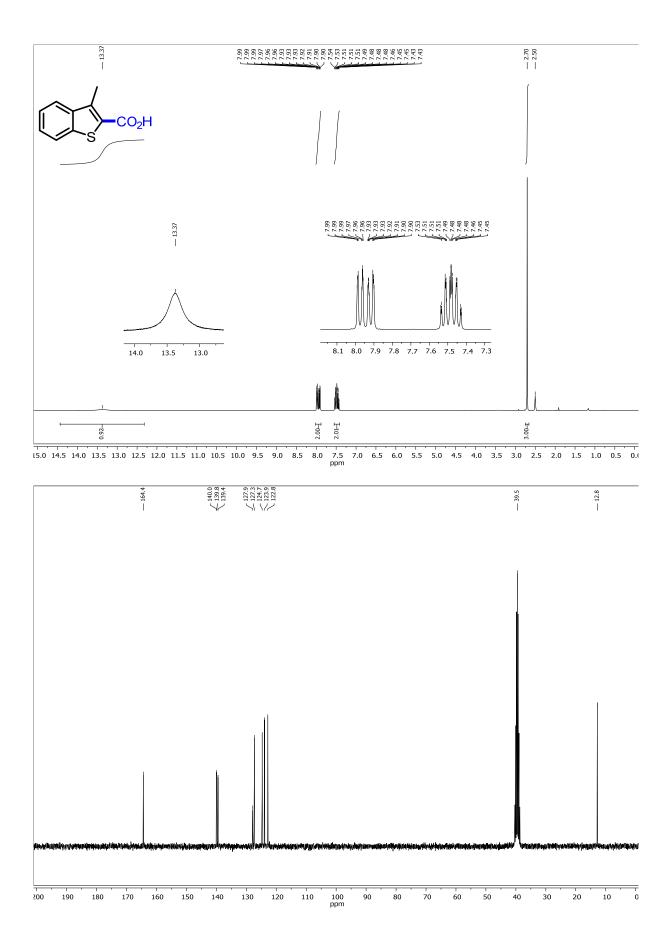


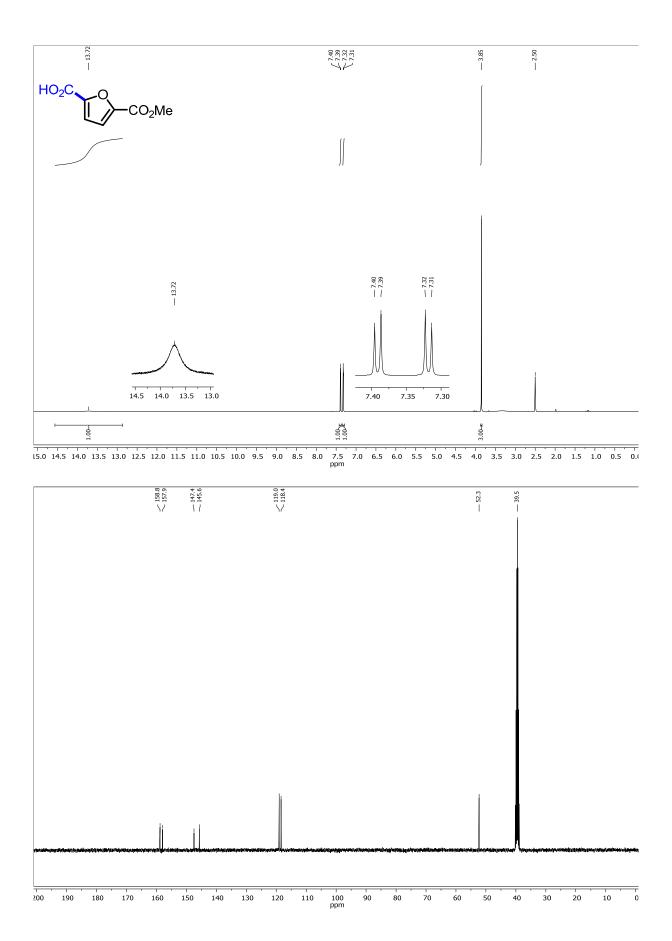


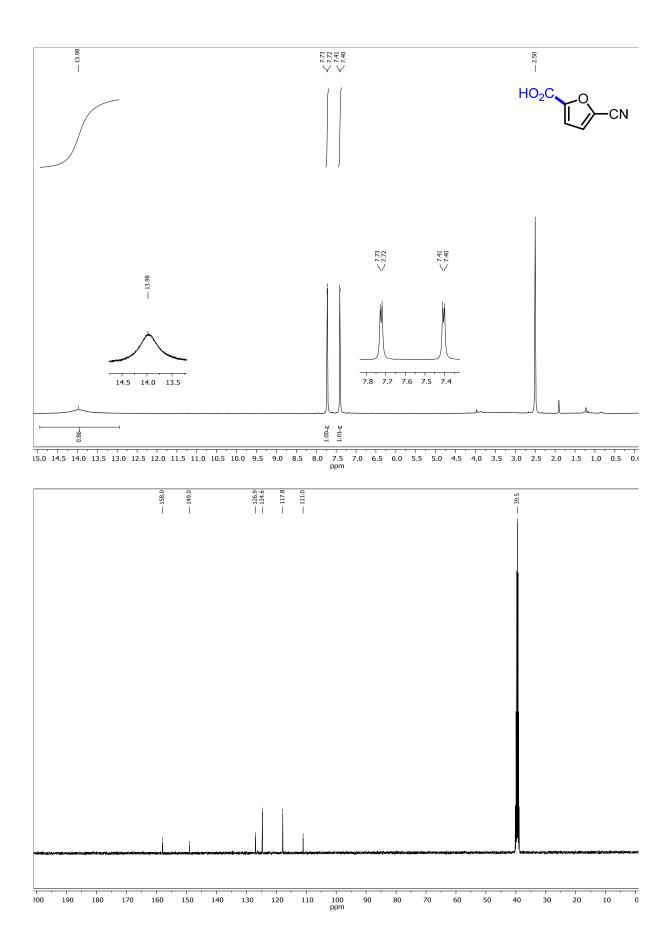


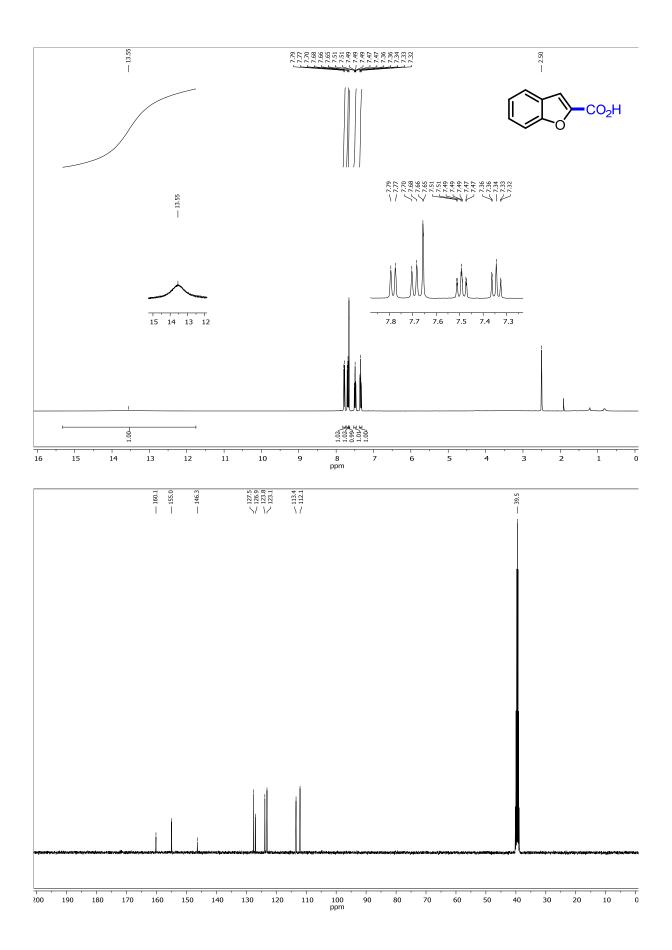


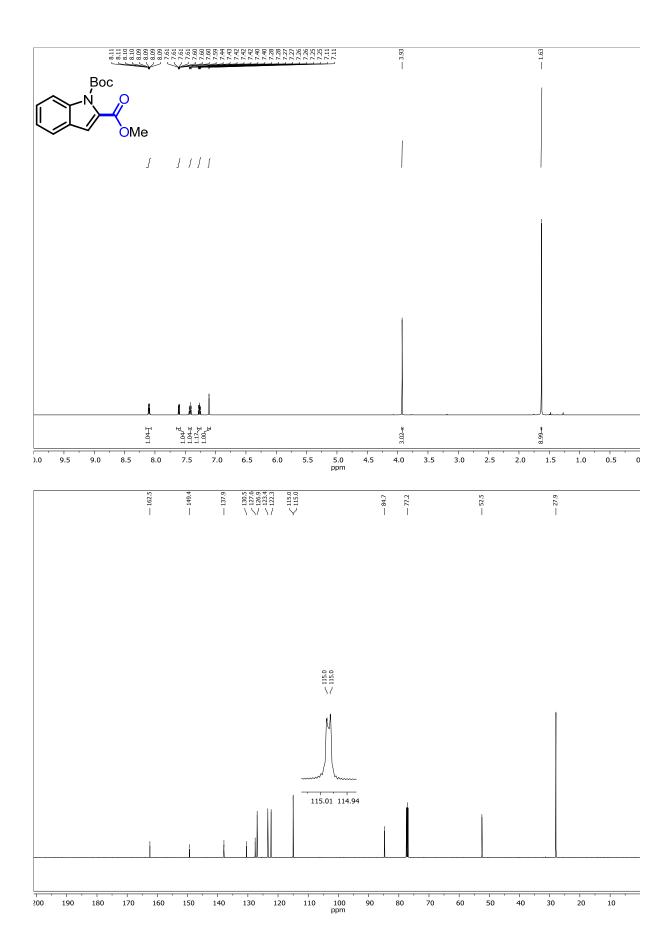


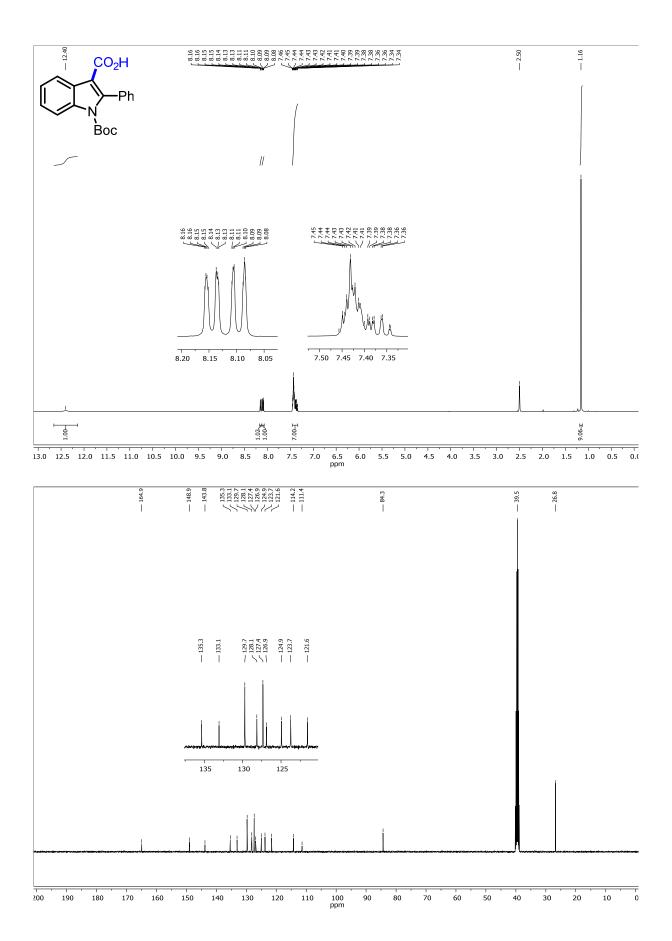


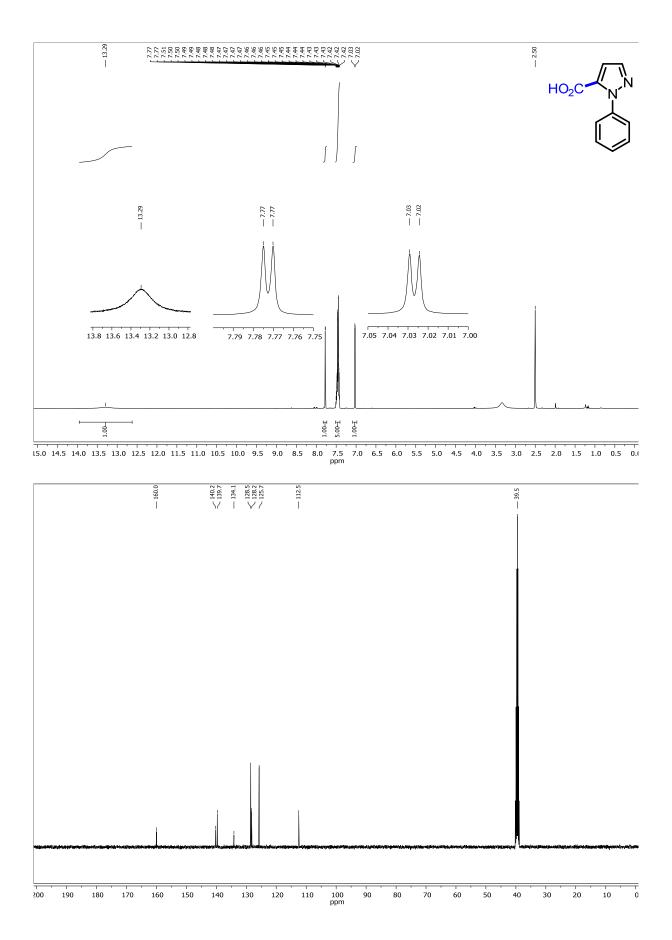


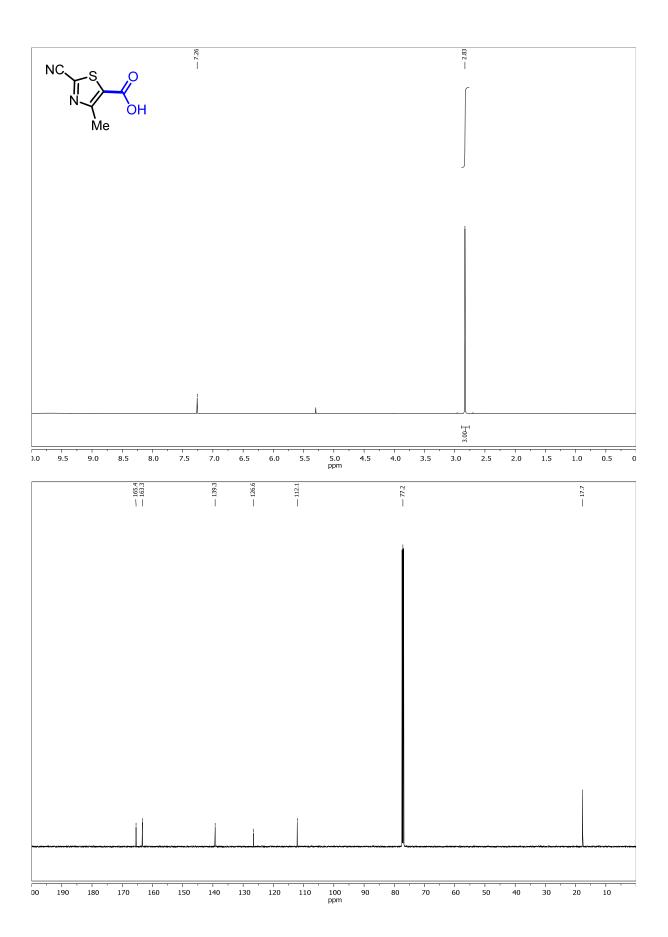


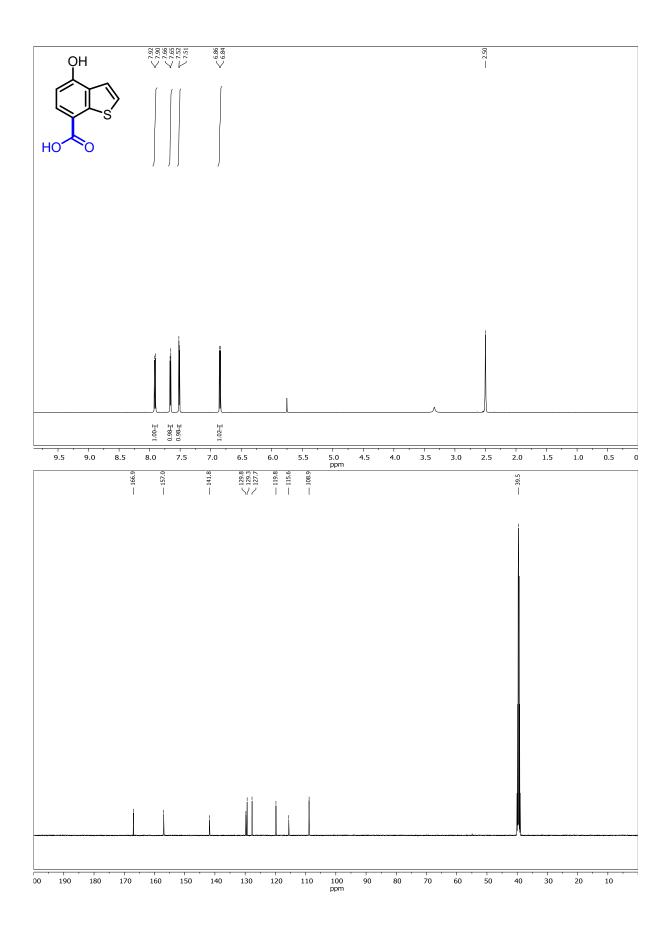


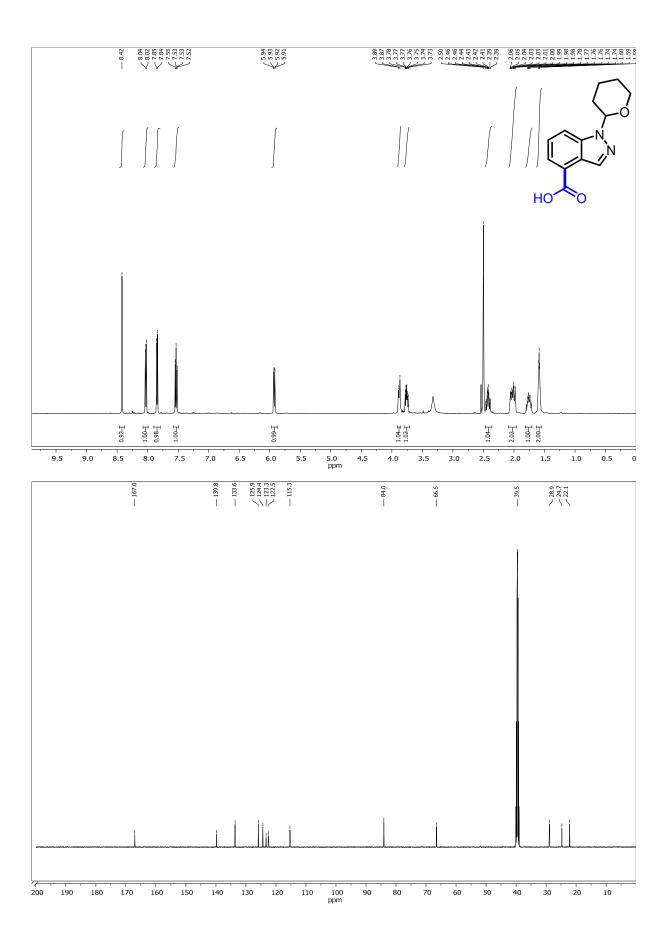


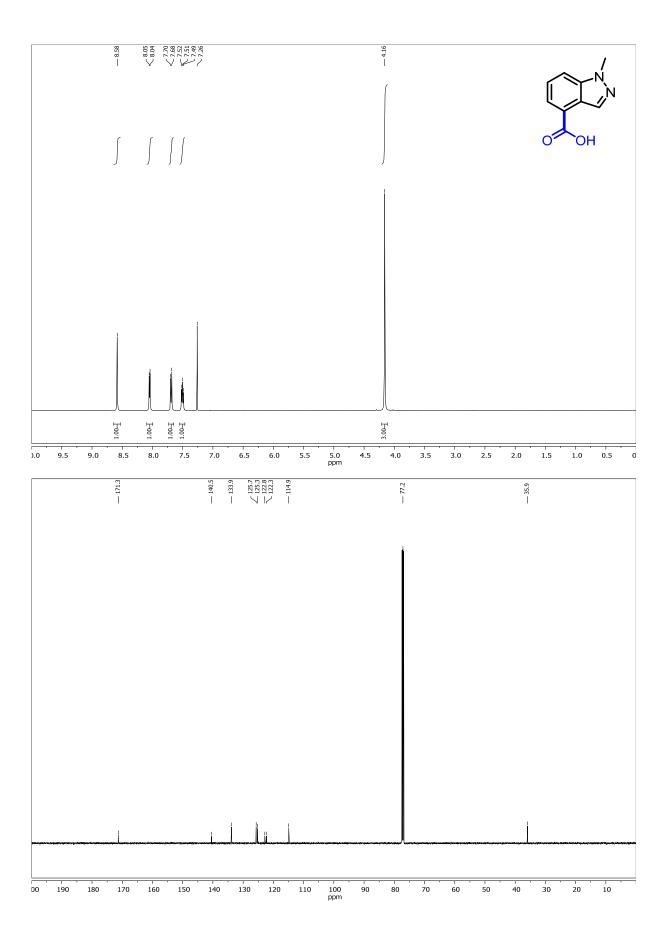


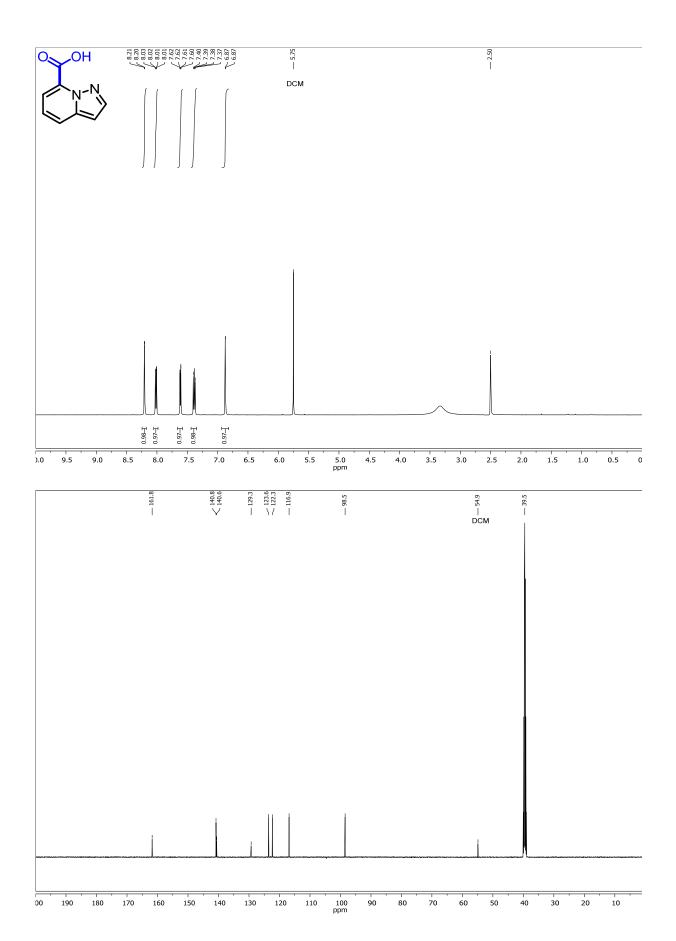


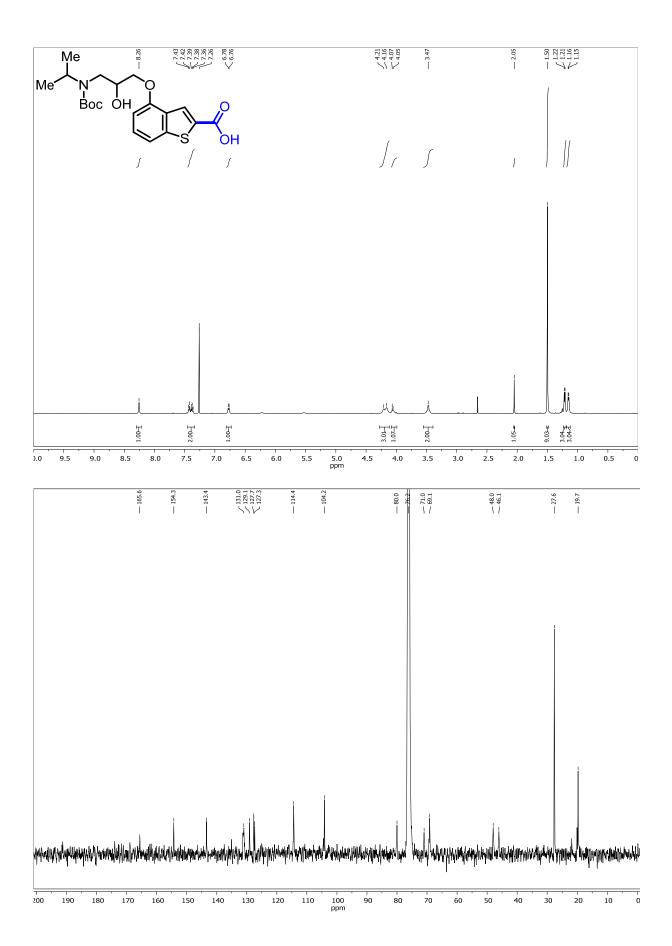


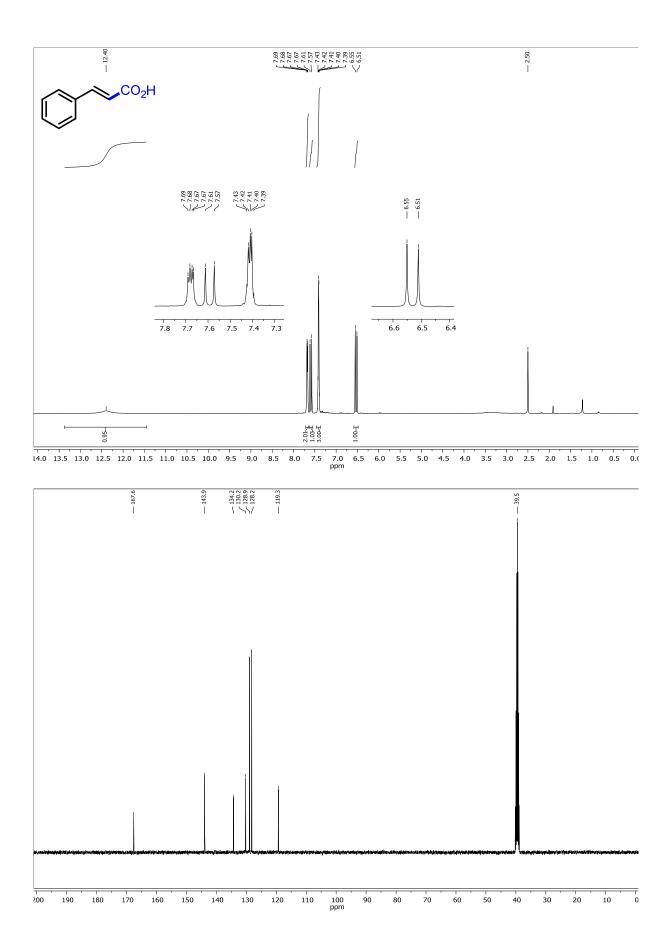


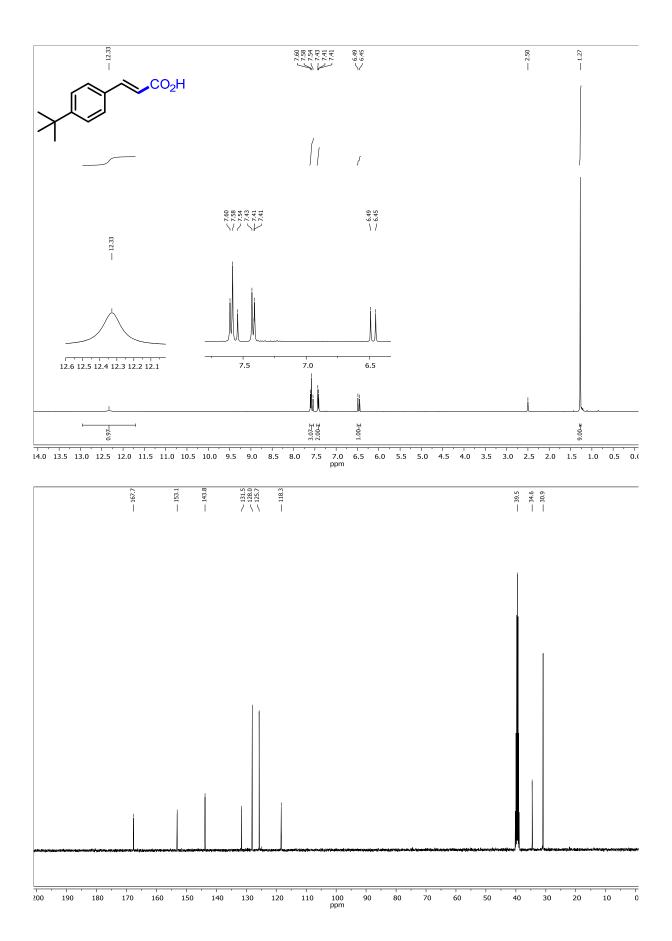


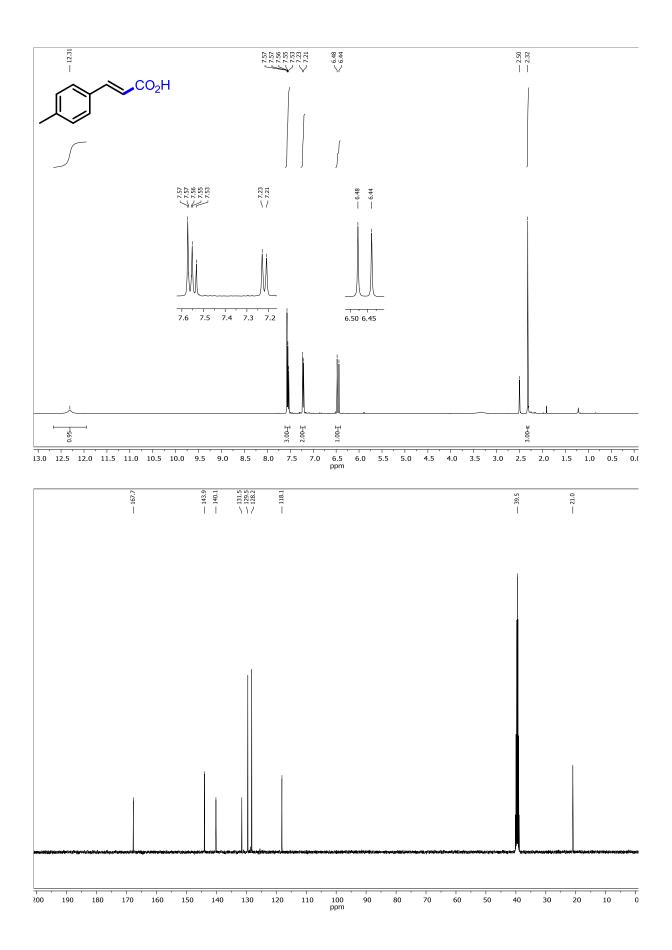


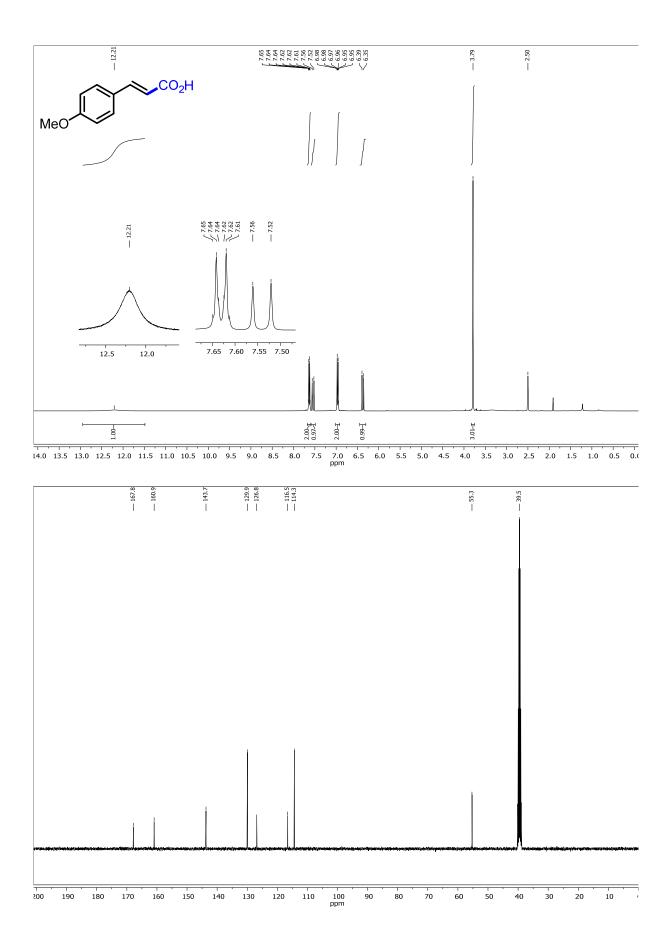


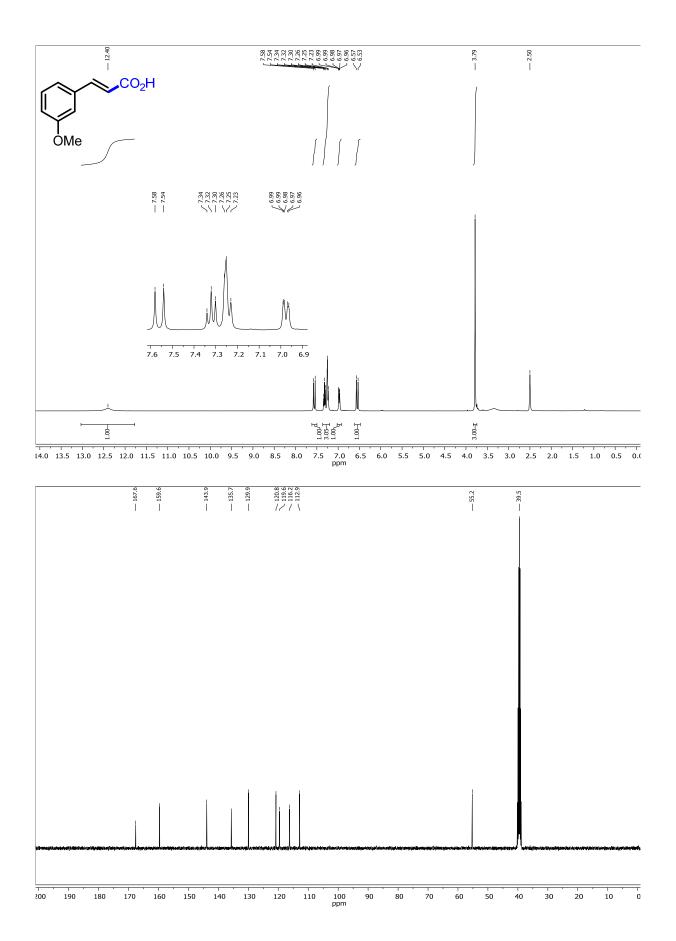


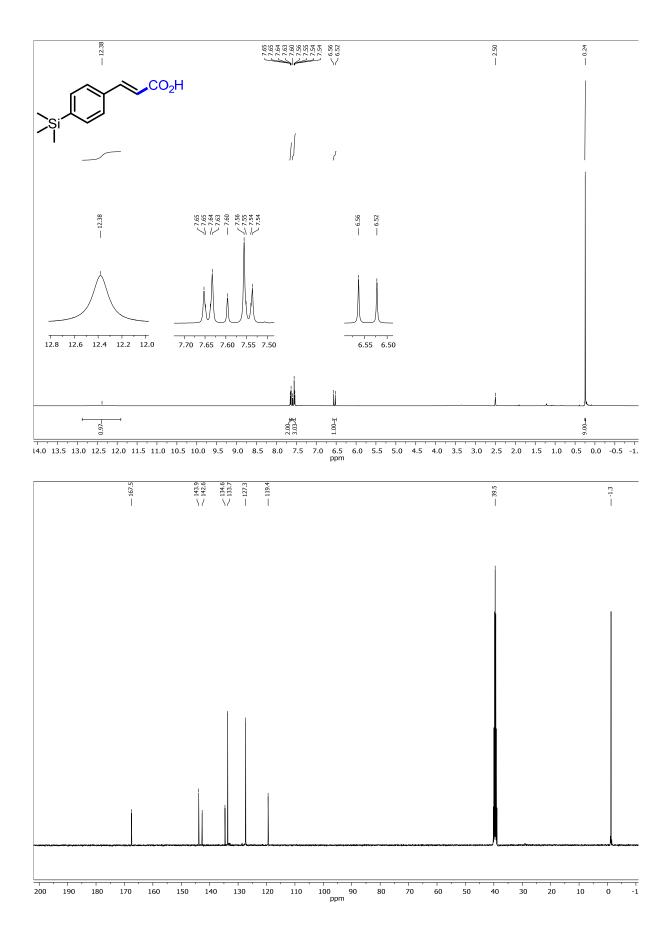


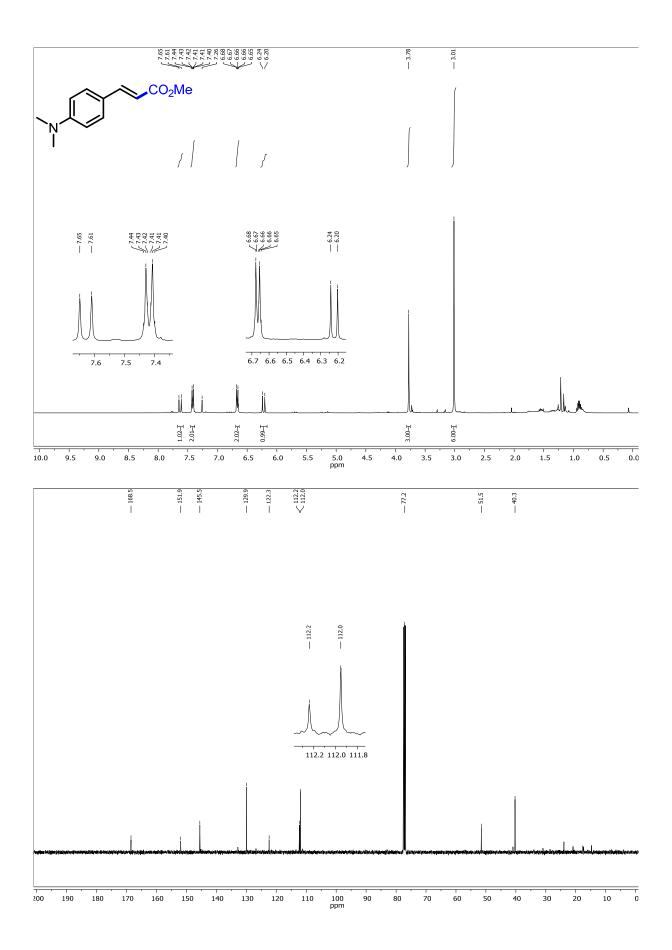


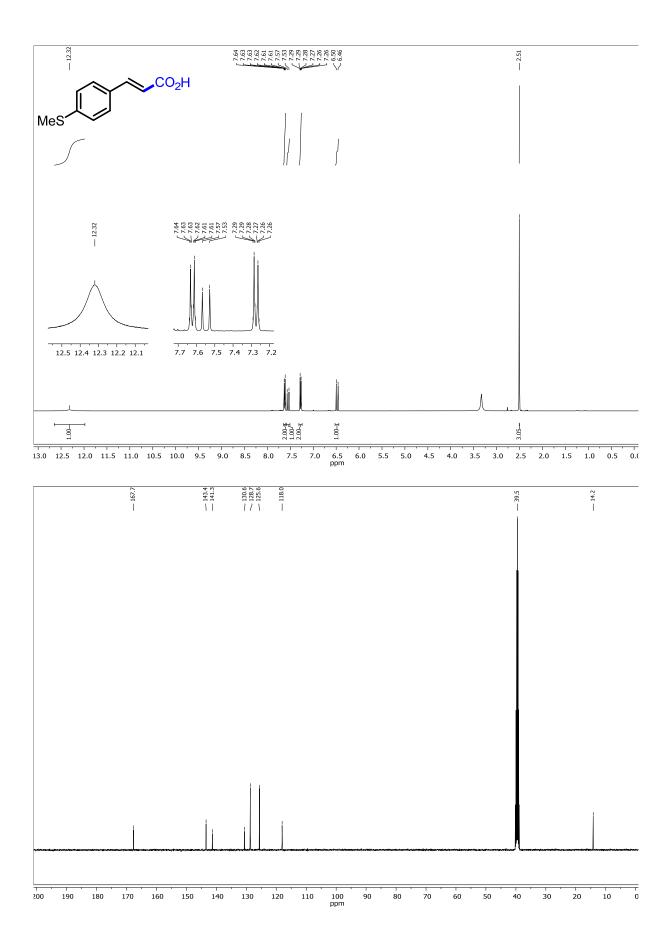


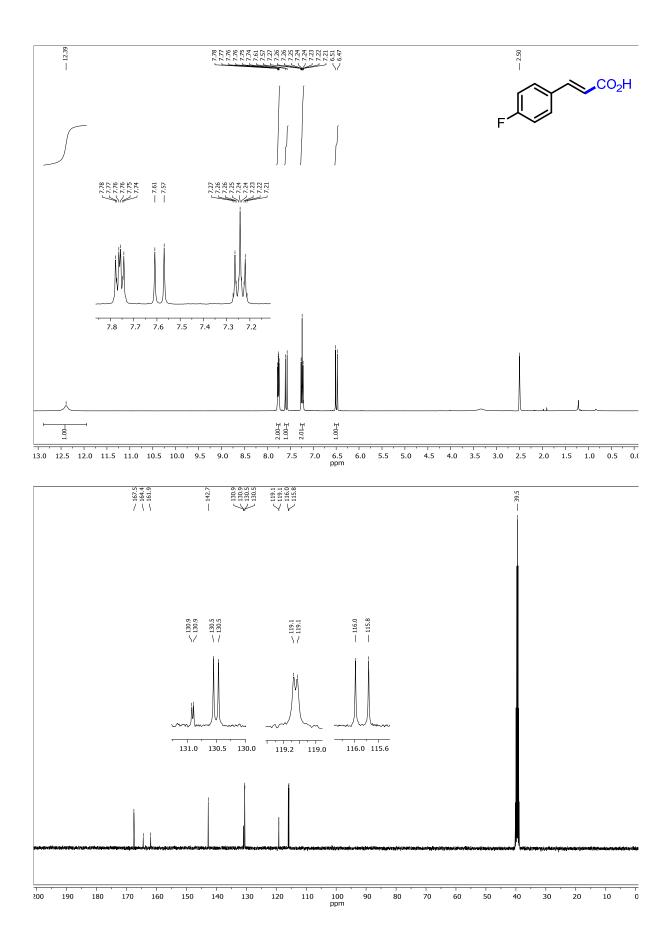


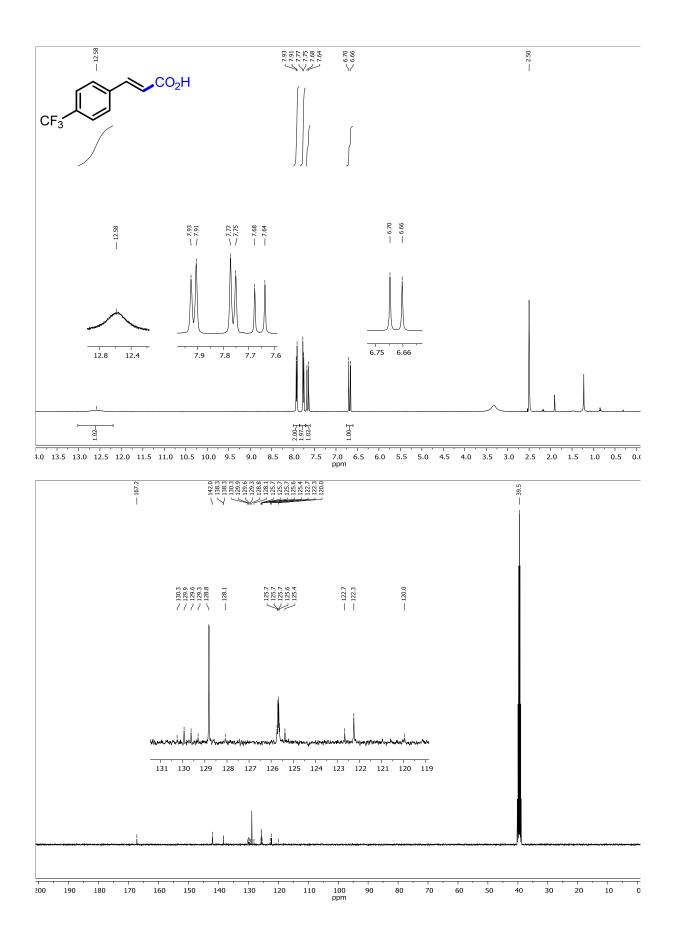


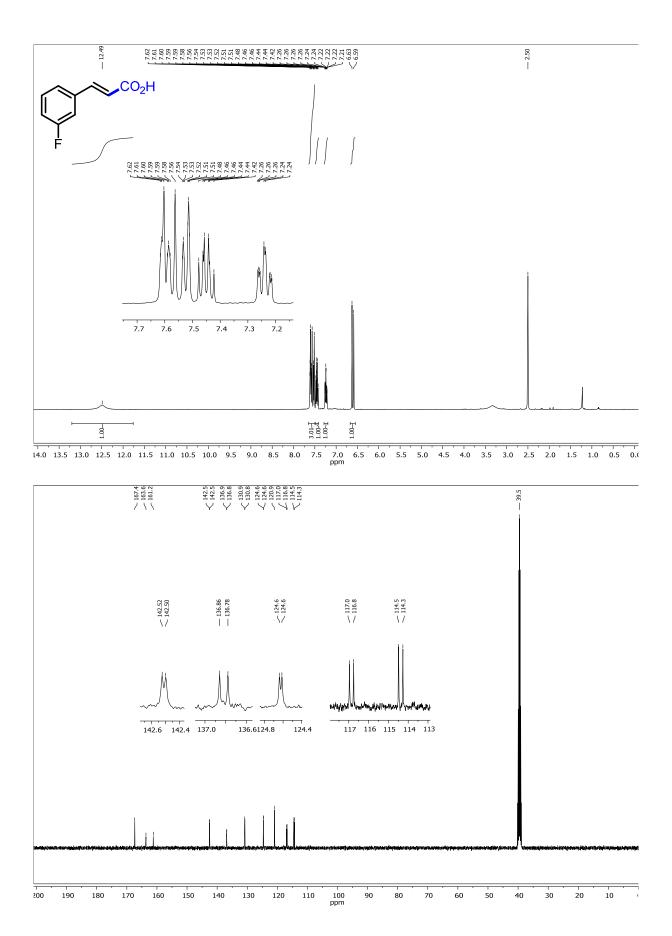


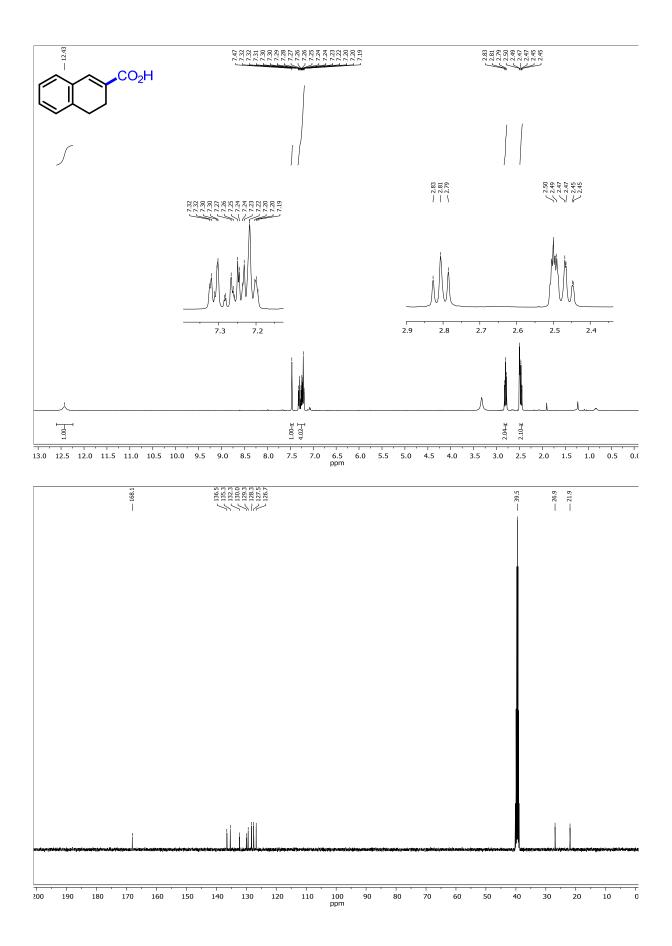


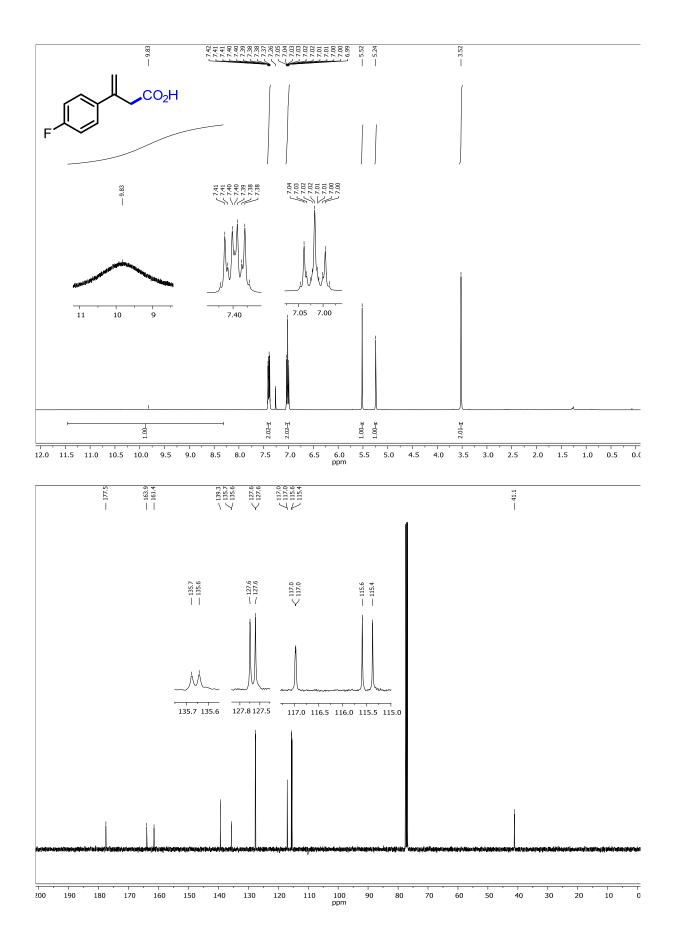


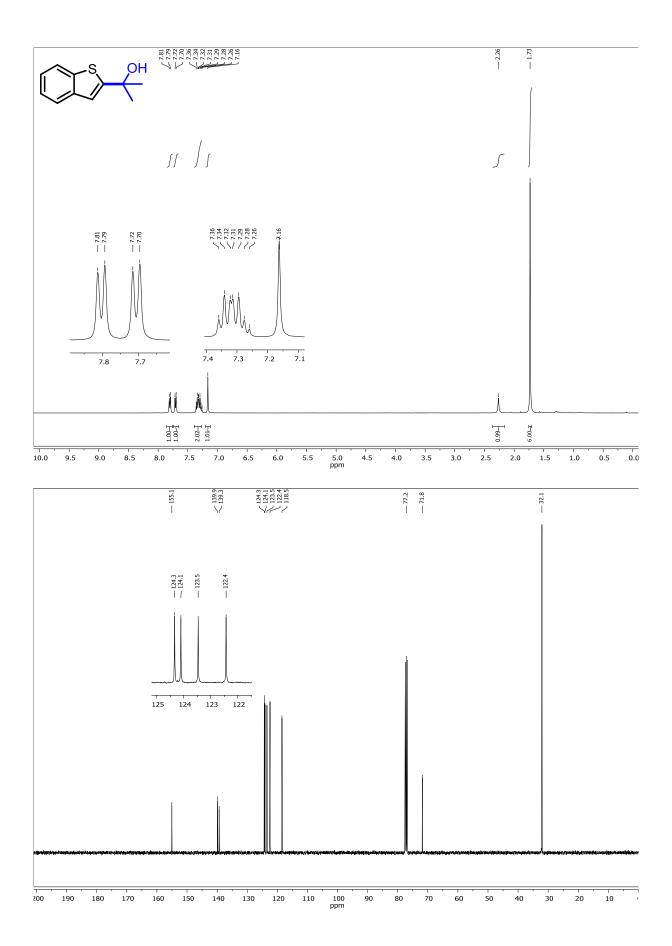


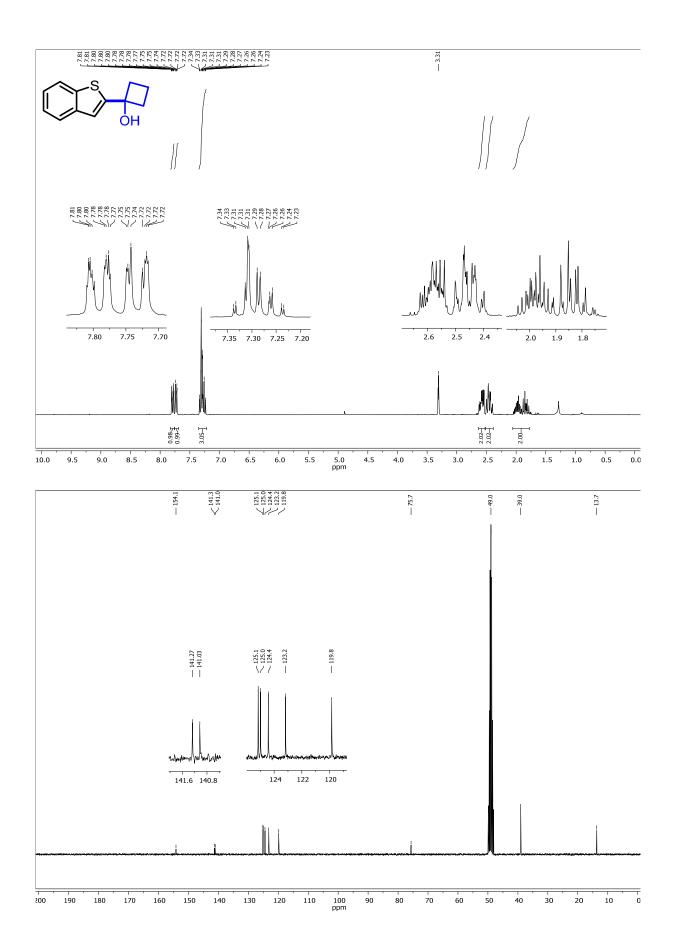


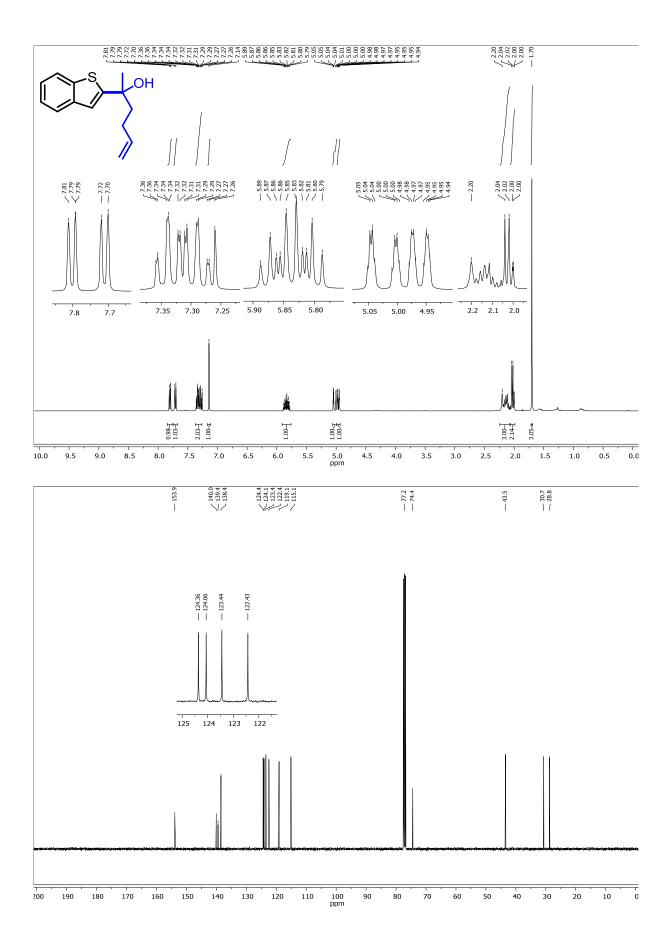












#### 6 - Mechanistic Studies

## 6.1 - General Procedure for High-throughput Screening of Arenes

To a paradox 96-well plate fitted with 0.5 mL glass vials containing magnetic stirrer bars was added the arene (0.03 mmol) and 2,3,6,7-tetramethoxyanthracen-9(10H)-one (2.0 mg, 20 mol%). The vials were partially-sealed and transferred to a glovebox antechamber after the vial had evacuated and refilled with N<sub>2</sub> (3×) within the antechamber. DMSO (0.3 mL) was dispensed to each of the vials followed by 1,1,3,3-tetramethylguanidine (11  $\mu$ L, 0.09 mmol) and the plate was sealed with the plate lid, two rubber mats and a Teflon TFA film. The plate was then removed and transferred to a glove-bag filled with an overpressure of CO<sub>2</sub>. The plate was then unsealed and placed on a stirrer plate with 3×456 nm Kessil lamps clamped overhead. The vial was then irradiated from above by two kessil lamps (vials approximately 10 cm away from the light source). After 18 hrs the irradiation was stopped and the plate was removed from the glove bag and quenched with aq. HCl (0.2 mL, 0.5M) and samples were taken and filtered before being placed on a plastic 96 well plate for HPLC analysis.





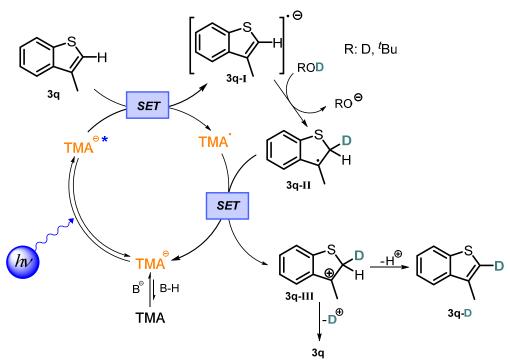
**Figure S9. a)** photo showing the removal of the plate lid within the CO<sub>2</sub> glove-bag for mass screening of substrates. **b)** 96 well plate reactions running in the CO<sub>2</sub> glove-bag set up.

#### **6.2 - Deuterium Labeling Experiments**

Upon formation of an arene radical anion 3q-I we envisioned a H/D exchange reaction giving rise to 3q-II in presence of a deuterium source. After reoxidation and deprotonation 3q-D would be formed (Scheme S3).

To a dry flat-bottomed crimp vial (5 mL) equipped with stirring bar, was added 3q (0.1 mmol) and 2,3,6,7-tetramethoxyanthracen-9(10*H*)-one (6.3 mg, 0.02 mmol, 20 mol%, only for entry 1-3 Table S3). Cs<sub>2</sub>CO<sub>3</sub> (98 mg, 3 equiv.) was quickly added and the vial was sealed with a Supelco aluminium crimp

seal with septum (PTFE/butyl). The vial was then evacuated and refilled with N<sub>2</sub> (5×) *via* syringe needle. The reaction mixture was dissolved in DMSO-d6 (1 mL, dry and degassed by bubbling with N<sub>2</sub>) and the deuterium source was added *via* syringe. The vial was then irradiated from the bottom side with blue LED light and a constant reaction temperature (25°C) was maintained by employing a water-cooling circuit connected to a thermostat. After 18 hrs of reaction time the reaction was quenched by the addition of water and the crude mixture was extracted with Et<sub>2</sub>O (3×). The combined organic layers were dried over Na<sub>2</sub>SO<sub>4</sub>, concentrated and purified *via* flash silica column chromatography using a mixture of hexanes and DCM (95:5) as eluent. The obtained product was dried in vacuo and analyzed by <sup>1</sup>H-NMR (Table S3 and Figures S10a-b) and GC-MS (Figure S10c).

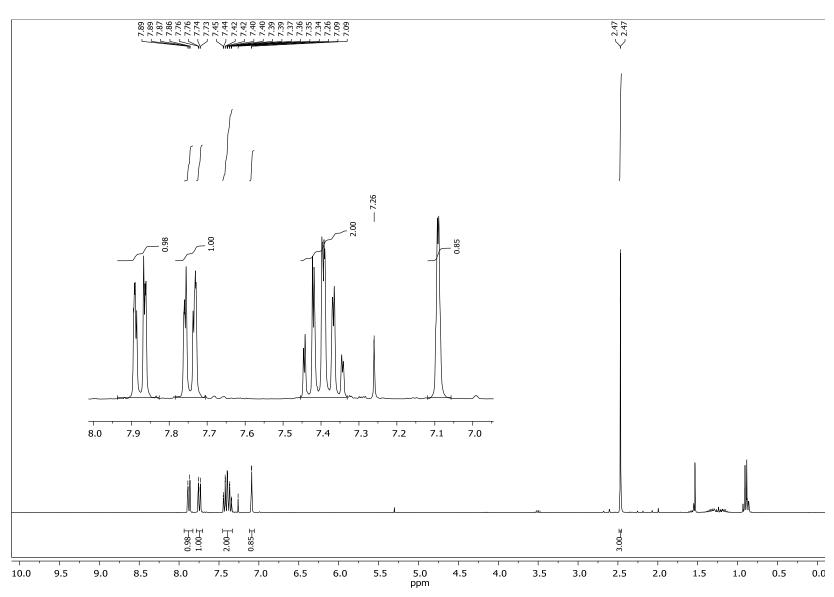


Scheme S3. Proposed mechanism for the deuterium labeling experiments using 3q as substrate.

Table S3. Deuterium labeling experiments

entry	catalyst	λ [nm]	D-source (eq.)	D-incorporation [%] <sup>a</sup>
1	TMAH	455	$D_2O(3)$	10
2	TMAH	455	$D_2O(15)$	14
3	TMAH	455	'BuOD (10)	15
$4^b$	-	dark	$D_2O(15)$	<1

<sup>&</sup>lt;sup>a</sup> determined by <sup>1</sup>H-NMR integration upon isolation and purification of the reaction mixture; <sup>b</sup> reaction was stirred in the dark.



**Figure S10a.** <sup>1</sup>H-NMR recorded after reaction work-up and column chromatography according to entry 3, Table S3. The signal at 7.1 ppm corresponds to the proton in position 2 of **3q**. Peak integration revealed a slightly reduced value of 0.85. The signal at 7.75 ppm served as reference and was set to integral 1.00.

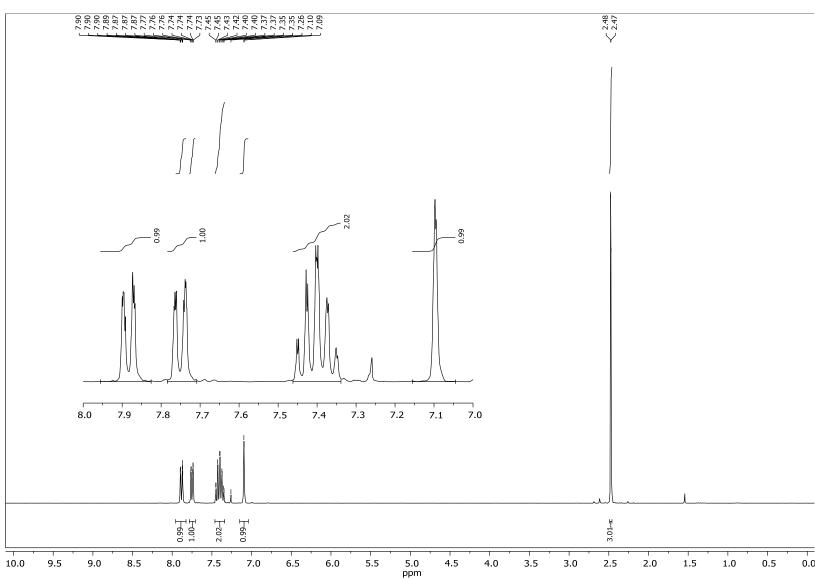
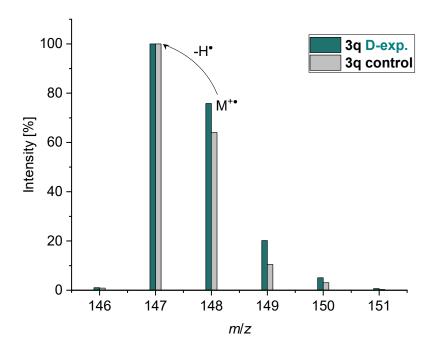


Figure S10b. <sup>1</sup>H-NMR recorded after reaction work up and column chromatography according to entry 4, Table S3. The signal at 7.1 ppm corresponds to the proton in position 2 of 3q. Peak integration revealed a value of 0.99. The signal at 7.75 ppm served as reference and was set to integral 1.00.



**Figure S10c.** GC-MS analysis after reaction work up and column chromatography according to entry 3 from Table S3 (green) and purchased 3q (grey). Mass spectra was recorded upon electron impact ionization (70 eV). The ionized 3q is prone to lose a hydrogen atom causing the most intense peak at m/z 147.

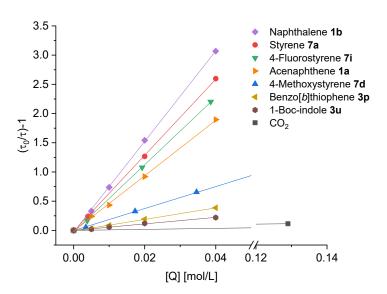
Figure S10a shows the <sup>1</sup>H-NMR spectrum from the isolated product of the deuterium labeling experiment in presence of <sup>1</sup>BuOD (entry 3, Table S3). Integration over the signal at 7.1 ppm, which corresponds to the proton in position 2 of benzothiophene **3q**, revealed a slightly decreased value (0.85 instead of 1.00). This deviance can be explained by the partial exchange of hydrogen by deuterium.

Figure S10b shows the <sup>1</sup>H-NMR spectrum from the isolated product of the control reaction (entry 4, Table S3). Integration over the signal at 7.1 ppm gave a value close to unity and suggests no or only traces of incorporated deuterium.

In addition to <sup>1</sup>H-NMR analysis, the incorporation of deuterium was verified by GC mass. Compared to the set of peaks caused by the purchased starting material **3q** (Figure S10c, grey), the different ratios in the isotope pattern suggest the partial incorporation of deuterium into the product isolated upon deuterium labeling reaction (entry 3, Table S3).

# 6.3 - Time-resolved Luminescence Quenching studies

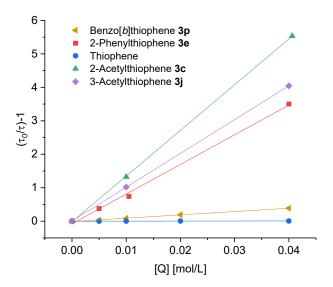
A first prediction regarding possibly working substrates was made by time-resolved luminescence quenching of the photoexcited catalyst. If there is an interaction between the excited PC and the substrate (electron transfer is proposed), the luminescence lifetime is shortened. Such processes can be easily followed by luminescence lifetime analysis and from the data obtained a Stern-Volmer plot of the time-resolved experiment was developed (Figure S11a-d and S13). A linear correlation between concentration of quencher [Q] and  $\tau_0 \times \tau^{-1}$  indicates a dynamic luminescence quenching. The luminescence lifetime was recorded in dry, degassed DMSO in presence of cesium carbonate by using a quartz cuvette (1×1 cm) with septum screw cap. The cuvette was degassed *in vacuo* and backfilled with N<sub>2</sub> (5×) before the stock solution of quencher and the catalyst solution were added *via* syringe. A **TMAH** concentration of c(TMAH) = 40  $\mu$ M in the cuvette was used for all experiments. For excitation of the sample, a 452 nm laser diode was used and an optical longpass filter (cut-on wavelength 500 nm) was installed before the detection unit. The time range for the measurement was set to 400 ns. The experimental data were fitted with a mono-exponential function. The quenching experiment using CO<sub>2</sub> as quencher was recorded as described above using a CO<sub>2</sub>-saturated DMSO solution. The approximated concentration of dissolved CO<sub>2</sub> was calculated from literature data. 41



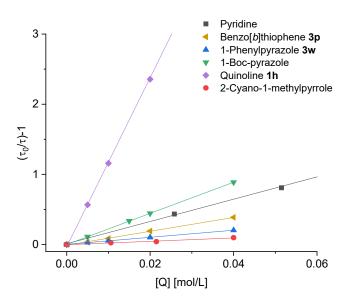
**Figure S8a.** Stern-Volmer plot developed with data obtained from time-resolved quenching experiments of **TMAH** in presence of Cs<sub>2</sub>CO<sub>3</sub> with tolerated substrates and a CO<sub>2</sub>-saturated solution of DMSO.

Thiophene derivatives 3c, e, j are excellent quenchers and are tolerated in the carboxylation reaction whereas benzo[b]thiophene (3p) was found to quench the excited state of  $TMA^-$ , however less efficiently. No quenching was observed in presence of thiophene and no carboxylation occurred when thiophene was used as substrate under the optimized reaction conditions (Figure S11b). The tested N-heteroarenes containing at least one nitrogen are quenching the excited photocatalyst. However, carboxylation products were only obtained using 3u (Figure S11a) or 3w (Figure S11c). Using acetone

**9a** as electrophile instead of CO<sub>2</sub> gave rise to the respective tertiary alcohol **9pa**. Quenching studies revealed that adding acetone (up to 2000 eq. regarding to catalyst concentration) does not quench the photoexcited **TMA**<sup>-</sup> (Figure S11d), supporting the hypothesis of a nucleophilic arene radical anion which attacks the electrophile.



**Figure S9b.** Stern-Volmer plot developed with data obtained from time-resolved quenching experiments of **TMAH** in presence of Cs<sub>2</sub>CO<sub>3</sub> with thiophene derivatives. In case of thiophene, no quenching was observed. No carboxylation occurred using thiophene as substrate under the optimized reaction conditions.



**Figure S10c.** Stern-Volmer plot developed with data obtained from time-resolved quenching experiments of **TMAH** in presence of  $Cs_2CO_3$  with *N*-heteroarenes and benzo[*b*]thiophene as reference.

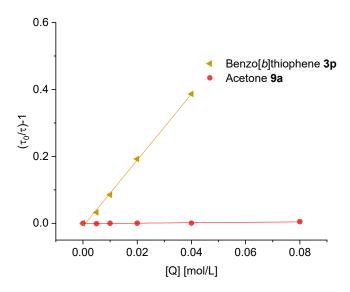
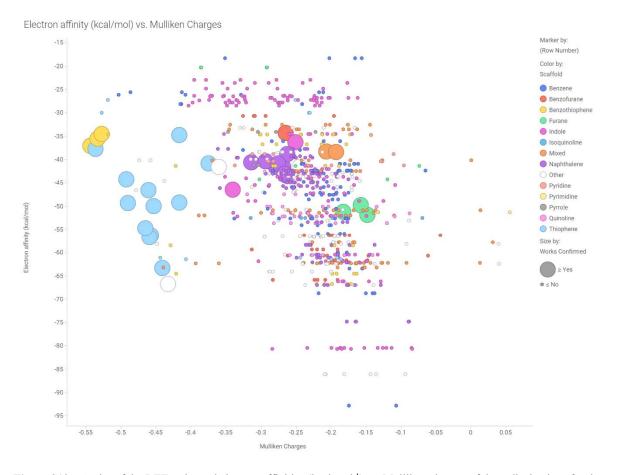


Figure S11d: Stern-Volmer plot developed with data obtained from time-resolved quenching experiments of TMAH in presence of  $Cs_2CO_3$  with acetone and benzo[b]thiophene as reference.

## 6.4 - Computational Analysis

Screening density functional theory (DFT) calculations were performed on substrates and radical anions to derive molecular and atomic properties that could rationalize the reaction outcomes. Geometries were optimized using the B3LYP-D3<sup>42</sup> *a posteriori*-corrected hybrid functional<sup>43</sup> with the LACVP\*\*+ basis set, and final energies and atomic properties were calculated using B3LYP-D3/LACV3P\*\*+ together with the PBF solvation model<sup>44</sup> for DMSO. The calculations were performed within the Schrödinger Small-Molecule Drug Discovery Suite 2019-2 using Jaguar version 10.4.<sup>45</sup> To facilitate convergence to a minimum, any apparent symmetry in the starting geometry was ignored in the optimizations (isymm=0). To facilitate SCF convergence for some radical anions the use of the pseudospectral method was turned off during all calculations (nops=1; *J* and *K* operators constructed from analytic two-electron integrals; no grid used). For each substrate and radical anion, Atomic Fukui indices, Mulliken charges and the spin population were calculated. The electron affinity for each substrate was roughly estimated by the direct DFT energy difference between the radical anion and the substrate and are given in kcal mol<sup>-1</sup>.

As seen in Figures S10-13, there is a strong correlation with reactivity and a positive outcome and calculated atomic descriptors and estimated electron affinities. However, there are also substrates that seem to fall within the acceptable range of estimated electron affinity, atomic charge, spin distribution and nucleophilicity that does not yield the desired products. This could be due to subsequent spontaneous decarboxylation as for 3u (requiring trapping the carboxyl acid as an ester) or the presence of nontolerated functional groups. According to the calculations, the regioselectivity is most strongly correlated to the Mulliken spin population.



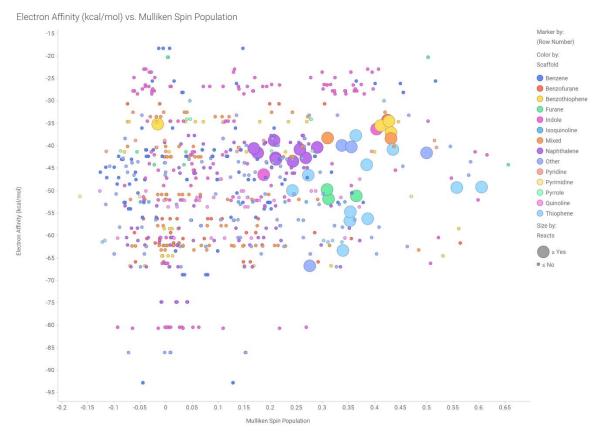
**Figure S12a.** A plot of the DFT estimated electron affinities (kcal mol<sup>-1</sup>) *vs.* Mulliken charges of the radical anions for the carbons reacting with CO<sub>2</sub> and for all CH carbons in the aromatic rings of non-reacting substrates, highlighting that most of the reactive substrates are located within a triangle. Three reacting furanes coloured in green are outliers.



**Figure S12b.** A plot of the DFT estimated electron affinities (kcal mol<sup>-1</sup>) vs. the Fukui f-NN-index (describing nucleophilicity) of the radical anions for the carbons reacting with  $CO_2$  and all CH carbons in the aromatic rings of non-reacting substrates illustrating that most of the reactive substrates have more nucleophilic radical anions.



**Figure S12c.** A plot of the DFT Mulliken charges of the radical anions vs. Fukui f-NN-index of the radical anions for the carbons reacting with CO<sub>2</sub> and all CH carbons in the aromatic rings of non-reacting substrates illustrating that most of the non-reactive carbons are less negatively charged and have lower predicted nucleophilicity.



**Figure S12d.** A zoom in plot of the DFT Mulliken charges of the radical anions *vs.* Fukui f-NN-index of the radical anions for the carbons reacting with CO<sub>2</sub> and all CH carbons in the aromatic rings of non-reacting substrates including only compounds with DFT estimated electron affinities within the values among the substrates that react. Highlighted are substrates with required electron affinities and nucleophilicity but not reacting due to non-compatible functional groups.

## 7 - Miscellaneous

# 7.1 - Synthetic route towards FDCA and DMFDC

Modifying the conditions during reaction work-up allows for the direct transformation of the crude reaction mixture of  $4\mathbf{r}$  to either 2,5-furandicarboxylic acid (FDCA) or dimethyl 2,5-furandicarboxylate (DMFDC). Both are important monomers for the manufacture of polyesters derived from biomass (Scheme S4). The reaction work-up with conc. HCl would cause the hydrolysis of the ester giving rise to the dicarboxylic acid FDCA. In contrast, the addition of MeI after releasing the  $CO_2$  overpressure allows for the formation of dimethyl dicarboxylate DMFDC.

Scheme S4: Synthetic route towards FDCA and DMFDC starting from crude reaction mixture of 4r.

### 7.2 - Carboxylation of biphenyl

Biphenyl acts as a good quencher but the resulting carboxylation product [1,1'-biphenyl]-4-carboxylic acid was only obtained in low yield (6%). Nevertheless, this result shows that also benzene derivatives can be activated towards a C–H carboxylation with our method. The thermodynamic driving force for this transformation with CO<sub>2</sub> however seems to be low.

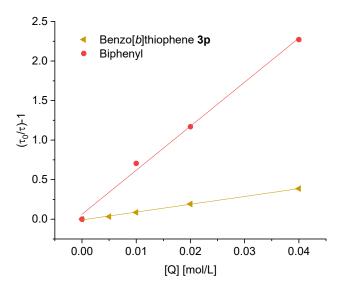


Figure S13. Stern-Volmer plot developed with data obtained from time-resolved quenching experiments of TMAH in presence of Cs<sub>2</sub>CO<sub>3</sub> with biphenyl and benzo[b]thiophene 3p as reference.

Scheme S5. TMAH-catalyzed carboxylation reaction of biphenyl.

<sup>1</sup>**H-NMR** (400 MHz, DMSO- $d_6$ ) δ 12.93 (s, 1H), 8.02 (d, J = 8.4 Hz, 2H), 7.80 (d, J = 8.4 Hz, 2H), 7.76 – 7.70 (m, 2H), 7.54 – 7.47 (m, 2H), 7.45 – 7.40 (m, 1H). **HRMS** (ESI+): calculated m/z for C<sub>13</sub>H<sub>11</sub>O<sub>2</sub> [(M+H)<sup>+</sup>] 199.0754; found 199.0751.

## 7.3 - Unsuccessful Substrates

The following substrates (excerpt of examined scope) were found to be not successful under the reported reaction conditions (Figure S14a-c). Structures marked in red do not quench the excited photocatalyst, structures marked in blue were found to quench the excited photocatalyst and structures in black were not tested regarding luminescence quenching.

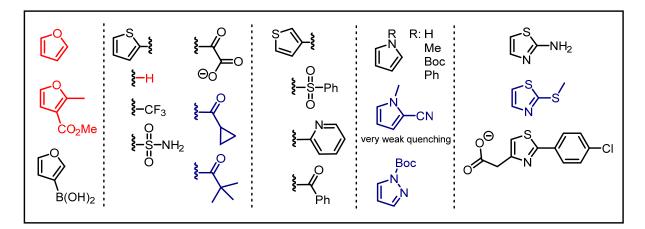


Figure S14a. Non-tolerated examined furans, thiophenes, pyrroles and thiazoles.

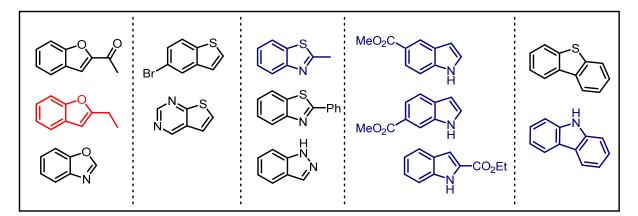


Figure S14b. Non-tolerated examined benzofurans, benzothiophenes, indoles, carbazoles and related heterocycles.

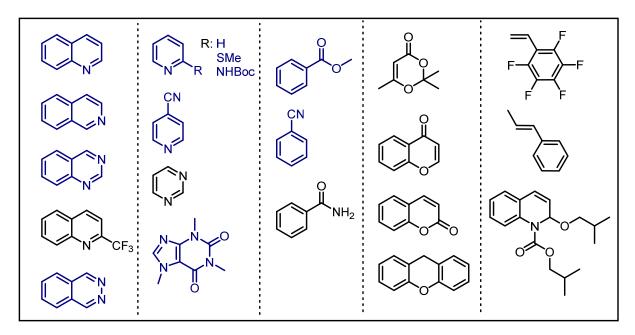


Figure S14c. Non-tolerated quinolines, pyridines, benzenes, benzopyrans, styrenes and related compounds.

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